

Fig. 1. Pseudaletis spolia Riley, \$\forall \text{ holotype.} \text{Fig. 2. Pseudaletis ugandae Riley, \$\partial \text{ holotype.} \text{ fig. 3. Telipna Sheffieldi Beth.-Baker, \$\forall \text{.} \text{ Fig. 4. Telipna rothi Smith, \$\forall \text{.} \text{ Fig. 5. Egumbia ernesti (Karsch), \$\forall \text{.} \text{ Fig. 6. Oxylides gloveri Hawker Smith, \$\forall \text{ holotype.} \text{ Fig. 7. Tumerepedes flava Beth.-Baker, \$\partial \text{ holotype.} \text{ Fig. 8. Aslauga aura H. H. Druce, \$\partial \text{ holotype.} \text{ Fig. 9. Pseudaletis dardanella Riley, \$\partial \text{ holotype.} \text{ All figures are natural size.}

# THE GENERA OF THE AFRICAN LYCAENIDAE (LEPIDOPTERA: RHOPALOCERA)



BY

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1 Colour plate, 348 text-figs.

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TRUSTEES OF
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#### CONTENTS

												Page
Introdu	JCTION											3
									s .			
Conclus	SION									. 1		264
Postscr	IPT (O	N THE	"B	UTTER	FLIES	OF	LIBERIA	"	CLENCH,	1965)		277
Index												300

#### SYNOPSIS

The genera of African Lycaenidae are revised and figures of wing venation and genitalia are included for each genus. Lists of the species included in each genus are given. The major classification of the Lycaenidae occurring in the area is rationalized.

#### INTRODUCTION

SINCE 1901, when Staudinger's Catalog der palaearktischen Lepidopteren was published, the classification of the palaearctic Lycaenidae has undergone profound modifications due to the importance now ascribed to the characters of the male genitalia. In order to establish a natural classification, we accord these characters, which we consider ancestral and of phylogenetic importance, precedence over external characters, which though much easier to appreciate, often present resemblances that are really due to fortuitous coincidence or convergent evolution.

In the very uniform subfamilies Theclinae and Lycaeninae, we have retained most of the genera included by Staudinger, but the omnibus genus *Lycaena*, in which species were arranged in a most arbitrary manner, has been broken up into the following subfamilies: Everinae, Lampidinae, Plebeiinae, Glaucopsychinae, Zizerinae and Lycaenopsinae. Most present day entomologists accept this classification and differ only as to the systematic rank, whether genus or subgenus, to be accorded to certain recently established groups of species.

We thus have a general concept of the palaearctic Lycaenidae which is in accordance with our present knowledge of systematics. The nearctic species have been classified on the same lines, but when we come to the Lycaenidae of the tropical regions, the situation is very different.

The only complete surveys that we have for the Ethiopian Lycaenidae are those of Aurivillius, firstly his *Rhopalocera Aethiopica* (1898), and secondly his account of the African Lycaenidae in Seitz, *Gross-Schmetterlinge der Erde* 13 (1914–25). In his first work Aurivillius based his classification solely on external characters, wing shape

and venation, shape of palpi, antennae, legs, etc. He divided the family into only two subfamilies, the Lipteninae peculiar to the Ethiopian regions, and the Lycaeninae. In his second work he retained his original classification with minor modifications and he intercalated the numerous species described between 1898 and the date of publication of the parts of Seitz's work. However, in the same period, Bethune Baker (1910, 1918, 1922 and 1924), published a number of monographs in which he made great use of genital characters. T. A. Chapman also published a Revision of the sub-family Zizeeriinae (*Trans. ent. Soc. Lond.* 1910: 480 et seq.). Aurivillius (in Seitz, 1914–25) alluded to these monographs but refused to modify the basis of his classification on the plea that classification could not be based on the characters of one sex only. He still included in a very extensive genus, *Cupido* (with typespecies *C. minimus* Fuessly, one of the Everinae), a motley crowd of species belonging to the Lampidinae, Plebeiinae, Zizeriinae, etc.

My disagreement with Aurivillius is not due to a difference of opinion concerning nomenclature and priority of description, it is a fundamental difference in concepts of classification. I think that classification should reflect phylogeny (as far as we can ascertain it), while Aurivillius contended that the best classification is that which enables an entomologist to determine easily and quickly any given specimen by the study of its external appearance.

However great our respect for the considerable achievement of Aurivillius, who was the first to bring some kind of order out of the chaos of the rich Ethiopian Fauna, I do not think that nowadays we can accept his method. I contend that we should attempt to do for the Ethiopian Fauna what has been done for the palaearctic Fauna, i.e. try to construct a natural classification.

Unfortunately the study of the Ethiopian Fauna lags far behind that of the palaearctic and nearctic faunas. Besides the excellent monographs of Bethune Baker and Chapman, which cover only a small part of the Ethiopian Fauna, we have only fragmentary studies consisting of faunistic lists and descriptions of new species. The male genitalia of many genera, especially in the subfamily Lipteninae, have never been studied or, at least, the results of such studies have never been published; up to now no comparison of the different genera has ever been attempted, neither has any attempt been made to group them into natural subfamilies.

It is this lack of a comprehensive outlook that I wish to try to remedy by making use of the knowledge available and adding to it such knowledge as I have acquired by personal observations. In the course of the last 25 years I have dissected and studied the male genitalia of many Ethiopian Lycaenidae, some from specimens sent me for determination, others from specimens kindly lent to me by Museums and private collectors. Altogether, I have examined more than 6,000 specimens belonging to 939 different species, out of a total of 1,263 known species. Previously I had examined some 500 palaearctic, nearctic and Indo-Malayan species; the knowledge thus acquired served me well in assessing relationship. Nevertheless I wish to make it clear that I have no intention to supersede the works of Aurivillius, which will still remain indispensable for the determination of species. My aim is quite different.

To facilitate the task of the reader, I have kept the genera in the same order as that followed by Aurivillius, although I do not always agree with him. I have reduced bibliographical references to old authors to a minimum since, as these can be found in the *Rhopalocera Aethiopica* of Aurivillius, their repetition would have overloaded the text to no purpose. Of modern authors I cite only the more comprehensive works which deal with a genus or an extensive regional fauna.

A few words are necessary about the descriptive text and the figures which make up the body of the present work. With regard to external characters I recapitulate only the more striking, truly generic features, but the condition of the fore tarsus of the male, the segments of which are generally fused, is always mentioned. As far as wing venation is concerned, I consider that a figure is more informative than a long description.

There follows a short description and schematic figure of the male genitalia of the

type-species of the genus, drawn with the aid of a camera lucida.

With very few exceptions the genital organs are drawn in ventral aspect, spread out and, where possible, flattened. Such a drawing differs considerably from the view obtained when the genitalia are examined in situ, and the reader who is not familiar with the dissection of male genitalia may have some difficulty in visualizing them in their natural position. A lateral drawing of the whole genitalia mounted in depth would have many disadvantages, such as lack of clarity and overlapping and foreshortening of some of the organs. Although I include some such figures, I have

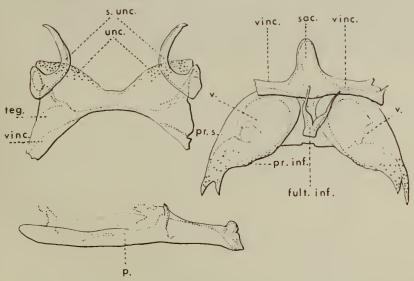


Fig. 1. Anthene definita Butler, 3 genitalia. unc., uncus = cheeks of Bethune Baker; s. unc., subunci = falces of Bethune Baker (these are not the same as the socii which are found in other groups of Lepidoptera); teg., tegumen; vinc., vinculum; sac., saccus; fult. inf., lower fulture = furca of Bethune Baker; v., valves = harpagones of Bethune Baker; pr. s., upper process of the valves; pr. inf., lower process of the valves; p., penis = aedeagus of Bethune Baker.

preferred to give drawings of the organs separately. Although the making of such mounts entails longer and more delicate operations, it is worth while because it gives a better idea of each organ and shows up the specific characters. Where there were imperfections in my preparation, I have reproduced them in my drawings so that my figures are faithful copies and not improvements on the original. To avoid confusion I have not drawn any hairs, but I have shown their points of insertion.

The terminology of the genital parts employed in the present work differs slightly from that which I used in some of my previous publications. In these, I followed Bethune Baker, and employed the term "tegumen" for the whole of the dorsal parts. I think now that it is more reasonable to call the part that corresponds to the tenth tergite "the uncus", and to confine the term "tegumen" to the derivative of the ninth tergite, even though the uncus, often closely fused to the tegumen, appears merely as a thickening on its posterior margin, or only consists of the two small lateral lobes, which Bethune Baker called "cheeks". For the part supporting the penis which is usually fused to the base of the valves, I employ the term "lower fultura", a more general term than "furca", which is a special form of the lower fultura found in the Plebeiinae.

The terminology of the parts, as used in the present work, is indicated in figures of *Anthene definita* Butler (Text-fig. 1) and *Lepidochrysops victoriae* Karsch (Text-fig. 2).

In most cases the description of the genitalia is followed by the comparison of the genitalia of the type-species with those of other species in the genus, in an attempt to establish whether the nominal genus under review is a natural taxonomic unit, or merely an artificial heterogeneous collection of species. But as I have seldom been able to examine all the species included in any genus (see Lists of Species) I have

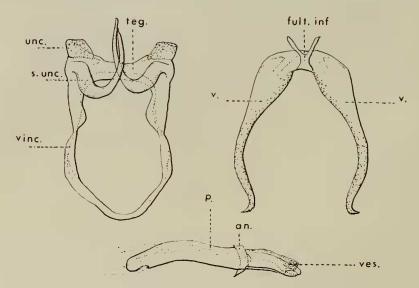


Fig. 2. Lepidochrysops victoriae Karsch, & genitalia, abbreviations as in Text-fig. 1, an., anellus; ves., vesica.

generally refrained from making new subdivisions or, in particular, introducing new generic names, which might add to the existing confusion.

At this point I have added brief indications of what is known of the life histories of the species referred to the genus. This is the only part of the present work which is pure compilation, for I have no personal knowledge of the early stages of any of the species concerned. It shows how strikingly ignorant we are in this respect.

Finally, there is appended to the account of each genus an alphabetical list of all the species, subspecies, varieties etc., referred to it, and their synonyms; those of which I have myself examined the male genitalia are marked with an asterisk, and I have been at pains to give references to descriptions of these organs published elsewhere by other authors. The alphabetical arrangement has been deliberately chosen because, in my opinion, the gaps in our knowledge are still so great as to render the production of a satisfactory natural systematic arrangement impossible. I am fully aware of the imperfections and lacunae of this work. It is not exactly attractive; the employment of a uniform order in the enumeration of characters and the necessarily concise style inevitably tend to produce a dry, monotonous text. My essay is more critical than constructive, and is meant to show that the present classification rests on imperfect bases. I have been content in the end to suggest a few new groupings of species and genera, nothing more, because to build up a sound system of classification, it would be necessary to examine all the species that exist, and I have not been able even to study all those that are known. It must be remembered that if it is comparatively easy to obtain sufficient material for the study of a limited and well known fauna, such as that of Europe, it is practically impossible to examine within a reasonable time all the Ethiopian species, many of which are represented by single specimens distributed in Museums and private collections all over the world. This will be the work of many authors specializing on single genera, or on limited groups of genera at a time.

Incomplete as this work is bound to be, I hope that it may be of some use, if only to incite other entomologists to further studies. I should not have dared to undertake it, had it not been for the friendly encouragement of Mr. N. D. Riley, formerly Keeper of Entomology, in the British Museum (Natural History), whose advice and help has always been available to me, and who made himself responsible for the final revision of the English text and for elucidating various nomenclatural obscurities etc. I also wish to thank the Trustees of the British Museum (Natural History) for their kindness in publishing this work.

I owe sincere thanks also to all the entomologists, both officials, and amateurs, who have sent me material for study, first and foremost Mr. T. H. E. Jackson of Kitale, Kenya, who sent me hundreds of specimens; further Mr. B. D. Barnes of Umtali; Mr. N. H. Bennett, Tring; Monsieur L. Berger, of the Musée Royal de l'Afrique central, Tervuren; Mr. R. H. Carcasson, Coryndon Museum, Nairobi; the late G. C. Clark, Port Elizabeth; Mr. H. Cookson, Umtali; Mr. C. G. C. Dickson, Cape Town; Dr. M. Fontaine, Brussels; Dr. W. Forster, Zoologische Sammlung des Bayerischen Staates, Munich; Dr. H. J. Hannemann, Berlin; Father Th. Massen, missionary in Ghana; Mr. K. M. Pennington, Balgowan, Natal; Dr. E. C. G. Pinhey, Bulawayo; Dr. Patrick Roche, London; Monsieur P. Rougeot,

Paris; Dr. van Someren, Ngong, Kenya; Dr. van Son, Pretoria; and Mr. G. E. Tite, Tring.

## DIAGNOSES OF THE GENERA, WITH LISTS OF SPECIES

## Genus ALAENA Boisduval

Alaena Boisduval, 1847, Voyage Delegorgue 2:591; Aurivillius, 1898:254; 1919-25:298; Desmond Murray, 1923:47; Pinhey, 1949:96, pl. 15, figs 1-3; Swanepoel, 1953:182, 184, pl. 8, figs 38-41. Type-species: Alaena amazoula Boisduval, by monotypy.

Male fore leg stout, very pilose, especially on the tibia; tarsus short, not distinctly segmented, strongly spinose ventrally, terminal claws absent;  $\varphi$  fore leg fully developed, less hairy than that of  $\Im$ .

Wing venation (Text-fig. 229). There is a short but distinct precostal vein at the base of the hind wing.

Male genitalia (Text-fig. 3). Uncus composed of two rounded lobes, densely covered with short hairs and broadly fused to a broad tegumen; subunci robust, curved near the base and ending in a hook; vinculum narrow, incomplete ventrally; the oblong valves are directly fused to the extremities of the tergite and are partially joined together on the lower edge, whilst their upper edges are bridged by a narrow membranous strip, thus forming a sheath for the strong, subcylindrical penis, which is somewhat broadened just before its truncated apex; there are a few short hairs on the distal portion of the valves.

The male genitalia of other species, as far as they are known, closely resemble those of A. amazoula; they differ only slightly in the shape of the valves. The genus seems homogeneous, both in structure and in general appearance. The species of Alaena are somewhat small with oblong, rounded wings, yellowish or black in colour, spotted with white. They remind one a little of the Acraeinae, in which family they were originally included.

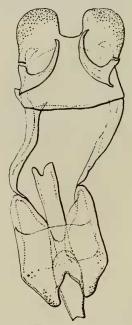


Fig. 3. Alaena amazoula Boisduval, & genitalia.

## LIST OF SPECIES OF Alaena

Alaena amazoula amazoula Boisduval, 1847. Fig. Trimen, 1862.

Alaena amazoula congoana Aurivillius, 1914.

Alaena amazoula nyasana Hawker Smith, 1933, Stylops 2: 1.

Aleana aurantiaca see interposita.

Alaena bicolora Bethune Baker, 1924, Ann. Mag. nat. Hist. (9) 14: 130.

Alaena caissa Rebel & Rogenhofer, 1894. Fig. Aurivillius in Seitz, 1914-25.

Alaena caissa kagera Talbot, 1935, Entomologists' mon. Mag. 71: 69, fig.

Alaena ferrulineata Hawker Smith, 1933, Stylops 2: 2.

Alaena interposita interposita Butler, 1883.

Alaena interposita hauttecoeuri Oberthur, 1888.

aurantiaca Butler, 1895.

Alaena johanna E. M. Sharpe, 1890. Fig. Sharpe, 1894.

Alaena johanna tsavoa Jackson, 1965, Ann. Mag. nat. Hist. (13) 8:527, fig.

Alaena kiellandi Carcasson, 1965, J. E. Afr. n. H. Soc. 25: 132, figs.

Alaena lamborni Gifford, 1965, Butt. Malawi: 41, figs.

Alaena maculata maculata Hawker Smith, 1933, Stylops 2: 3.

Alaena maculata ochrea Hawker Smith, 1933, Stylops 2: 3.

Alaena madibirensis Wichgraf, 1921, Int. ent. Z. 14: 195.

Alaena major, see nyassae.

\*Alaena margaritacea Eltringham, 1929, Trans. ent. Soc. Lond. 77: 492. Fig. Murray, 1935.

Alaena mulsa, see picata.

Alaena ngonga Jackson, 1965, l.c.: 521, fig.

\*Alaena nyassae nyassae Hewitson, 1877. Fig. Trimen, 1894 = major Oberthur, 1894.

Alaena nyassae marmorata Hawker Smith, 1933, Stylops 2:1.

Alaena nyassae ab. ochracea Butler, 1893.

\*Alaena oberthuri Aurivillius, 1898. Fig. Aurivillius in Seitz, 1914-25.

\*Alaena picata picata E. M. Sharpe, 1896. Fig. Aurivillius in Seitz, 1914–25. mulsa Thieme, 1904; rollei Suffert, 1904.

Alaena picata interrupta Hawker Smith, 1933, Stylops 2: 2.

Alaena picata connectens Talbot, 1935, Entomologist's mon. Mag. 71:69.

Alaena reticulata Butler, 1896. Fig. Butler, 1897.

Alaena rollei see picata.

\*Alaena subrubra Bethune Baker, 1915, Ann. Mag. nat. Hist. (8) 16: 186.

\*Alaena unimaculata Hawker Smith, 1926, Revue Zool. Bot. afr. 14: 237.

## Genus TELIPNA Aurivillius

Telipna Aurivillius, 1895, Ent. Tidskr. 16: 198; 1898: 256; 1914–25: 300. Type-species: Liptena acraea Westwood, 1851; 1959: I.C.Z.N. Opinion 566.

This type-designation, made under the plenary powers of the International Commission for Zoological Nomenclature, is justified by the facts, which are clearly

set out in the *Opinion* quoted above and in the application submitted to the Commission by Francis Hemming and myself. The strict application of the Rules would have resulted in great confusion in the generic names of some 150 species of Lycaenidae, would have upset usages which had been customary since 1898, and served no useful purpose.

Frons and palpi bearing closely adpressed hair-scales. Eyes smooth, palpi short, third segment much reduced, button-shaped. Thorax laterally compressed, abdomen rather long. Legs black, white-ringed; of fore tarsi unsegmented, without terminal claws.

Wing venation (Text-fig. 230). A short precostal vein is present at the base of the hind wings. Male genitalia (Text-fig. 4). It is difficult to give a clear description of the 3 genitalia of T. acraea; only by carrying out an actual dissection can a satisfactory idea be obtained. Bethune Baker's description (1914: 319, pl. 59, fig. 12) is very brief and his photograph showing the organs in profile, in situ, is rather confusing. I have chosen to figure the genitalia with the different parts separated, in ventral aspect, and spread out as much as possible.

Uncus bilobed, without clearly articulated subunci, but with lateral expansions fused to the very large tegumen. Vinculum broad; on the tergite-sternite suture there are two supplementary processes of irregular outline enclosing the penis and resembling a secondary pair of reduced valves; the true valves broadly fused basally to the vinculum, the upper process ending in a pair of rounded lobes, the much shorter lower process having a similar rounded end. Penis uniformly tapering, ending in a truncated cone. Uncus and distal half of valves hairy.

The male genitalia of all the species of *Telipna* examined are very uniform, except for *T. carnuta* which really should be excluded from the genus. The venation, too, of this species is slightly different; on the fore wing vein 7 arises slightly before the upper angle of the cell; on the hind wing vein 7 is stalked with vein 6, separating from it very near its origin.

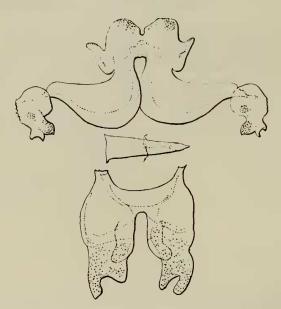


Fig. 4. Telipna acraea semirufa Smith & Kirby, & genitalia.

The male genitalia of T. carnuta (Text-fig. 5), as Bethune Baker (1914: 319) has already pointed out, are far removed from those of the true Telipnas. Tegumen and subunci similar to these parts in species of *Alaena*; valves directly united to the tergal portion of the vinculum, of which the sternal part is lacking; the valves, to use Bethune Baker's expression, are shaped like a ham in which the knuckle end is deeply divided at its extremity; they are joined together close to their bases by a broad chitinous band at their upper edges, and by a narrower band at their lower edges, the latter band bearing also a long tapering process which is directed caudad. Penis robust, slightly curved, tapering uniformly to the obliquely truncate apex; vesica armed with strong spines.

The early stages of T. consanguinea Rebel and T. sanguinea depuncta Talbot have been noted by T. H. E. Jackson (1937, Trans. R. ent. Soc. Lond. 86: 205). The larvae feed on lichens and mosses on the bark of trees.

# LIST OF SPECIES OF Telipna

- \*Telipna acraea acraea (Doubleday & Hewitson), 1852.
- Telipna acraea nigra Suffert, 1904, Dt. ent. Z., Iris 17: 42.
- \*Telipna acraeoides acraeoides (Smith & Kirby), 1890. Fig. Aurivillius in Seitz, 1914–25. Fig. Smith & Kirby, 1887, as sanguinea Plötz, figs 1, 2 only. Fig. Hewitson, 1866, Exot. Lep. III, fig. 12, as acraea Doubleday & Hewitson, nec Westwood, 1852.
- Telipna acraeoides laplumei Devos, 1919, Revue zool. afr. 6:62.
- Telipna actinotina Lathy, 1903, Trans. ent. Soc. Lond. 1903: 194, fig. \*Telipna angustifascia angustifascia Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1: 77, fig.
- \*Telipna angustifascia neavei Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17:316.
  - Telipna anneckei, see sanguinea.

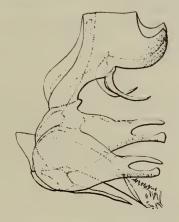


Fig. 5. Telipna carnuta Hewitson, of genitalia.

Telipna atrinervis Hulstaert, 1924, Revue zool. afr. 12: 113.

\*Telipna aurivillii Rebel, 1914, Annln naturh. Mus. Wien 28: 262, fig.

\*Telipna bimacula bimacula (Plötz), 1880, Fig. Aurivillius in Seitz, 1914–25. fervida Smith & Kirby, 1890.

Telipna bimacula echo (Smith & Kirby), 1890.

\*Telipna bimacula semirufa (Smith & Kirby), 1889.

\*Telipna bimacula albofasciata Aurivillius, 1910.

Telipna bimacula f. nigrita Talbot, 1935, Entomologist's mon. Mag. 71: 70.

\*Telipna carnuta carnuta (Hewitson), 1873. Fig. Smith & Kirby, 1893. Telipna carnuta parva (Kirby), 1887. Fig. Smith & Kirby, 1888.

Telipna citrimacula Schultze, 1916, Arch. Naturgesch. 81 (A) Heft 12: 141.

\*Telipna consanguinea Rebel, 1914, Annln Naturh. Mus. Wien 28: 262, fig. Telipna consanguinea ab. extincta Schultze, 1923, Ergeb. 2te D. Zent. Afr. Exp. 1910–11, 1: 1150, fig.

\*Telipna erica erica Suffert, 1904, Dt. ent. Z. Iris 17: 41. Fig. Aurivillius in Seitz,

1914-25.

\*Telipna erica ugandae Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 388. Telipna exsuperia, see hollandi.

Telipna fervida, see bimacula.

\*Telipna hollandi Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1: 80, fig. exsuperia Hulstaert, 1924.

Telipna ja Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 388.

Telipna kamitugensis Dufrane, 1945, Bull. Annls Soc. R. ent. Belg. 81: 114.

\*Telipna katangae Stempffer, 1961: 9, fig.

Telipna mariae Dufrane, 1945, Bull. Annls. Soc. R. ent. Belg. 81: 112. Telipna medjensis Holland, 1920, Bull. Am. Mus. nat. Hist. 43: 214, fig.

\*Telipna nyanza Neave, 1904, Novit. zool. 11: 335, fig.

\*Telipna plagiata Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1: 79, fig. Telipna rothi (Smith), 1898.

Telipna rothioides Holland, 1920, Bull. Am. Mus. nat. Hist. 43: 214, fig.

Telipna rufilla (Smith), 1901.

Telipna ruspinoides Schultze, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910-11, 1:1151, fig.

\*Telipna sanguinea sanguinea (Plötz), 1880. Fig. Smith & Kirby, 1887 (figs 3, 4 only)

anneckei Dewitz, 1886.

Telipna sanguinea bistrigatus Aurivillius, 1925, Ark. Zool. 17A, 32:7.

\*Telipna sanguinea depuncta Talbot, 1937, Trans. R. ent. Soc. Lond. 86: 59. Telipna sheffieldi Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 387. Telipna subhyalina Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1: 78, fig. Telipna sulpitia Hulstaert, 1924, Revue zool. afr. 12: 114.

\*Telipna transverstigma H. H. Druce, 1910, Proc. zool. Soc. Lond. 1910: 356, fig. Telipna venanigra Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 387. Telipna villiersi Stempffer, 1965, Bull. Inst. fr. Afr. noire 27: 1450, figs.

# Genus COOKSONIA H. H. Druce

Cooksonia H. H. Druce, 1905, Trans. ent. Soc. Lond. 1905: 256; Aurivillius, 1914-1925: 302. Type-species: Cooksonia trimeni H. H. Druce, 1905, by original designation.

Sheffieldia H. H. Druce, 1912, Entomologist's mon. Mag. 48: 128; Talbot, 1935, Entomologist's mon. Mag. 71: 202, synonymy. Type-species: Sheffieldia neavei H. H. Druce, by monotypy.

Eyes smooth; palpi of medium length, erect, clothed with adpressed black scales, third segment very short; antennae slender, half as long as costa; club well differentiated, flattened, spatulate.

Wing venation (Text-fig. 231).

The only known specimen of C. trimeni is a Q, so I have been unable to examine the  $\partial$  genitalia of the type-species; on the other hand, I have examined the  $\partial$  genitalia of C. neavei H. Druce and C. aliciae Talbot.

In C. neavei the  $\mathcal{J}$  fore legs are short, almost glabrous, tibia a trifle shorter than the femur, tarsus unsegmented and bearing fine spines below. The venation (Text-fig. 232) of C. neavei differs slightly from that of C. trimeni inasmuch as veins 5 and 6 of the fore wing have a short common stem which arises from the upper angle of the cell, but this difference is not constant, for I have seen in the British Museum a  $\mathcal{J}$  specimen of C. neavei in which the veins arise as in C. trimeni. The synonymy given by Talbot, viz: Cooksonia = Sheffieldia, is undeniable.

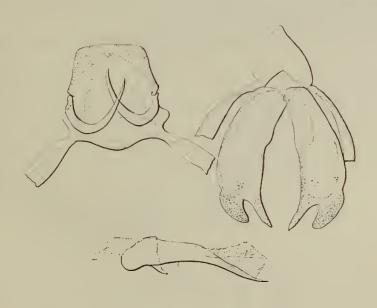


Fig. 6. Cooksonia neavei (Druce), & genitalia.

Male genitalia of C. neavei (Text-fig. 6): uncus crescentic with an almost straight anterior margin; subunci long, curved, tapering regularly; tegumen triangular, separated from the uncus by a membrane which is translucent under the microscope; vinculum fairly wide, saccus triangular, turned towards the apex of the abdomen and not towards the eighth segment; valves oblong, the two processes separate at the apex, the apex of the upper process rounded, that of the lower one more pointed; penis elongate, ending in a sharp, slightly curved point; uncus and apices of valves densely pilose.

In C. aliciae the  $\Im$  fore legs are like those of C. neavei, and the male genitalia (Text-fig. 7) almost identical with those of C. neavei, except that the lateral angles of the uncus are more rounded.

The male genitalia of these two species are of the commonplace pattern, totally different from that found in species of *Telipna* and *Pentila*, in which the genitalia are highly specialized. There is, therefore, no good reason for leaving the genus *Cooksonia* in the company of the above two genera, the only character it has in common with them being the presence of a precostal vein in the hind wing, a character which, incidentally, was not mentioned in the description of the genus but which nevertheless is present in the three known species.

The chrysalis of *C. aliciae* has been described and figured by Talbot (1935: 204, pl. V, fig. 3).

## LIST OF SPECIES OF Cooksonia

- \*Cooksonia aliciae Talbot, 1935, Entomologist's mon. Mag. 71: 203, fig.
- \*Cooksonia neavei (H. H. Druce), 1912.

Cooksonia trimeni trimeni H. H. Druce, 1905.

Cooksonia trimeni terpsichore Talbot, 1935, Entomologist's mon. Mag. 71: 204.

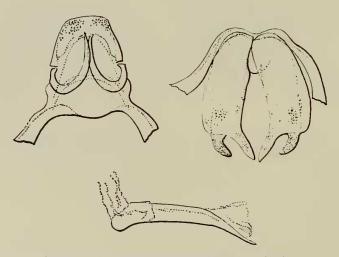


Fig. 7. Cooksonia aliciae Talbot, of genitalia.

## Genus PENTILA Westwood

Pentila Westwood, 1851, Genera Diurnal Lepidoptera, pl. 76; Aurivillius, 1898: 258, 1914: 303; Murray, 1935: 52; Swanepoel, 1953: 191. Type-species: Tingra tropicalis Boisduval, 1847; 1935: I.C.Z.N. Opinion 566.

Head small, vertex bearing adpressed hairs; eyes large and naked; palpi very small and very short, underside of 1st and 2nd segments bearing long scales, last segment reduced, button-shaped; antennae short, fairly robust, with a very distinct club which is an elongated oval in

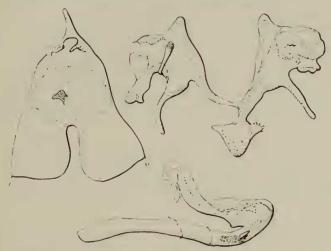


FIG. 8. Pentila tropicalis (Boisduval), & genitalia.

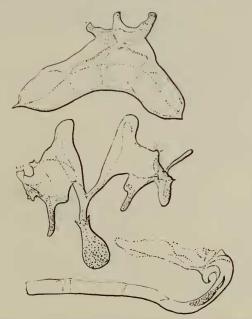


Fig. 9. Pentila tachyroides Dewitz, & genitalia.

shape; thorax short and slender; abdomen long, swollen in its apical portion; legs short, robust, bearing a few scales but no hairs, tarsi spinose ventrally;  $\eth$  fore leg with the tibia finely spinose ventrally, tarsus short, unsegmented and ventrally spinose.

Wing venation (Text-fig. 233). The cell is very long in both wings; hind wing with a short precostal vein at the base.

Male genitalia (Text-figs 8, 9). The genitalia of the 3 are asymmetric and highly specialized, but very constant in form throughout the genus. In order to appreciate their configuration thoroughly it is necessary to separate the dorsal and ventral portions and to arrange the parts separately flat on a slide; a preparation showing the parts in situ, in profile, is only confusing and of no use for the recognition of specific characters. Uncus trifurcate, the median process much the longest, the two lateral lobes asymmetric, curved inwards; tegumen very large, hood-shaped, also slightly asymmetric; gnathos strongly chitinized and trumpet-shaped in lateral aspect; sternite prolonged towards the eight segment by two rounded expansions and ending in an asymmetric feebly chitinized portion towards the apex of the abdomen; there are no true articulated valves but five asymmetric expansions of the vinculum, the shapes of which provide excellent specific characters; penis very long and cylindrical, slightly incurved towards the apex; the fully eversible vesica was found everted in all specimens examined, producing the appearance of a flail.

Female genitalia (Text-fig. 10). The \$\pi\$ genitalia are also very specialized. The anal papillae are covered with fine silky hair and are devoid of posterior apophyses. The wide ostium bursae opens on the seventh sternite and is easily visible to the naked eye, which enables the sex to be recognized with ease. The ductus bursae is at first strongly chitinized, and the sclerotized portion, which makes a swelling on the underside of the abdomen, takes an even curve that seems to correspond with the curve of the terminal part of the penis; the membranous portion of the duct is folded back at an acute angle towards the apex of the abdomen. In this way one can understand the peculiar situation imposed on the vesica at the time of copulation, a situation which is retained after copulation and is found on dissecting the males.

I have been able to examine the male genitalia of all known species of *Pentila* except *P. umbra* Holland. As indicated above, they are all of the same general type as *tropicalis*, yet they provide good specific characters. The genus, though numerous in species, is perfectly homogeneous.

Nothing is known of the early stages of any species, which is surprising, for many of the species are abundant in their habitats and they are widely distributed geographically.

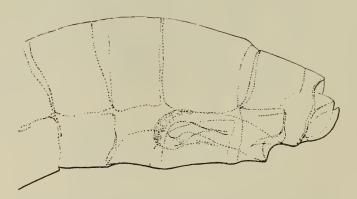


Fig. 10. Pentila tachyroides f. roidesta Suffert, 2 genitalia.

## LIST OF SPECIES OF Pentila

Pentila abraxas abraxas (Doubleday & Hewitson), 1852. Fig. genitalia, Stempffer & Bennett, 1961: 1181.

tripunctata Aurivillius, 1895.

Pentila abraxas ab. affixa Schulze, see abraxas pardalena.

\*Pentila abraxas maculata (Kirby), 1887. Fig. Smith & Kirby, 1888. Fig. genitalia, Stempffer & Bennett, 1961: 1182.

hedwiga Suffert, 1904; telesippe and elpinice Grünberg, 1910.

\*Pentila abraxas pardalena H. H. Druce, 1910. Fig. genitalia, Stempffer & Bennett, 1961: 1185.

abraxas ab. affixa Schultze, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910-11,

1:1157.

- \*Pentila abraxas phidia Hewitson, 1874. Fig. Smith & Kirby, 1893. Fig. genitalia, Stempffer & Bennett, 1961: 1184.

  nunu (Karsch), 1893.
- \*Pentila abraxas subochracea Hawker Smith, 1933, Stylops 2: 4. Fig. and fig. genitalia, Stempffer & Bennett, 1961: 1186.
- \*Pentila alba Dewitz, 1886. Fig. genitalia, Stempffer & Bennett, 1961: 1202.
- \*Pentila amenaida Hewitson, 1873. Fig. genitalia, Stempsfer & Bennett, 1961:1130.
- \*Pentila amenaidoides (Holland), 1892. Fig. and fig. genitalia, Stempffer & Bennett, 1961: 1138.
- \*Pentila auga auga Karsch, 1895. Fig. H. H. Druce, 1910, Ill. African Lycaenidae. Fig. genitalia, Stempffer & Bennett, 1961: 1198.

Pentila auga congoensis Joicey & Talbot, see cloetensi catauga.

Pentila bertha (Smith & Kirby), see nero.

- \*Pentila bitje H. H. Druce, 1910. Fig. genitalia, Stempffer & Bennett, 1961: 1201.
- \*Pentila camerunica Stempffer & Bennett, 1961: 1196, fig. and fig. genitalia.
- \*Pentila carcassoni Stempffer & Bennett, 1961: 1168, fig. and fig. genitalia.
- \*Pentila christina Suffert, 1904. Fig. H. H. Druce, 1910, Ill. African Lycaenidae, pl. 2. Fig. genitalia, Stempffer & Bennett, 1961: 1195.
- \*Pentila cloetensi cloetensi Aurivillius, 1897. Fig. Holland, 1920, Bull. Am. Mus. nat. Hist. 43. Fig. genitalia, Stempffer & Bennett, 1961: 1188. elfrieda Suffert, 1904.
- \*Pentila cloetensis albida Hawker Smith, 1933, Stylops 2:4. Fig. and fig. genitalia, Stempffer & Bennett, 1961:1192.
- \*Pentila cloetensis aspasia Grünberg, 1910. Fig. and fig. genitalia, Stempffer & Bennett, 1961.
- \*Pentila cloetensi f. elfriedana Strand, 1918, Int. ent. Z. 12: 101.
- Pentila cloetensi f. latefascia Dufrane, 1953, Bull. Annls Soc. R. ent. Belg. 89:47.
- \*Pentila cloetensi catauga Rebel, 1914, Annln naturh. Mus. Wien 28: 263, pl. 22. Fig. genitalia, Stempffer & Bennett, 1961: 1189. auga congoensis Joicey & Talbot, 1921.

- \*Pentila cloetensi lucayensis Schultze, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910–11, 1:1158. Fig. and fig. genitalia, Stempffer & Bennett, 1961:1193.
- \*Pentila cloetensi uelensis Stempffer & Bennett, 1961:1190, fig. and fig. genitalia.
- \*Pentila cloetensi condamini Stempffer, 1963, Bull. Inst. fr. Afr. noire 25: 954, fig. and fig. genitalia.

Pentila elfrieda Suffert, see cloetensi.

Pentila elpinice Grünberg, see abraxas maculata.

- \*Pentila fallax Bethune Baker, 1915, Ann. Mag. nat. Hist. (8) 16: 187. Fig. and fig. genitalia, Stempffer & Bennett, 1961: 1148.
- \*Pentila fidonioides Schultze, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910-11, 1:1155. Fig. and fig. genitalia, Stempffer & Bennett, 1961:1133.
- \*Pentila glagoessa (Holland), 1893. Fig. genitalia, Stempffer & Bennett, 1961: 1203.

Pentila hedwiga Suffert, see abraxas maculata.

\*Pentila hewitsoni hewitsoni (Smith & Kirby), 1887. Fig. Aurivillius in Seitz, 1914–25. Fig. genitalia, Stempffer & Bennett, 1961: 1199.

Pentila hewitsoni f. leura (Kirby), 1890. Fig. Smith & Kirby, 1891.

Pentila hewitsoni limbata (Holland), 1893.

- \*Pentila inconspicua H. H. Druce, 1910. Fig. and fig. genitalia, Stempffer & Bennett, 1961: 1162.
- \*Pentila landbecki Stempffer & Bennett, 1961: 1165, fig. and fig. genitalia.

Pentila lavinia (Kirby), see torrida.
Pentila lunaris (Weymer), see preussi.

\*Pentila mesia Hulstaert, 1924, Revue zool. afr. 12:115. Fig. and fig. genitalia, Stempffer & Bennett, 1961:1136.

Pentila mylothrina Butler, see tachyroides.

Pentila nero (Smith & Kirby), 1894. Fig. genitalia, Stempffer & Bennett, 1961: 1132.

bertha (Smith & Kirby), 1894.

- \*Pentila nigeriana Stempffer & Bennett, 1961: 1158, fig. and fig. genitalia. Pentila nunu (Karsch), see abraxas phidia.
- \*Pentila nyassana nyassana Aurivillius, 1898. Fig. genitalia, Stempffer & Bennett, 1961: 1113.
- \*Pentila nyassana alberta Hulstaert, 1924, Revue zool. afr. 12: 116. Fig. and fig. genitalia, Stempffer & Bennett, 1961: 1123.

Pentila nyassana amenaidena Strand, 1911.

- \*Pentila nyassana benguellana Stempffer & Bennett, 1961: 1116, fig. and fig. genitalia.
- \*Pentila nyassana clarensis Neave, 1903. Fig. Holland, 1920, Bull. Am. Mus. nat. Hist. 43. Fig. genitalia, Stempffer & Bennett, 1961: 1120.

Pentila nyassana dama Suffert, 1904.

\*Pentila nyassana elisabetha Hulstaert, 1924, Revue zool. afr. 12:116. Fig. and fig. genitalia, Stempffer & Bennett, 1961:1121.

Pentila nyassana f. nigribasis Hulstaert, 1924, Revue zool. afr. 12: 116.

\*Pentila nyassana leopardina Schultze, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910–11, 1: 1154, fig. Fig. genitalia, Stempffer & Bennett, 1961: 1126.

\*Pentila nyassana multiplagata Bethune, Baker 1908. Fig. and fig. genitalia, Stempffer & Bennett, 1961: 1125.

\*Pentila nyassana obsoleta Hawker Smith, 1933, Stylops 2: 3. Fig. Stevenson, 1940, J. ent. Soc. sth. Afr. 3.

Pentila nyassana f. cataractae Stevenson, 1940, l.c.: 101, fig.

\*Pentila nyassana pauli Staudinger, 1888. Fig. Aurivillius, 1898. Fig. genitalia, Stempffer & Bennett, 1961: 1127.

\*Pentila nyassana f. radiata Lathy, 1903.

Pentila nyassana f. multipunctata Lathy, 1903.

\*Pentila nyassana ras Talbot, 1935, Entomologist's mon. Mag. 71:70. Fig. genitalia, Stempffer & Bennett, 1961:1124.

\*Pentila occidentalium occidentalium Aurivillius, 1898. Fig. genitalia, Stempffer & Bennett, 1961: 1156.

Pentila occidentalium f. congoana, Strand, see occidentalium f. immaculata Suffert.

\*Pentila occidentalium gabunica Stempffer & Bennett, 1961:1157, fig. and fig. genitalia.

Pentila occidentalium f. immaculata Suffert, 1904.

occidentalium f. congoana Strand, 1918.

Pentila parapetreia ab. derema Strand, see rogersi parapetreia

Pentila paucipuncta (Kirby), see preussi.

\*Pentila petreia Hewitson, 1874. Fig. Smith & Kirby, 1889. Fig. genitalia, Stempffer & Bennett, 1961: 1170. tripunctata (H. H. Druce), 1888.

\*Pentila petreoides Bethune Baker, 1915, Ann. Mag. nat. Hist. (8) 16: 187. Fig. and fig. genitalia, Stempffer & Bennett, 1961: 1146.

\*Pentila picena picena Hewitson, 1874. Fig. Smith & Kirby, 1893. Fig. genitalia, Stempffer & Bennett, 1961: 1177.

\*Pentila picena catori Bethune Baker, 1906. Fig. and fig. genitalia, Stempffer & Bennett, 1961: 1179.

\*Pentila picena cydaria (Smith), 1898. Fig. genitalia, Stempffer & Bennett, 1961: 1178.

\*Pentila preussi preussi Staudinger, 1888. Fig. Smith & Kirby, 1891. Fig. genitalia, Stempffer & Bennett, 1961: 1171.

paucipuncta (Kirby), 1890, and lunaris Weymer, 1892.

\*Pentila preussi fayei Stempffer, 1963, Bull. Inst. fr. Afr. noire 25: 957, fig.

\*Pentila pseudorotha Stempffer & Bennett, 1961: 1142, fig. and fig. genitalia.

\*Pentila rogersi rogersi (H. H. Druce), 1907. Fig. genitalia, Stempffer, 1953, Annls Mus. R. Congo belge 27: 7.

\*Pentila rogersi parapetreia Rebel, 1908. Fig. genitalia, Stempffer & Bennett, 1961: 1175.

parapetreia ab. derema Strand, 1911.

- \*Pentila rotha rotha Hewitson, 1873. Fig. genitalia, Stempffer & Bennett, 1961:1140.
- \*Pentila rotha marianna Suffert, 1904. Fig. H. H. Druce, 1910, Ill. African Lycaenidae pl. 1. Fig. genitalia, Stempffer & Bennett, 1961: 1142.

\*Pentila subfuscata Hawker Smith, 1933, Stylops 2: 3. Fig. and fig. genitalia, Stempffer & Bennett, 1961: 1172.

\*Pentila tachyroides Dewitz, 1799. Fig. genitalia, Stempffer & Bennett, 1961: 1176.

mylothrina Butler, 1888.

\*Pentila tachyroides f. roidesta Suffert, 1904.

Pentila telesippe Grünberg, see abraxas maculata.

\*Pentila torrida (Kirby), 1887. Fig. Smith & Kirby, 1887. Fig. genitalia, Stempffer & Bennett, 1961: 1204.

lavinia (Kirby), 1890.

Pentila tripunctata Aurivillius, see abraxas.

Pentila tripunctata H. H. Druce, see petreia.

\*Pentila tropicalis tropicalis (Boisduval), 1847. Fig. Hewitson, 1866. Fig. genitalia, Stempffer & Bennett, 1961: 1103.

\*Pentila tropicalis chyulu van Someren, 1939, Jl E. Africa Uganda nat. Hist. Soc. 14: 145. Fig. genitalia, Stempffer & Bennett, 1961: 1110.

\*Pentila tropicalis mombasae (Smith & Kirby), 1889. Fig. genitalia, Stempffer & Bennett, 1961: 1108.

\*Pentila tropicalis f. lasti (Smith & Kirby), 1889. Fig. genitalia, Stempffer & Bennett, 1961: 1109.

Pentila tropicalis f. sigiensis Strand, 1910.

\*Pentila tropicalis swynnertoni Stevenson, 1940 J. ent. Soc. sth. Afr. 3: 101, fig.

\*Pentila umangiana umangiana Aurivillius, 1898. Fig. and fig. genitalia, Stempffer & Bennett, 1961: 1150.

\*Pentila umangiana connectens Hulstaert, 1924, Revue zool. afr. 12:115. Fig. and fig. genitalia, Stempffer & Bennett, 1961:1154.

\*Pentila umangiona fontainei Stempffer & Bennett, 1961: 1153, fig. and fig. genitalia.

\*Pentila umangiana prodita Schultze, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910–11, 1:1156. Fig. and fig. genitalia, Stempffer & Bennett, 1961:1152. Pentila umbra Holland, 1892. Fig. Aurivillius in Seitz, 1914–25.

**Pentila yaunda** Karsch, 1895. Fig. H. H. Druce, 1910, *Ill. African Lycaenidae*, (probably only a synonym of *abraxas maculata* Kirby).

## Genus LIPTENARA Bethune Baker

Liptenara Bethune Baker, 1915, Ann. Mag. nat. Hist. (8) 16: 186. Type-species: Liptenara batesi Bethune Baker, 1915, by original designation.

Eyes palpi and legs similar to those of Pentila.

Wing venation (Text-fig. 234). Only differs from that of *Pentila* in a single detail, namely vein 11 arises somewhat nearer to the base of the wing.

Male genitalia (Text-fig. 11). Quite similar to those of Pentila.

I do not see any important character on which to separate *Liptenara* from *Pentila*. The species are easy to recognize on account of the large white subapical mark, but in my opinion they scarcely merit generic status.

# LIST OF SPECIES OF Liptenara

\*Liptenara batesi Bethune Baker, 1915, Ann. Mag. nat. Hist. (8) 16: 187. Fig. genitalia, Stempffer & Bennett, 1961: 1205.

\*Liptenara hiendlmayri (Dewitz), 1886. Fig. genitalia, Stempffer & Bennett,

1961:1208.

\*Liptenara schoutedeni (Hawker Smith), 1926, Revue zool. afr. 14:238. Fig. genitalia, Stempffer & Bennett, 1961: 1207.

# Genus ORNIPHOLIDOTOS Bethune Baker

Ornipholidotos Bethune Baker, 1914, Trans. ent. Soc. Lond. 1914; 319. Type-species: Pentila kirbyi Aurivillius 1895.

Pentila Westwood (partim); Aurivillius 1898: 263; 1914: 310.

In the original description of this genus, Bethune Baker specified *Pentila muhata* Dewitz, 1886, as its type-species, and figured (l.c. pl. 58, fig. 10) the male genitalia of what he took to be this species.

Through the kindness of Dr. Hannemann of the Berlin Museum, I obtained on loan the two specimens which served Dewitz as the "types" of his original description.

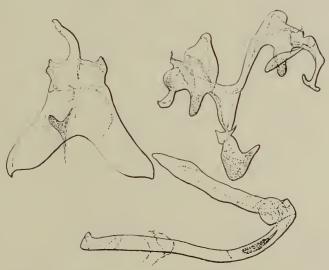


Fig. 11. Liptenara batesi Bethune Baker, & genitalia.

One of them does in fact have a label referring to this description. Examination of the fore tarsi shows that one of these is a male, the other a female but both lack the extremity of the abdomen and, besides, are rather discoloured, probably on account of age. Among the hundreds of specimens of *Ornipholidotos* which I have had the opportunity of examining, collected more or less recently, I have found none which can undoubtedly be regarded as exactly matching Dewitz's "type" specimens of *muhata*. In the absence of genitalia for comparison and in view of the inconclusive nature of any comparison based on external characters in this genus, it is, therefore, impossible at present to identify Dewitz's species, which must remain a *species dubium*.

On the other hand the male genitalia figured by Bethune Baker as those of *muhata* are instantly recognizable as those of the well known species *O. kirbyi* (*Pentila kirbyi* Aurivillius, 1895), the holotype of which I was able to dissect in 1947, and of which I have examined the genitalia of some fifty specimens from various African localities.

We have therefore a case in which (I) the species named as the type-species of the genus is a *species dubium* (*muhata*) and (2) the species on which the description of the genus was in fact based is an easily recognized species, currently known as *kirbyi*. In order to overcome the uncertainties of this situation I am applying to the International Commission on Zoological Nomenclature to set aside all type-fixations for the genus *Ornipholidotos* made prior to their ruling and to rule that the type-species of that genus be *Pentila kirbyi* Aurivillius (1895, *Ent. Tidskr.* 16: 198) as defined by Stempffer (1947, *Revue Zool. Bot. afr.* 40: 169).

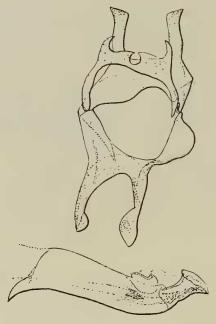


Fig. 12. Ornipholidotos kirbyi (Aurivillius), & genitalia.

If, as seems possible, it should transpire in the light of further evidence, that muhata and kirbyi become synonyms, the type-species will remain the same. Only its name will be changed.

In anticipation of a favourable decision on my application, I am proceeding on

the basis that the type-species of Ornipholidotos is O. kirbyi.

Head small; eyes large and naked; palpi very small and very short, divergent, clothed with scales below; antennae similar to those of Pentila: thorax short and slender; abdomen long, swollen in its apical portion, especially in the Q. Legs similar to those of Pentila.

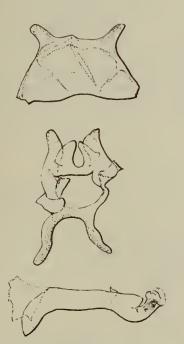
Wing shape: fore wing not so long as in Pentila: in all the species except O. paradoxa H. H. Druce, all four wings are translucent white with costal margin and the apex of the fore wings

and the hind margin of all four wings more or less broadly blackish.

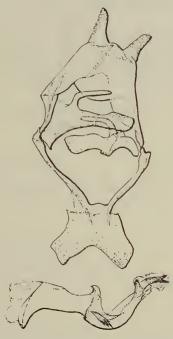
Wing venation (Text-fig. 235). Cell much elongated, though on the whole slightly shorter

than that of Pentila: discocellular shorter than in Pentila.

Male genitalia (Text-fig. 12, kirbyi). (See also Bethune Baker, Trans. ent. Soc. Lond. 1914: lviii, fig. 10, and Stempffer, 1947, Rev. Zool. Bot. afr. 30: 167, fig. 2). Uncus composed of two long robust arms; tegumen hood-shaped; articulated on the vinculum, on the tergite-sternite suture, there are two large asymmetrical processes, which are widened and fused distally and are provided with a strong hook; because of their point of attachment these processes cannot be regarded as subunci; these peculiar processes are found, though modified in shape, in all the other species of Ornipholidotos that I have studied; vinculum broad; there are no distinct articulated valves but only simple expansions of the vinculum which have a spatulate apex; penis elongate, its distal end bent back towards the dorsum, slightly constricted just before the



Ornipholidotos ntebi Fig. 13. (Bethune Baker), ♂ genitalia.



Ornipholidotos peucetia Fig. 14. (Hewitson), & genitalia.

broadened apex; vesica enclosing a strong spine surrounded by a cluster of smaller spines and, in addition, covered with little cornuti which give it a shagreened appearance; uncus bearing long strong bristles; there are long fine hairs at the apices of the vinculum expansions.

Elsewhere I have figured the male genitalia (see list below) of bakotae, bitjeensis, gabonensis, jacksoni, katangae, onitshae, overlaeti, teroensis, and ugandae. Here I figure in addition the genitalia of ntebi (Text-fig. 13), peucetia (Text-fig. 14), paradoxa (Text-fig. 15) and latimargo (Text-fig. 16).

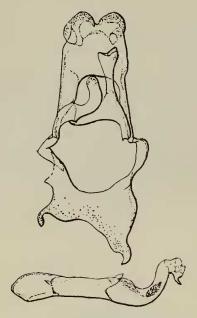


Fig. 15. Ornipholidotos paradoxa (Druce), & genitalia.

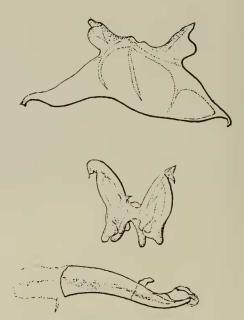


Fig. 16. Ornipholidotos latimargo (Hawker Smith), 3 genitalia.

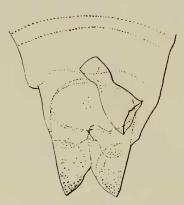


Fig. 17. Ornipholidotos peucetia (Hewitson), Q genitalia.

As one can ascertain by comparing all these figures, the male genitalia of the species of *Ornipholidotos* show an extraordinary diversity. Nevertheless they present good generic characters in the constant shape of the penis, the absence of true valves articulated on the vinculum, the presence of articulated processes on the tergite-sternite suture and the general asymmetry. The female genitalia of the species of *Ornipholidotoros* are also asymmetrical. In *O. peucetia* (Text-fig. 17, abdomen figured flattened dorso-ventrally), for example, notice that the ostium bursae, instead of being situated below the ostium oviductus, is twisted to the left through an angle of about 45 degrees and is visible to the naked eye as a small chitinous trumpet-shaped excrescence on the side of the abdomen.

There are two hypotheses to account for the external resemblance coupled with the diversity in the structure of the genitalia in the species of *Ornipholidotos*. Either we can suppose lines of descent from widely different ancestors, when the similarity in external characters would be due to convergent evolution similar to that which resulted in mimicry; or we can suppose a single line of descent giving rise to species whose external characters have remained pretty well constant, whilst the genitalia have evolved in a disorderly, anarchical manner, tending to produce species which are very diverse and sometimes unstable, as in *O. overlaeti*, in which species the dissection of over 50 males produced no two specimens the genitalia of which were rigorously identical, the variations in the shape of the uncus being sometimes quite considerable. Nevertheless this individual variation remains within such limits that one has no cause to doubt their specific identity. It is this observation which inclines me to accept the second hypothesis.

The caterpillar of O. muhata (?) has been described by T. H. E. Jackson (1937, Trans. R. ent. Soc. Lond. 86: 206). It feeds on lichens and resembles a Lepisma, being broadly rounded anteriorly and ending in a sharp point posteriorly. Each segment bears 3-4 black spines. The head is protected by a collar. Its green, grey and black colour blends with that of the lichen.

# LIST OF SPECIES OF Ornipholidotos

- \*Ornipholidotos bakotae Stempffer, 1962: 1137, fig. and fig. genitalia.
- \*Ornipholidotos bitjeensis Stempffer, 1957, Bull. Inst. fr. Afr. noire 19: 209, fig. genitalia.
- \*Ornipholidotos camerunensis Stempffer, 1964: 1227, fig.
- \*Ornipholidotos congoensis Stempffer, 1964: 1228, fig.
- Ornipholidotos emarginata (Hawker Smith), 1933, Stylops 2: 4.
- \*Ornipholidotos gabonensis Stempffer, 1947, Revue Zool. Bot. afr. 40: 169, fig. and fig. genitalia.
- \*Ornipholidotos jacksoni Stempffer, 1961: 11, fig. and fig. genitalia.
- \*Ornipholidotos katangae Stempffer, 1947, Revue Zool. Bot. afr. 40: 170, figs.
- \*Ornipholidotos kirbyi Aurivillius, 1895. Fig. Smith & Kirby, Rhop. exot. 2 (as muhata). Fig. genitalia, Stempffer, 1947, Revue Zool. Bot. afr. 40: 169.

Ornipholidotos kirbyi fumata f. (Schultze), 1923, Ergebn. 2te D. Z. Afr. Exp. 1910–11, 1: 1160.

\*Ornipholidotos latimargo (Hawker Smith), 1933, Stylops 2: 3.

Ornipholidotos muhata (Dewitz), 1886.

\*Ornipholidotos nigeriae Stempffer, 1964: 1230, fig.

\*Ornipholidotos ntebi (Bethune Baker), 1906.

\*Ornipholidotos onitshae Stempffer, 1962: 1135, fig. and fig. genitalia.

- \*Ornipholidotos overlaeti Stempffer, 1947, Revue Zool. Bot. afr. 40: 171, fig. and fig. genitalia.
- \*Ornipholidotos paradoxa (H. H. Druce), 1910.

\*Ornipholidotos perfragilis (Holland), 1890.

O. sylphida (Staudinger), 1891. Fig. Smith & Kirby, 1892.

Ornipholidotos peuceda (Smith), see peucetia.

\*Ornipholidotos peucetia (Hewitson), 1866. peuceda (Smith), 1889.

\*Ornipholidotos peucetia chyluensis (van Someren), 1939, Jl E. Africa Uganda nat. Hist. Soc. 14: 146.

Ornipholidotos peucetia orientalis (Storace), 1947, Annali Mus. civ. Stor. nat. Giacomo Doria 63: 77.

Ornipholidotos peucetia penningtoni (Riley), 1944, Entomologist 77: 29.

\*Ornipholidotos sylpha (Kirby), 1890. Fig. Smith & Kirby, 1892. Ornipholidotos sylphida (Staudinger), see perfragilis Holland.

\*Ornipholidotos teroensis Stempffer, 1957, Bull. Inst. fr. Afr. noire 19: 211, fig. genitalia.

Ornipholidotos tirza (Hewitson), 1873. Fig. Smith & Kirby, 1893.

\*Ornipholidotos ugandae Stempffer, 1947, Revue Zool. Bot. afr. 40: 170, fig. and fig. genitalia.

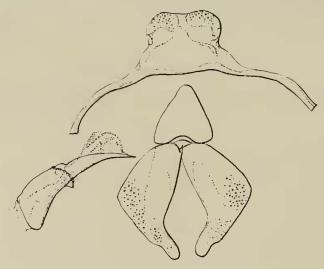


Fig. 18. Durbania amakosa Trimen, & genitalia.

## Genus DURBANIA Trimen

D'Urbania Trimen, 1862, Trans. ent. Soc. Lond. (3) 1:400; Aurivillius, 1898: 264. Durbania Trimen; Aurivillius, 1914:302; Murray 1935:49. Type-species: D'Urbania amakosa Trimen, by monotypy.

Head small, frons and vertex shortly pilose; eyes smooth; palpi fairly long, densely scaly, second segment long and robust, third short, slightly acuminate; antennae short, composed of 24 to 27 segments, rather thick, with a distinct cylindrical gradual club having the last four segments partly fused; thorax short, densely clothed with short hairs above and with scales and hair-scales below; abdomen of medium length, scaly; legs robust, clothed with scales but no hairs, tibiae with 2-3 delicate spines below, tarsi long, spinose beneath; fore tarsi of 3 unsegmented.

Wing venation (Text-fig. 236). Fore wing: vein 11 originates from the cell at a point nearer to the origin of vein 10 than to the base. Hind wing with a short precostal vein at the base of

the wing.

Male genitalia (Text-fig. 18). Uncus rounded with a shallow median depression, fused to a fairly large tegumen; no subunci; vinculum narrow and prolonged to form a large triangular saccus; valves oblong with a regular outline and rounded apex; base of penis robust, gradually becoming more slender distally, and ending with a sharp point with a wide opening on its dorsal surface; uncus and valves abundantly clothed with fine hairs.

The male genitalia of *D. limbata* are very close to those of *D. amakosa*, only the apex of the valves is lightly excised.

The early stages of D. amakosa have been described by Trimen (1887, S. Afr. Butterflies 2: 216).

## LIST OF SPECIES OF Durbania

Durbania amabilis Staudinger, see limbata.

\*Durbania amakosa amakosa Trimen, 1862.

Durbania amakosa ayresi van Son, 1941, Jl ent. Soc. sth. Afr. 4: 182.

Durbania amakosa natalensis van Son, 1959, Novos Taxa ent. 16: 8. Fig.

\*Durbania amakosa penningtoni van Son, 1959, Novos Taxa ent. 16: 17. Fig.

\*Durbania limbata Trimen, 1887.

amabilis Staudinger, 1888.

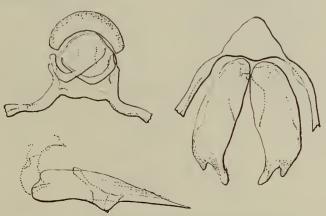


Fig. 19. Durbaniella clarki (van Son), & genitalia.

## Genus DURBANIELLA van Son

Durbaniella van Son, 1959, Novos Taxa ent. 16:10. Type-species: Durbania clarki van Son, 1941, by monotypy.

Differs from the genus *Durbania* in the following characters: *Palpi*, third joint very much reduced, not more than 1/5th the length of the second joint; antennae with 20 segments, club large and flattened.

Wing venation (van Son, 1959, Novos Taxa ent. 16, fig. 2). Fore wing, vein 11 arises from the cell midway between the base of the wing and the origin of vein 10. Hind wing with a small precostal vein.

Male genitalia (Text-fig. 19). Uncus crescent-shaped, fused to the margin of the rather large oval tegumen. Subunci long, curved, evenly tapering. Vinculum narrow, produced to form a wide triangular saccus. Valves oblong, the two processes separated apically, their extremities rounded, the lower process longer than the upper one. Penis basally stout, tapering evenly and ending in a sharp point broadly open dorsally. Uncus and apex of valves hairy.

## LIST OF SPECIES OF Durbaniella

\*Durbaniella clarki (van Son), 1941, Jl ent. Soc. sth. Afr. 4: 183; fig., 1959, Novos Taxa ent. 16: 10.

## Genus DURBANIOPSIS van Son

Durbaniopsis van Son, 1959, Novos Taxa ent. 16:12. Type-species: Durbania saga Trimen, 1883, by monotypy.

D'Urbania Trimen (partim); Aurivillius, 1898: 265.

Durbania Trimen (partim); Aurivillius, 1914; 303; Murray, 1935: 52.

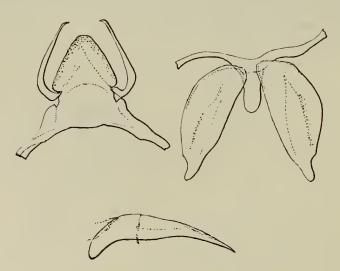


Fig. 20. Durbaniopsis saga (Trimen), & genitalia.

Palpi densely scaled, third joint much reduced, subconic, barely one-fourth the length of second joint; antennae with 32 segments, the club large and flattened.

Wing venation (see van Son, 1959, Novos Taxa ent. 16: fig. 3). Vein 11 of fore wing arises much nearer to the origin of vein 10 than to the base of the wing. Hind wing cell much shorter

than in Durbaniella and with a small precostal vein.

Male genitalia (Text-fig. 20). Uncus crescent-shaped with rounded apex and straight sides, fused to the subtriangular tegumen. Subunci very long, slender, curved near their base; shown spread out in the illustration, in their normal position they lie folded under the tegumen. Vinculum narrow, prolonged in a triangular saccus. Valves oblong, the apex rounded, not divided. Penis like that of *Durbania* and *Durbaniella*. Uncus and distal part of the valves covered with fine hair.

The simple symmetrical genitalia of *Durbania*, *Durbaniella* and *Durbaniopsis* show common characters, although subunci are lacking in *Durbania*. They are very far removed from those of *Telipna*, *Pentila* and *Ornipholidotos*. In spite of the presence of a precostal vein on the hind wings in both these groups of genera, I consider they are not truly related.

# LIST OF SPECIES OF Durbaniopsis

\*Durbaniopsis saga (Trimen), 1883. Fig. Aurivillius in Seitz, 1914.

## Genus MIMACRAEA Butler

Mimacraea Butler, 1872, Lep. exot.: 104; Aurivillius, 1898: 265, 1918: 312; Pinhey, 1949: 97. Type-species: Mimacraea darwinii Butler, 1872, by original designation.

Eyes smooth; palpi moderately long, clothed below with black scales, the third segment bearing a few white scales at the apex; antennae about half as long as the costa, club elongated and progressively swollen; thorax rather slender; abdomen long, reaching slightly beyond the anal angle; legs robust, black, white-annulated, fore tarsi of 3 unsegmented and finely spinose below.

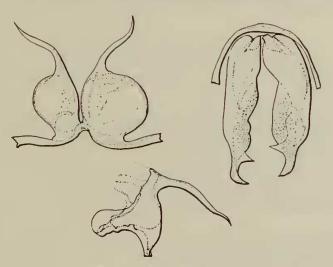


Fig. 21. Mimacraea darwinii Butler, & genitalia.

Wing venation (Text-fig. 237). Fore wing cell short and not reaching the middle of the wing; vein 7 arising from 6, 8 and 9 from 7. Hind wing cell short, not reaching the middle of the wing; 7 arising from 6.

Male genitalia (Text-fig. 21). Uncus composed of two large hemispherical lobes which are hollow and distinctly asymmetrical, fused together at the base for a very short distance, each lobe bearing anteriorly a very long, tapered, horn-shaped process; no subunci; tegumen reduced to a narrow strip; vinculum narrow. Lower fultura welded to a fold at the base of the valves and closely sheathing the base of the penis; valves oblong, with the upper process bearing two strong apical teeth, the lower process semi-membranous. Penis of a peculiar shape, the massive base widely open proximally for the passage of the ductus, the distal half a long, slender, slightly curved point at right-angles to the base; there are some pubescence on the uncus, many long hairs on the lower process of the valves and a few short hairs on the upper process.

The male genitalia of all the species of *Mimacraea* that I have been able to examine are extremely similar and of the *darwinii* type; as specific characters one can only point to slight modifications in the form of the apex of the valves.

The species of *Mimacraea* have a very characteristic facies, which enables one to distinguish them at once from related genera; they resemble, often in an almost startling manner, certain species of Acraeinae of the genera *Acraea* and *Bematistes*. Since the Acraeinae are not attacked by insectivorous predators, because of their nauseating smell, *Mimacraea* is often quoted as an example of Batesian mimicry. As in the case of most mimetic species, the colours and patterns in *Mimacraea* are very variable individually. It seems to me probable that some forms, described as species, are in fact really subspecies or even individual forms.

The early stages of M. marshalli dohertyi Rothschild have been observed by van Someren (see Poulton, 1924, Trans. ent. Soc. Lond. 1924: 152). The larva is clothed in small barbed spicules and very long fine hair which spreads laterally and gives it the appearance of a moth larva; it feeds on lichens and is nocturnal. The larva of M. krausei Dewitz has been briefly described by T. H. E. Jackson (1937, Trans. R. ent. Soc. Lond. 86: 207). It is similar to that of M. dohertyi but much darker brown and of the same habits.

## LIST OF SPECIES OF Mimacraea

Mimacraea angustata Schultze, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910–11, 1:1163.

\*Mimacraea apicalis Smith & Kirby, 1890.

\*Mimacraea charmian charmian Smith & Kirby, 1890.

Mimacraea charmian ertli Talbot, 1924, Entomologist 57: 38.

Mimacraea costleyi H. H. Druce, 1912.

\*Mimacraea darwinii Butler, 1872. (darwinia emend.).

Mimacraea eltringhami eltringhami H. H. Druce, 1912.

Mimacraea eltringhami burgeoni Hawker Smith, 1928, Revue Zool. Bot. afr. 16: 214.

\*Mimacraea flavefasciata Schultze, 1912.

Mimacraea fulvaria Aurivillius, 1895.

Mimacraea gelinia (Oberthur), 1893.

Mimacraea graeseri Schultze, 1912.

\*Mimacraea krausei krausei Dewitz, 1889.

\*Mimacraea krausei ab. obsolescens Hawker Smith, 1926, Revue zool. afr. 14:238.

Mimacraea krausei f. citrifascia Talbot, 1935, Entomologist's mon. Mag. 71: 71. Mimacraea krausei f. viviana Talbot, 1935, Entomologist's mon. Mag. 71: 71, fig.

Mimacraea krausei elgonae Talbot, 1935, Entomologist's mon. Mag. 71: 71, fig. Mimacraea krausei karschioides Carpenter & Jackson, 1950, Proc. R. ent. Soc. Lond. (B) 19: 106.

\*Mimacraea krausei masindae Bethune Baker, 1913.

\*Mimacraea krausei poultoni Neave, 1904.

luteomaculata Grünberg, 1908.

\*Mimacraea lacta Schultze, 1912. Fig. Aurivillius in Seitz, 1918.

\*Mimacraea landbecki landbecki H. H. Druce, 1910.

Mimacraea landbecki ab. latifascia Rebel, 1914, Annln naturh. Mus. Wien 28: 263, fig.

Mimacraea landbecki ab. flavescens Rebel, 1914, Annln naturh. Mus. Wien 28: 263, fig.

Mimacraea luteomaculata Grünberg, see krausei poultoni.

Mimacraea mariae Dufrane, 1945, Bull. Annls Soc. R. ent. Belg. 81: 115.

\*Mimacraea marshalli marshalli Trimen, 1898.

\*Mimacraea marshalli dohertyi Rothschild, 1901.

\*Mimacraea marshalli dohertyi f. somereni Talbot, 1937, Trans. R. ent. Soc. Lond. 86: 63, fig.

\*Mimacraea marshalli media Talbot, 1937, Trans. R. ent. Soc. Lond. 86: 63, fig. Mimacraea marshalli nzoia Talbot, 1937, Trans. R. ent. Soc. Lond. 86: 64, fig. Mimacraea neavei Eltringham, 1909.

Mimacraea neokoton H. H. Druce, 1907.

\*Mimacraea neurata neurata Holland, 1895.

Mimacraea neurata alciopina Joicey & Talbot, 1924, Entomologist 57: 38.

\*Mimacraea neurata f. incurvata Talbot, 1935, Entomologist's mon. Mag. 71:205.

Mimacraea neurata lineata Talbot, 1935, Entomologist's mon. Mag. 71: 205.

\*Mimacraea paragora Rebel, 1911.

Mimacraea pseudepaea Schultze, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910–11, 1: 1165.

Mimacraea pulverulenta Schultze, 1912.

Mimacraea schmidtii Schultze, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910-11, 1:1163.

Mimacraea schubotzi Schultze, 1912.

Mimacraea skoptolos H. H. Druce, 1907.

Mimacraea telloides Schultze, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910-11, 1:1165.

## Genus MIMERESIA Stempffer

Mimeresia Stempffer, 1961 Annls Mus. R. Afr. cent. Sér. 4to 94: 24. Type-species: Liptena libentina Hewitson, 1886, by original designation.

Pseuderesia Butler (pro parte); Aurivillius, 1898: 267; 1918: 318.

Eyes large and smooth; palpi rather long, ascending, extending broadly beyond the frons, the second joint densely scaly, the third short and pointed; antennae long and delicate, white-banded, the club progressively expanding; thorax moderately stout, abdomen long and reaching beyond the anal angle; in the  $\Im$  the tibiae of the fore leg often swollen (Text-fig. 23, libentina) and clothed in closely adpressed scales, tarsi not jointed; in the  $\Im$  the tibiae not swollen.

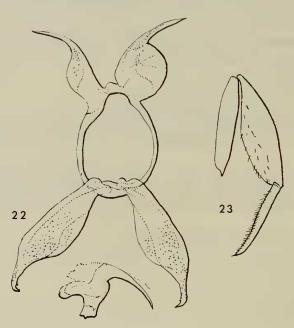
Wing venation (Text-fig. 238). On all four wings the cell is longer than in Mimacraea and reaches about midway. On the hind wing vein 5 arises nearer to 6 + 7 than to 4.

Male genitalia. Very similar to those of Mimacraea, except that the apex of the valves is falcate and not crescentic.

The genitalia of all the species of *Mimeresia* are very like those of *libentina* (Textfig. 22) and far removed from *Pseuderesia*. Such close similarity cannot be attributed to chance. One cannot doubt the ancestral affinity of *Mimacraea* and *Mimeresia*, the two genera only being separable in form, pattern and slight differences in wing venation. The resemblance in facies between *Mimeresia* and *Pseuderesia* is at present not satisfactorily explained.

## LIST OF SPECIES OF Mimeresia

Mimeresia alberici (Dufrane), see (3) neavei and  $(\mathfrak{P})$  moreelsi purpurea. Mimeresia ashira (Holland), see debora.



Figs 22-23. Mimeresia libentina (Hewitson), & genitalia and & anterior leg.

Mimeresia carlota (Suffert), see dinora.

- \*Mimeresia cellularis (Kirby), 1890 (Sept.). Fig. Smith & Kirby, 1891. mondo (Holland), 1890 (November).
- \*Mimeresia debora debora (Kirby), 1890 (Sept.). Fig. Smith & Kirby, 1891. ashira (Holland), 1890 (November).
- \*Mimeresia debora barnsi (Hawker Smith), 1933, Stylops 2:5. Fig. Stempffer, 1961.
- \*Mimeresia debora catori (Bethune Baker), 1904. Fig. Stempffer, 1961.
- \*Mimeresia debora deborula (Aurivillius), 1898. Fig. Smith & Kirby, 1894, as debora Q.
- \*Mimeresia dinora dinora (Kirby), 1890. Fig. Smith & Kirby, 1891. carlota (Suffert), 1904.
- \*Mimeresia dinora discirubra (Talbot), 1937, Trans. R. ent. Soc. Lond. 86: 60. Fig. Stempffer, 1954, Bull. Soc. ent. Fr. 59: 89.
- \*Mimeresia drucei drucei (Stempffer), 1954, Bull. Soc. ent. Fr. 59:89.
- \*Mimeresia drucei owerri Stempffer, 1961: 33, fig.
- \*Mimeresia drucei ugandae (Stempffer), 1954, Bull. Soc. ent. Fr. 59: 91.
- \*Mimeresia favillacea favillacea (Grünberg), 1910. Fig. Aurivillius in Seitz, 1918.
- \*Mimeresia favillacea griseata (Talbot), 1937, Trans. R. ent. Soc. Lond. 86: 61.
- \*Mimeresia libentina libentina (Hewitson), 1866.
- Mimeresia libentina f. isabellae (Schultze), 1916, Arch. Naturgesch. 82 A3: 36.
- \*Mimeresia libentina f. zerita (Plötz), 1880. rubrica (H. H. Druce), 1888; zoraida (Smith & Kirby), 1890.
  - Mimeresia mondo (Holland) see cellularis (Kirby).
- \*Mimeresia moreelsi moreelsi (Aurivillius), 1901. Fig. Aurivillius in Seitz, 1918.
- \*Mimeresia moreelsi purpurea (Hawker Smith), 1933, Stylops 2:5. alberici ♀ (Dufrane), 1945.
- \*Mimeresia moreelsi tessmanni (Grünberg), 1910. Fig. Aurivillius in Seitz, 1918.
- Mimeresia moreelsi tessmanni f. decolorata (Hulstaert), 1924, Revue zool. afr. 12:117.
- \*Mimeresia moyambina (Bethune Baker), 1904. Fig. Stempffer, 1961.
- \*Mimeresia neavei (Joicey & Talbot), 1921, Bull. Hill Mus. Witley 1:80. Fig.
- Mimeresia rubrica (H. H. Druce), see libentina f. zerita.
- \*Mimeresia russulus russulus (H. H. Druce), 1910.
- \*Mimeresia russulus katangae (Hawker Smith), 1926, Revue zool. afr. 14: 239.
- \*Mimeresia russulus unyoro Stempffer, 1961: 39. Fig.
- \*Mimeresia semirufa (Smith), 1902.
  - Mimeresia zoraida (Smith & Kirby), see libentina f. zerita.

## Genus PSEUDERESIA Butler

- Pseuderesia Butler, 1874, Trans. ent. Soc. Lond. 84: 532. Type-species: Pseuderesia catharina Butler, 1874 (= Pentila eleaza Hewitson, 1873) by original designation.
- Pseuderesia Butler (pro parte); Aurivillius, 1898: 266; 1918: 318.

Eyes large, smooth; palpi fairly long, ascending and projecting well beyond the frons, second segment clothed with long, erect hair, third segment short; antennae long, slender, white-banded, club well differentiated, clavate; thorax moderately stout, abdomen long and reaching beyond anal angle in the male;  $\eth$  fore leg with tibia not swollen, tarsus unsegmented, finely spinose below.

Wing venation (Text-fig. 239). In the  $\circlearrowleft$  of eleaza on the fore wings vein 10 arises from the upper angle of the cell; in the  $\circlearrowleft$  it branches from the common stem of vein 6 and 7 + 8 + 9; in isca vein 10 arises from the anterior margin of the cell. On the hind wing vein 5 is nearer to 6 + 7 than to 4 in all species.

Male genitalia (Text-fig. 24). Uncus deeply divided, formed of two long symmetrical leaf-like expansions with pointed apices. Subunci absent. Tegumen reduced to a narrow strip. Vinculum narrow with an indistinct saccus. Lower fultura fused to the base of the valves and sheathing the internal part of the penis. Valves oblong, slightly falcate at the apex. Penis small, proximally widely open dorsally, the distal (outer) part strongly recurved. Uncus delicately pubescent, distal half of valves thickly hairy.

All the male genitalia of *Pseuderesia* that I have examined bear a definite resemblance to those of *P. eleaza*: the uncus is always bifid, though less deeply divided, the subunci are absent, the valves oblong. The penis of *P. paradoxa* shows a certain resemblance to that of *eleaza*, but those of *isca* (Text-fig. 25), *ouesso*, *nigeriana*, *phaeochiton*, *osheba*, *clenchi*, *rougeoti*, *beni* and *rutilo*, differ in that the distal portion of the penis is not recurved. The penis of *bicolor* (Text-fig. 26) is of a very different type, the outer portion very long, cylindrical, bent at an obtuse angle and open dorsally to allow the passage of the vesica, which bears numerous cornuti.

#### LIST OF SPECIES OF Pseuderesia

- \*Pseuderesia beni Stempffer, 1961: 19. Fig.
- \*Pseuderesia bicolor Smith & Kirby, 1890.
- \*Pseuderesia clenchi Stempffer, 1961: 22. Fig.

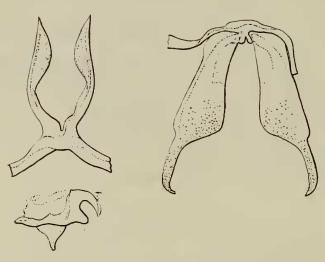


Fig. 24. Pseuderesia eleaza eleaza (Hewitson), of genitalia.

- \*Pseuderesia eleaza eleaza (Hewitson), 1873. picta Smith, 1898; variegata (Smith & Kirby), 1890.
- \*Pseuderesia eleaza catharina Butler, 1874.
- \*Pseuderesia eleaza katera Stempffer, 1961: 14.
- \*Pseuderesia eleaza nigra Cator, 1904.
- \*Pseuderesia eleaza vidua Talbot, 1937, Trans. R. ent. Soc. Lond. 86: 60. Fig.
- \*Pseuderesia isca isca (Hewitson), 1873.
  - Pseuderesia isca ab. demaculata Hulstaert, 1924, Revue zool. afr. 12:117.
- \*Pseuderesia isca magnimaculata Rebel, 1914, Annln naturh. Hofmus. Wien 28: 264.

Pseuderesia mildbraedi Schulze, see osheba.

Pseuderesia minium H. H. Druce, see osheba.

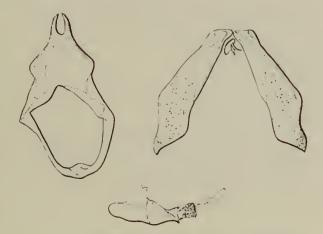


Fig. 25. Pseuderesia isca (Hewitson), of genitalia.

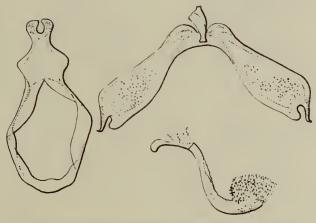


Fig. 26. Pseuderesia bicolor Smith & Kirby, & genitalia.

- \*Pseuderesia nigeriana Stempffer, 1962: 1139. Fig.
- \*Pseuderesia osheba (Holland), 1890. minium H. H. Druce, 1910; mildbraedi Schultze, 1912.
- \*Pseuderesia onesso Stempffer, 1962: 1143. Fig.

Pseuderesia paradoxa paradoxa Schultze, 1916, Arch. Naturgesch. 82, A3: 37.

\*Pseuderesia paradoxa orientalis Stempffer, 1962: 1145. Fig.

- \*Pseuderesia phaeochiton Grünberg, 1910. Fig. Aurivillius in Seitz, 1918. Pseuderesia picta Smith, see eleaza.
- \*Pseuderesia rougeoti Stempffer, 1961: 21. Fig.
- \*Pseuderesia rutilo H. H. Druce, 1910.

Pseuderesia variegata Smith & Kirby, see eleaza.

## Species doubtfully referred to Pseuderesia

Pseuderesia cornucopiae (Holland), 1892.

Pseuderesia fusca Cator, 1904.

Pseuderesia mapongua (Holland), 1893.

Not having been able to examine these species I am unable to decide whether they belong to *Mimeresia* or to *Pseuderesia*.

Peters (1952, Check List Butt. Eth. Region) included in his list of species of Pseuderesia, under No. 1313a, Pseuderesia "amaurina Neu. 1928". This is an evident confusion with Pseudacraea amaurina Neustetter (1928, Int. ent. Z. 21:445). Neustetter did not describe any species of Pseuderesia under that name.

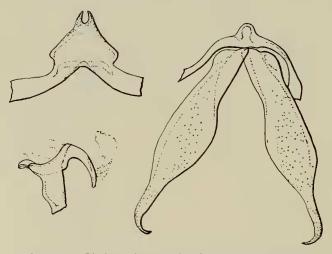


Fig. 27. Citrinophila marginalis Kirby, 3 genitalia.

## Genus CITRINOPHILA Kirby

Citrinophila Kirby, 1887, Ann. Mag. nat. Hist. (5) 19: 367; Aurivillius, 1898: 268; 1918: 325. Type-species: Citrinophila marginalis Kirby, by original designation.

Eyes naked: palpi fairly long, second segment laterally compressed, third segment slender, acuminate; antennae black and white annulated, club sharply distinct, elongate, flattened; legs black and white annulated, fore tarsi of 3 unsegmented, finely spinose below.

Wing venation (Text-fig. 240). Differs from Pseuderesia in that the discoidal on the hind wing is straight between the points of origin of veins 4 and 5.

Male genitalia (Text-fig. 27). Uncus composed of two narrow lateral bands on the posterior margin of the tegumen each ending in a small sharply pointed process and separated by a deep concavity; no subunci; tegumen very large, subtriangular; vinculum broad, with a small

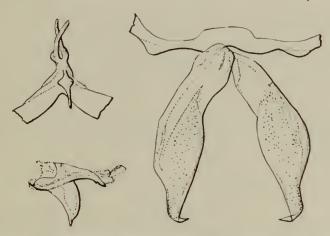


Fig. 28. Citrinophila unipunctata Bethune Baker, & genitalia.

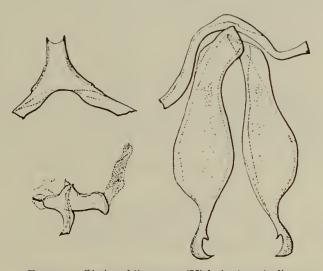


Fig. 29. Citrinophila tenera (Kirby), & genitalia.

rounded saccus; lower fultura fused to the base of the valves and sheathing the internal portion of the penis; valves markedly oblong, with sharp, slightly falcate apices; penis short, the external portion strongly recurved, with a wide opening on the dorsal surface, uncus and valves pilose.

Kirby has described several species which closely resemble marginalis, one of them (limbata) in the genus Citrinophila, the others tenera, serena and similis in the genus Teriomima. It is difficult to identify these species with certainty, as the available specimens do not agree precisely with the original descriptions, and Kirby did not state the sex of his holotypes. Aurivillius (1918: 326) established the following pairs of synonyms, similis and marginalis, and tenera (Text-fig. 29) with limbata.

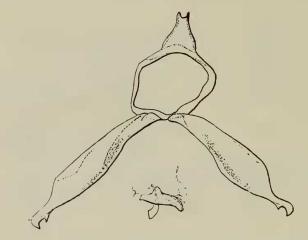


Fig. 30. Citrinophila terias Joicey & Talbot, & genitalia.

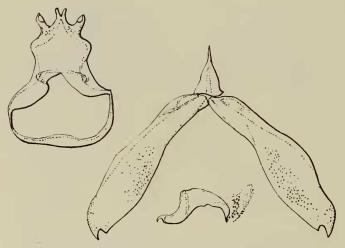


Fig. 31. Citrinophila erastus (Hewitson), & genitalia.

A specimen which I believe to be referable to *limbata*, because of its faintly orange colour, has genitalia exactly like those of *marginalis*.

Citrinophila terias (Text-fig. 30) has armature in which the dorsum and the valves resemble the corresponding parts in marginalis but the distal part of the penis is only very slightly recurved.

In Citrinophila unipunctata (Text-fig. 28) the dorsum is very different from that of the preceding species: the uncus is formed of two long finger-like processes with rounded apices, and there is a lanceolate process fused at the base of the uncus and directed anteriorly; subunci lacking; tegumen much reduced; vinculum broad with faintly indicated saccus; valves like those of marginalis; penis feebly curved; uncus and valves pilose.

Citrinophila erastus (Text-fig. 31). Uncus composed of four short processes, those of the median pair divergent from the median line, the lateral pair with a shallow apical cavity; subunci lacking; tegumen lozenge-shaped; vinculum rather broad; valves oblong, the upper process terminating in a stout tooth; penis short, curved, vesica bearing stout cornuti. Uncus and lower margin and apices of the valves pilose.

It is apparent that the unci in the four species described above are of very different types. The genus *Citrinophila* is thus not very homogeneous, and the similarities of the facies of the included species may be the result of convergent evolution.

# LIST OF SPECIES OF Citrinophila

\*Citrinophila erastus erastus (Hewitson), 1866.

Citrinophila erastus ab. erasmus (Kirby), 1887.

Citrinophila erastus ab. flaveola (Kirby), 1887.

Citrinophila erastus ab. vulcanica Schulze, 1916, Arch. Naturgesch. 82 A 3:37.

- \*Citrinophila erastus pallida Hawker Smith, 1933, Stylops 2:6.
- \*Citrinophila limbata Kirby, 1887. Fig. Smith & Kirby, 1888.
- \*Citrinophila marginalis Kirby, 1887. Fig. Smith & Kirby, 1888.

Citrinophila pusio (Smith), 1898.

Citrinophila regularis Schulze, see terias.

Citrinophila serena (Kirby), 1890. Fig. Smith & Kirby, 1891.

Citrinophila similis (Kirby), 1887. Fig. Smith & Kirby, 1888.

Citrinophila tenera (Kirby), 1887. Fig. Smith & Kirby, 1888.

- \*Citrinophila terias Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1:82. Fig. regularis Schulze, 1923.
- \*Citrinophila unipunctata Bethune Baker, 1908.

# Genus TERIOMIMA Kirby

Teriomima Kirby, 1887, Ann. Mag. nat. Hist. (5) 19: 364; Stempsfer & Bennett, 1953, Bull. Br. Mus. nat. Hist. (Ent.) 3: 81. Type-species: Teriomima subpunctata Kirby, by original designation.

Teriomima Kirby (pro parte); Aurivillius, 1898: 270; 1918: 327; Swanepoel, 1953: 188.

Eyes smooth; palpi rather long, ascending, projecting beyond the frons, clothed with adpressed scales, the second segment stout, laterally compressed, the third segment slender, acuminate; antennae rather short with a distinct, subcylindrical club;  $\beta$  fore legs with unsegmented tarsi, clothed below with fine spines.

Wing venation (Text-fig. 241).

Male genitalia (Text-fig. 32). Uncus bifid, lobes dilated at the base and tapering to blunt points bent ventrad; no subunci; tegumen rather large, vinculum rather wide; fultura inferior sheathing the base of the penis; valves oblong, slightly falcate at the apex and bearing a long gently curved harpe, penis long and stout, curved in the shape of a flattened S and bearing, near the blunt distal end, two ventrally directed lobes.

The male genitalia of all the species of *Teriomima* are figured by Stempffer and Bennett (1953). They are of the same type as those of *subpunctata*, although in *micra* and *parva* the valves are devoid of harpes.

## LIST OF SPECIES OF Teriomima

Teriomima delicatula Kirby, see subpunctata.

\*Teriomima micra (Grose Smith), 1898.

\*Teriomima parva Hawker Smith, 1933, Stylops 2:6.

\*Teriomima puella Kirby, 1887. Fig. Smith & Kirby, 1888.

\*Teriomima puellaris (Trimen), 1894.

\*Teriomima subpunctata Kirby, 1887. Fig. Smith & Kirby, 1888. delicatula Kirby, 1890.

\*Teriomima zuluana van Son, 1949, Ann. Transv. Mus. 21: 211. Fig.

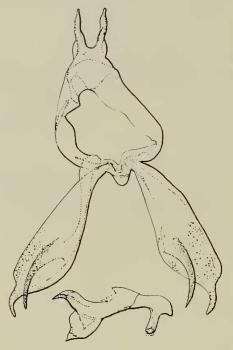


Fig. 32. Teriomima subpunctata Kirby, & genitalia.

## Genus EUTHECTA Bennett

Euthecta Bennett, 1954, Entomologist 87:170. Type-species: Euthecta cooksoni Bennett, 1954, by monotypy.

Eyes, palpi, antennae, legs and wing venation as in Teriomina.

Male genitalia (Text-fig. 33): uncus conical, rather concave at the apex; no subunci; no special processes as in Baliochila. Penis very long, sickle-shaped, the basal part swollen, the external part ending in a sharp point.

### LIST OF SPECIES OF Euthecta

\*Euthecta cooksoni Bennett, 1954, Entomologist 87: 171.

# Genus BALIOCHILA Stempffer & Bennett

Baliochila Stempsfer & Bennett, 1953, Bull. Br. Mus. nat. Hist. (Ent.) 3:85. Type-species: Liptena aslauga Trimen, 1873, by original designation.

Teriomima Kirby (pro parte); Anrivillius, 1898: 271; 1918: 328; Murray, 1935: 54; Swanepoel, 1953: 187.

Eyes, palpi, antennae, legs and wing venation as in Teriomima.

Male genitalia (Text-fig. 34). The of genitalia of all the species of Baliochila have been described and figured by Bennett & Stempsfer (1953). With the exception of B. singularis they present a common character which has justified the erection of the genus, viz. the presence of a pair of symmetrical processes, of various length, the bases of which are firmly attached to the dorsal face of the anellus internally, the outer margin being attached to the tegumen by a semi-membranous union. In the natural position these processes extend horizontally, on the axis of the abdomen, between the uncus and the penis; they are sometimes crowned with spines, sometimes entirely covered with short stiff hair. One cannot regard them as subunci, since they are not articulated to the uncus-tegumen suture; on the other hand, one cannot homologize them exactly with the asymmetrical processes of Ornipholidotos, which are articulated with the suture between tergite and sternite. For these reasons they are designated "special processes"

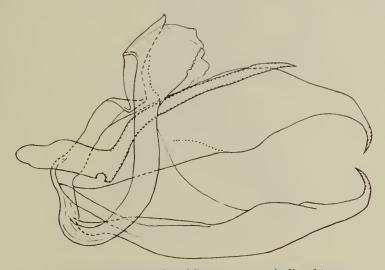


Fig. 33. Euthecta cooksoni Bennett, & genitalia of type.

in the descriptions of these species. The valves of *Baliochila* are oblong, more or less falcate at the apex, and the penis never bears lobes distally like those of *Teriomima subpunctata*.

On the basis of the morphology of the male genitalia, *Baliochila* can be divided into four groups of species:—

a. Uncus bifid, the two branches widely separate, more or less slender, special processes rather short (Stempffer & Bennett, figs. 32-43):—

Baliochila aslauga, barnesi, neavei, hildegarda, dubiosa, nyasae, stygia.

b. Uncus in form of a flattened median process excavate at the extremity, the special processes long and slender (figs. 44-45):—

Baliochila woodi, fragilis.

c. Uncus bifid, the two branches slender and ending in a point, special processes long and stout (figs. 46-50) :—

Baliochila minima, lipara.

d. Uncus bifid, its two branches carried on a long common stem, no special processes, but the anellus bearing paired long slender processes (fig. 51):—

Baliochila singularis.

### LIST OF SPECIES OF Baliochila

- \*Baliochila aslauga (Trimen), 1873. Fig. Trimen, 1887.
- \*Baliochila barnesi Stempffer & Bennett, 1953: 86. Fig.
- \*Baliochila dubiosa Stempffer & Bennett, 1953: 90. Fig.
- \*Baliochila fragilis Stempffer & Bennett, 1953: 95. Fig.

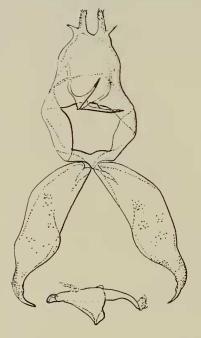


Fig. 34. Baliochila aslauga (Trimen), of genitalia.

Baliochila freya (Smith & Kirby), see hildegarda.

\*Baliochila hildegarda (Kirby), 1887. Fig. Smith & Kirby, 1888. freya (Smith & Kirby), 1894.

\*Baliochila lipara Stempffer & Bennett, 1953: 99. Fig.

\*Baliochila minima minima (Hawker Smith), 1933, Stylops 2: 6. Fig. Stempffer & Bennett, 1953: pl. 7, fig. 44.

\*Baliochila minima amanica Stempffer & Bennett, 1953: 97. Fig.

\*Baliochila minima latimarginata (Hawker Smith), 1933, Stylops 2:7. Fig. Stempffer & Bennett, 1953: pl. 7, fig. 47.

\*Baliochila neavei Stempffer & Bennett, 1953: 88. Fig.

\*Baliochila nyasae Stempffer & Bennett, 1953: 92. Fig.

\*Baliochila petersi Stempffer & Bennett, 1956, Bull. Inst. fr. Afr. noire 18: 503.

\*Baliochila singularis Stempffer & Bennett, 1953: 100. Fig.

\*Baliochila stygia (Talbot), 1935, Entomologist's mon. Mag. 71: 72. Fig.

\*Baliochila woodi (Riley), 1943, Entomologist 76: 225.

# Genus CNODONTES Stempffer & Bennett

Cnodontes Stempffer & Bennett, 1963, Bull. Br. Mus. nat. Hist. (Ent.) 3: 101. Type-species: Durbania pallida Trimen, 1898, by original designation.

Teriomima Kirby (pro parte); Aurivillius, 1898: 271; 1918: 328; Murray, 1935: 55; Swanepoel, 1953: 188.

Eyes, palpi, antennae, legs and venation as in Teriomima and Baliochila.

Male genitalia (Text-fig. 35). The 3 genitalia of all the species of Cnodontes have been figured either by Stempffer & Bennett in the paper referred to above, or by Bennett in the Entomologist (1954, 87: 172; 1956, 89: 115). They present a structural peculiarity not observed in any

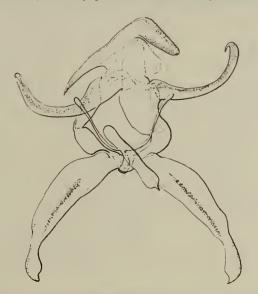


Fig. 35. Cnodontes pallida (Trimen), of genitalia.

other holarctic, aethiopian or indo-malayan Lycaenid: the uncus is fused with the eighth tergiter which forms a kind of hood or sheath above it. Subunci are lacking. Tegumen triangular, its base bearing a pair of long finger-like processes which are not in any way attached to the anellus. Vinculum broad. Valves as in *Baliochila*. To the dorsal surface of the anellus are fused two long finger-like processes lying parallel to those of the tegumen. It seems that in *Cnodontes* the "special processes" of *Baliochila* are divided to form two independent pairs, one attached to the tegumen, the other to the anellus. Penis long. Uncus and valves pilose.

### LIST OF SPECIES OF Cnodontes

- \*Cnodontes pallida (Trimen), 1898.
- \*Cnodontes penningtoni Bennett, 1954, Entomologist 87: 171. Fig.
- \*Cnodontes vansomereni Stempffer & Bennett, 1953: 103. Fig.
- \*Cnodontes vansoni Stempffer & Bennett, 1956, Entomologist 89: 115. Fig.

### Genus LARINOPODA Butler

Larinopoda Butler, 1871, Trans. ent. Soc. Lond. 172. Aurivillius, 1898: 271; 1918: 328. Type-species: Larinopoda lycaenoides Butler, 1871 (= Liptena lircaea Hewitson, 1866), by monotypy.

Eyes smooth; palpi with second segment long and clothed with adpressed scales, third segment slender, acuminate; antennae short, slender, with a slightly swollen, flattened club; thorax short, robust; abdomen long with the apical segments much swollen in the  $\mathsep$ ; legs strong, clothed with scales, the  $\mathsep$  fore tarsi unsegmented, finely spinose below.

Wing venation (Text-fig. 242). Fore wing cell short, produced at the lower angle. Hind

wing cell much produced at its lower angle.

Male genitalia (Text-fig. 36). Both the anterior and posterior edges of the uncus have a rounded depression, their two apices form a blunt point, the sides are slightly concave; subunci long and curved, the basal portion swollen in the middle, the apex of the free portion rounded and without a terminal hook; tegumen fairly large; vinculum wide; valves oval with rounded

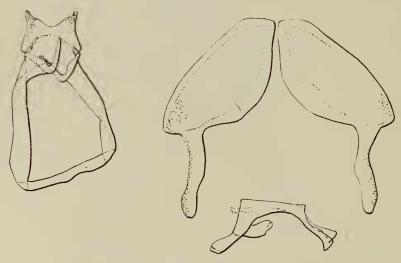


Fig. 36. Larinopoda lircaea (Hewitson), of genitalia.

apex, the lower processus near its apex bearing a long finger-like process of which the apex is rounded; internal portion of penis curved, and connected with the base of the valves by a short peduncle representing the lower fultura; the external portion of the penis forming an obtuse angle with the internal portion, the distal third divided into two branches with spatulate apices; uncus, distal portion and finger-like process of the valves all bear a few fine hairs.

Eltringham (1922, Trans. ent. Soc. Lond. 1922: 254) described and figured the genitalia of most of the species of Larinopoda. They are all of a type very similar to those of lircaea. The genus is very homogeneous both in male genitalia and in facies, which recalls the European Pieridae; Butler even placed his genus Larinopoda between Euchloe and Nepheronia in that family.

# LIST OF SPECIES OF Larinopoda

Larinopoda aspidos aspidos H. H. Druce, 1890. Fig. Eltringham, 1922.

Larinopoda aspidos f. brenda H. H. Druce, 1903.

Larinopoda aspidos f. latimarginata Smith, 1898. Fig. Aurivillius in Seitz, 1918.

Larinopoda batesi Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 389.

\*Larinopoda eurema (Plötz), 1880.

varipes Kirby, 1887; libussa Staudinger, 1888.

\*Larinopoda lagyra (Hewitson), 1866.

lara Staudinger, 1891.

Larinopoda lagyra f. emilia Suffert, 1904.

Larinopoda lagyra f. emilia ab. deficiens Dufrane, 1953, Bull. Annls Soc. R. ent. Belg. 89: 48.

Larinopoda lagyra f. gyrala Suffert, 1904. Fig. Aurivillius in Seitz, 1918.

Larinopoda lagyra f. punctata H. H. Druce, 1910.

Larinopoda lara Staudinger, see lagyra.

Larinopoda libussa Staudinger, see eurema. \*Larinopoda lircaea (Hewitson), 1866.

lycaenoides Butler, 1871.

Larinopoda lircaea f. hermansi Aurivillius, 1896.

Larinopoda lircaea f. innocentia Gaede, 1915, Int. ent. Z.: 111.

Larinopoda lircaea f. spuma H. H. Druce, 1910.

Larinopoda lircaea ab. alenica Strand, 1914, Arch. Naturgesch. 79 A 12: 133.

Larinopoda lircaea ab. alenicola Strand, 1914, l.c.: 133.

Larinopoda lircaea ab. benitonis Strand, 1914, l.c.: 134.

Larinopoda lircaea ab. makoniensis Strand, 1914, l.c.: 134.

Larinopoda lircaea ab. simekoa Strand, 1914, l.c.: 134.

Larinopoda lircaea ab. bibundica Strand, 1914, l.c.: 134.

Larinopoda lycaenoides Butler, see lircaea.

Larinopoda soyauxii (Dewitz), see tera.

\*Larinopoda tera (Hewitson), 1873. Fig. Aurivillius in Seitz, 1918. soyauxii Dewitz, 1879.

Larinopoda varipes Kirby, see eurema.

## Genus FALCUNA Stempffer & Bennett

Falcuna Stempffer & Bennett, 1963, Bull. Br. Mus. nat. Hist. (Ent.) 13: 174. Type-species: Liptena libyssa Hewitson, 1866, by original designation.
Liptena Hewitson (pro parte); Aurivillius, 1898: 275; 1918: 331.

Eyes smooth; palpi rather long, extending beyond the frons, clothed with adpressed scales, the second segment laterally compressed, the third slender, blunt-pointed; antennae short, white-ringed, with distinct subcylindrical club; legs ringed with yellow, the fore tarsi of the 3 not articulated.

Wing venation (Text-fig. 243). Only differs from that of Liptena undularis (type-species of the genus Liptena) in that veins 3 and 4 of the hind wing are separate at their points of origin whereas in undularis they arise from a short common stalk. However, this is not a valid generic character, for it also occurs in fatima and submacula which are true Liptenas.

Male genitalia (Text-fig. 37). Uncus subtriangular, shield-shaped, pointed caudad, almost completely separated from the tegumen to which it is only attached by two slender ligaments running from the centre of its ventral margin to the lateral margins of the tegumen. Subunci heavily chitinized, fused together along their inner margins distally, the distal margin strongly serrate. Tegumen large. Vinculum broad, the saccus little developed. Valves oblong, apically finger-like. Penis long, subcylindrical, heavily curved. Uncus densely, apex of valves lightly, pilose.

Fusion of the subunci is not known to occur in any other genus of African Lycaenidae. Its occurrence in every species of *Falcuna*, on the other hand, justifies the erection of the genus. All the species of the genus are rather uniform, besides, in external appearance.

## LIST OF SPECIES OF Falcuna

- \*Falcuna campimus campimus (Holland), 1890. Fig. Aurivillius in Seitz, 1918; genitalia, Stempffer & Bennett, 1963: 191, figs.
- \*Falcuna campimus dilatata (Schultze), 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910–11, 1:1177. Fig. Stempffer & Bennett, 1963: 192, figs.

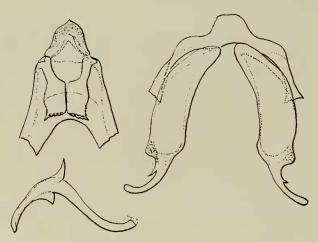


Fig. 37. Falcuna libyssa libyssa (Hewitson), of genitalia.

- \*Falcuna dorothea Stempffer & Bennett, 1963: 190, figs.
- \*Falcuna hollandi (Aurivillius), 1895. Fig. Aurivillius in Seitz, 1918 : genitalia, Stempffer & Bennett, 1963 : 185.
- \*Falcuna hollandi nigricans Stempffer & Bennett, 1963: 186, figs.
- \*Falcuna hollandi suffusa Stempffer & Bennett, 1963: 184, figs.
- \*Falcuna iturina Stempffer & Bennett, 1963: 187, figs.
- \*Falcuna kasai Stempffer & Bennett, 1963: 182, figs.
- \*Falcuna lacteata Stempffer & Bennett, 1963: 180, figs.
- \*Falcuna leonensis Stempffer & Bennett, 1963: 174, figs.
- \*Falcuna libyssa libyssa (Hewitson), 1866. Fig. genitalia, Stempffer & Bennett, 1963: 176.
- \*Falcuna libyssa angolensis Stempffer & Bennett, 1963: 177, figs.
- \*Falcuna libyssa cameroonica Stempffer & Bennett, 1963: 176, figs.
- Falcuna libyssa confluens (Grünberg), see orientalis Bethune Baker.
- Falcuna libyssa latemarginata Schultze, see margarita Suffert.
- \*Falcuna lybia (Staudinger), 1891. Fig. Aurivillius in Seitz, 1918. Fig. genitalia, Stempffer & Bennett, 1963: 193.
- \*Falcuna margarita (Suffert), 1904. Fig. Stempffer & Bennett, 1963: 181. libyssa latemarginata Schultze, 1916.

Falcuna melandeta (Holland), 1893.

- \*Falcuna orientalis orientalis (Bethune Baker), 1906. Fig. Aurivillius in Seitz, 1918. Fig. genitalia, Stempffer & Bennett, 1963: 183, figs. libyssa confluens Grünberg, 1908.
- \*Falcuna orientalis bwamba Stempffer & Bennett, 1963: 183, figs.
- \*Falcuna overlaeti Stempffer & Bennett, 1963: 190, figs.
- \*Falcuna reducta Stempffer & Bennett, 1963: 189, figs.
- \*Falcuna semliki Stempffer & Bennett, 1963: 188, figs.
- \*Falcuna synesia synesia (Hulstaert), 1924, Revue zool. afr. 12:118. Fig. Stempffer & Bennett, 1963:178, figs.
- \*Falcuna synesia f. landana Stempffer & Bennett, 1963: 178.
- \*Falcuna synesia fusca Stempffer & Bennett, 1963, : 179, figs.
- \*Falcuna synesia gabonensis Stempffer & Bennett, 1963: 178, figs.

## Genus LIPTENA Westwood

- Liptena Westwood, 1851, in Westwood, Doubleday & Hewitson, Gen. Diurn. Lep. 2: plate 77; Aurivillius, 1898: 273; 1918: 329. Type-species: Liptena undularis Hewitson, 1866, designated 1959, in Opinion 566, Int. Comm. Zool. Nomenclature.
- Parapontia Röber, 1892, in Staudinger & Schatz, Exot. Schmett. 2: 280. Type-species: Liptena undularis Hewitson, 1866.
- Leucolepis Karsch, 1893, Berl. ent. Z. 38: 216. Type-species: Teriomima decipiens Kirby, 1890.
- Pseudoliptena (partim) Stempffer, 1946, Revue fr. Ent. 13:8. Type-species Pseudoliptena bitje Stempffer, l.c.; artefact. See Hemming, 1963, Entomologist 96:292.

Eyes smooth, palpi reaching well beyond frons, 2nd joint long and laterally compressed, bearing adpressed scales, 3rd joint rather long, acuminate; antennae of moderate length, white ringed and with a gradually swollen club flattened apically; fore tarsi of 3 not segmented, delicately spinose beneath.

Wing-venation. Wing venation is not uniform throughout the genus. In undularis (Text fig. 244) vein 7 on the fore wing ends on the outer margin slightly below the apex, veins 3 and 4 on the hind wing arise from a short common stalk. This is the position in a number of species, e.g. xanthostola, evanescens, flavicans, rochei, undina, fulvicans, eukrines, homeyeri despecta, modesta etc. In Liptena decipiens (Text-fig. 245) vein 7 on the fore-wing similarly ends below the apex, but veins 3 and 4 on the hind wing arise from a common point at the lower angle of the cell, which is the condition also in alluaudi, tulliana, tullia, o-rubrum, rubromaculata etc. The venation of albomacula, ideoides, and gordoni is identical with that of Falcuna. In fatima and submacula vein 7 of the fore wing terminates at the apex.

However, the systematic divisions which could be established on the basis of these slight differences in wing venation in no way correspond with those based on the characters of the 3 genitalia, which in my opinion are far more important and significant. Neither do they correspond with the divisions which could more easily be made on such obvious characters as colour, e.g. black and white, yellow, brown or black species. For this reason I consider Karsch's *Leucolepis* (type *decipiens*) to be merely a synonym of *Liptena*.

Male genitalia (Text-fig. 38, Liptena undularis). Uncus crescentic with a shallow depression on the posterior distal margin; subunci long, curved and swollen midway; tegumen oval; vinculum rather broad and produced to form a very long spatulate saccus, directed caudad and

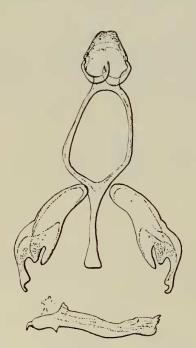


Fig. 38. Liptena (Liptena) undularis (Hewitson), & genitalia.

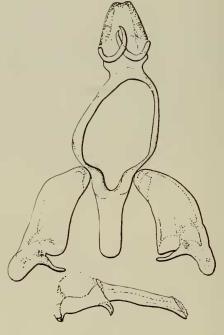


Fig. 39. Liptena (Liptena) homeyeri Dewitz, 3 genitalia.

bearing at its extremity a tuft of large scales; lower fultura small, sheathing the base of the penis; valves rather long, the two processes widely separate apically, the upper one much the longer and ending in a small hook; penis elongate, subcylindric, only slightly curved; uncus and distal portion of valves pilose.

Not all the species usually placed in *Liptena* conform to a uniform pattern of male genitalia. One can tentatively establish groups and subgroups of species based more or less on these organs:—

A. Dorsum (of genitalia) and valves as in undularis :—

a. penis elongate, subcylindrical:—
undularis, homeyeri (Text-fig. 39), ferrymani (Text-fig. 40), eukrines,
subvariegata, batesana.

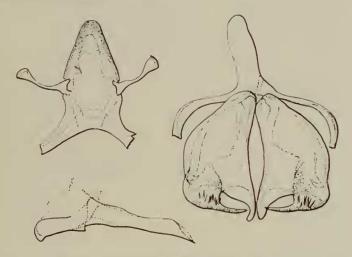


Fig. 40. Liptena (Liptena) ferrymani (Smith & Kirby), o genitalia.

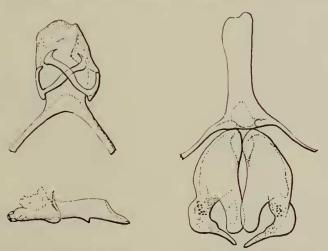


Fig. 41. Liptena (Liptena) fatima (Kirby), & genitalia.

- b. penis short, stout, sometimes with a tooth on the lower lip:—
  fatima (Text-fig. 41), decipiens (Text-fig. 42), alluaudi, submacula, xanthostola, undina, evanescens.
- c. penis long, the tip more or less delated:—
  albomaculata (Text-fig. 43), o-rubrum (Text-fig. 44), nigromarginata, opaca, septistrigata.

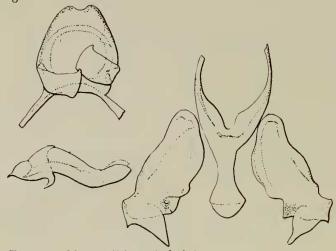


Fig. 42. Liptena (Liptena) decipiens (Kirby), & genitalia.

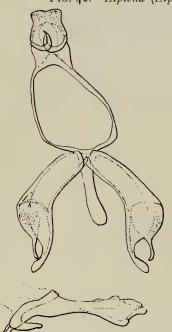


Fig. 43. Liptena (Liptena) albomacula Hawker Smith, 3 genitalia.

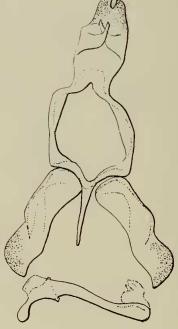


FIG. 44. Liptena (Liptena) o-rubrum (Holland), 3 genitalia.

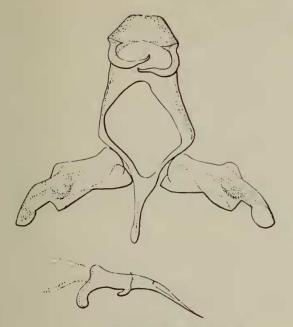


Fig. 45. Liptena (Liptena) modesta (Kirby), 3 genitalia.

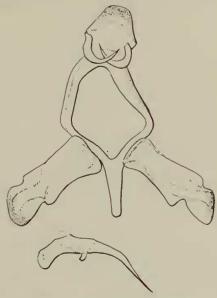


Fig. 46. Liptena (Liptena) helena (Druce), & genitalia.

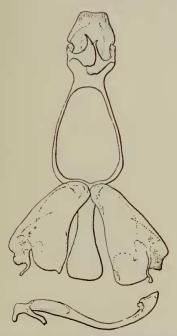


Fig. 47. Liptena (Liptena) flavicans (Smith & Kirby), & genitalia.

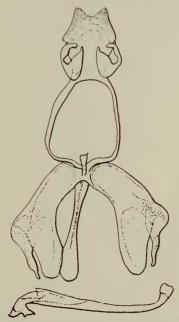


Fig. 48. Liptena (Liptena) similis (Kirby), & genitalia.

- d. penis ending in a tapering point:—
  modesta, (Text-fig. 45), helena (Text-fig. 46), despecta, catalina, hapale,
  augusta, rubromacula.
- e. penis very long, subcylindrical, strongly curved :— flavicans (Text-fig. 47), similis (Text-fig. 48), rochei, fulvicans, durbania.
- B. Uncus subrectangular, the distal margin deeply excised; two pairs of subunci; penis long, feebly curved:—

  tullia (Text-fig. 49), tulliana (Text-fig. 50).
- C. Dorsal structures greatly reduced, feebly sclerotized, subtriangular and rounded above, attached directly to the vinculum; no subunci; valves oblong, the lower processes united basally; penis long, tapering gradually, apically bulbous and with a stout hook. Dorsum smooth, the ends of the valves moderately pilose:

ideoides (Text-fig. 51, ventral aspect), gordoni (Text-fig. 52, lateral view), infima, otlauga, ferruginea.

A general revision of the genus *Liptena* auctorum would be most valuable; I have not been able to undertake it myself as it has not yet been possible for me to examine all the species so far described, which alone would have permitted me to draw

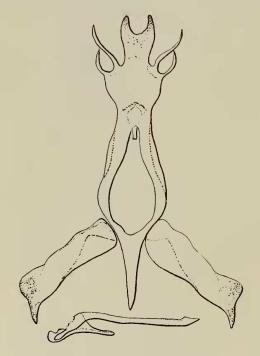


Fig. 49. Liptena (Liptena) tullia (Staudinger), & genitalia.



Fig. 50. Liptena (Liptena) tulliana Smith, 3 genitalia.

definite conclusions. However, one can say at once that the species included in groups B and C ought to be excluded from the genus *Liptena*, the genitalia being of a type plainly different from those of *undularis*.

# LIST OF SPECIES OF Liptena (Liptena)

\*Liptena albicans Cator, 1904. Fig. Aurivillius in Seitz, 1914-25.

\*Liptena albomacula Hawker Smith, 1933, Stylops 2:7.

Liptena albula (H. H. Druce), see simplicia.

\*Liptena alluaudi Mabille, 1890. Fig. genitalia, Stempsfer, 1957, Bull. Inst. fr. Afr. noire 19: 213.

Liptena amabilis Schultze, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910-11, 1: 1181, fig.

\*Liptena augusta Suffert, 1904. Fig. H. H. Druce, 1910; genitalia, Stempffer, 1957 Bull. Inst. fr. Afr. noire 19: 214.

Liptena bakeriana (Cator), see gordoni.

Liptena bassae Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 390. subpunctata Bethune Baker, 1906 (nom. praeoc).

Liptena batesana Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 390.

Liptena bolivari Kheil, 1905.

\*Liptena catalina Smith & Kirby, 1887.

Liptena citronensis Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 389. Liptena confusa Aurivillius, 1898.

\*Liptena congoana Hawker Smith, 1933, Stylops 2: 9.

Liptena daemon H. H. Druce, see o-rubrum.

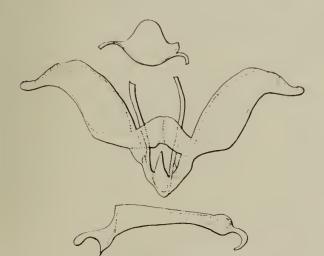


Fig. 51. Liptena (Liptena) ideoides Dewitz, & genitalia.

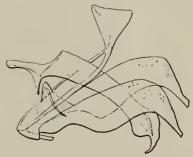


Fig. 52. Liptena (Liptena) gordoni (Druce), & genitalia of type.

- Liptena decempuncata Schultze, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910-11, 1: 1187.
- \*Liptena decipiens (Kirby), 1890, (Sept.). Fig. Smith & Kirby, 1891. leucostola Holland, 1890, (Dec.).
- \*Liptena decipiens cameroona Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 389.
- \*Liptena despecta (Holland), 1890. Fig. Aurivillius in Seitz, 1914–25. modestissima Rebel, 1914.
- \*Liptena durbania Bethune Baker, 1915, Ann. Mag. nat. Hist. (9) 16: 189. rectifascia Hawker Smith, 1933.

Liptena eketi Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 391.

Liptena erycinoides (Smith & Kirby), see helena.

Liptena eukrinaria Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 391.

\*Liptena eukrines H. H. Druce, 1905.

Liptena eukrines obsoleta Dufrane, 1953, Bull. Annls Soc. R. ent. Belg. 89: 49.

Liptena eukrinoides Talbot, 1937, Trans. R. ent. Soc. Lond. 86: 64, fig.

- \*Liptena evanescens (Kirby), 1887. Fig. Smith & Kirby, 1887. xanthis (Holland), 1890.
- \*Liptena fatima (Kirby), 1890. Fig. Smith & Kirby, 1891.
- \*Liptena ferruginea Schultze, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910-11, 1:1184, fig.
- \*Liptena ferrymani ferrymani (Smith & Kirby), 1891.
- \*Lipena ferrymani bigoti Stempffer, 1964: 1233.
- \*Liptena flavicans flavicans (Smith & Kirby), 1891.
- \*Liptena flavicans aequatorialis Stempffer, 1956: 8, fig.
- \*Liptena flavicans katera Stempffer, 1956: 8, fig.
- \*Liptena flavicans oniens Talbot, 1935, Entomologist's mon. Mag. 71: 72, fig.
- \*Liptena flavicans praeusta Schultze, 1916, Arch. Naturgesch. 82, A3: 38.
- \*Liptena fulvicans Hawker Smith, 1933, Stylops 2:8.

Liptena girthii Dewitz, see ideoides.

- \*Liptena gordoni (H. H. Druce), 1903. Fig. Lathy, 1903. bakeriana (Cator), 1904.
- \*Lipena hapale Talbot, 1935, Entomologist's mon. Mag. 71: 72, fig.
- \*Liptena helena (H. H. Druce), 1888. Fig. Smith & Kirby, 1890. erycinoides (Smith & Kirby), 1890.
- \*Liptena homeyeri Dewitz, 1884.
- \*Liptena hulstaerti Hawker Smith, 1926, Revue zool. afr. 14: 239.
- \*Liptena ideoides Dewitz, 1886. girthii Dewitz, 1886.
- \*Liptena infima (Smith & Kirby), 1890.

Liptena inframacula Hawker Smith, 1933, Stylops 2:7.

Liptena intermedia Grünberg, 1910.

Liptena jacksoni Stempffer, see nigromarginata.

\*Liptena kelle Stempffer, 1964: 1231, fig.

Liptena latruncularia (Holland), see modesta.

Liptena leucostola (Holland), see decipiens.

\*Liptena modesta (Kirby), 1890 (Sept.). Fig. Smith & Kirby, 1892. latruncularia (Holland), 1890 (December).

Liptena modestissima Rebel, see despecta.

Liptena mwagensis Dufrane, 1953, Bull. Annls Soc. R. ent. Belg. 89: 49.

\*Liptena nigromarginata Stempffer, 1961: 43.

jacksoni Stempffer, 1953 (nom. praeoc.).

Liptena occidentalis Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 390.

Liptena ochrea Hawker Smith, 1943, Stylops 2:8.

Liptena olombo (Holland), 1890.

Liptena opaca opaca (Kirby), 1890. Fig. Smith & Kirby, 1892.

\*Liptena opaca immaculata Grünberg, 1910.

\*Liptena o-rubrum o-rubrum (Holland), 1890. Fig. Aurivillius in Seitz, 1914-25.

daemon H. H. Druce, 1910.

\*Liptena o-rubrum teroana Talbot, 1935, Entomologist's mon. Mag. 71: 72.

Liptena o-rubrum tripunctata (Smith & Kirby), 1894.

\*Liptena otlauga (Smith & Kirby), 1890.

Liptena perobscura H. H. Druce, 1910.

Liptena praestans praestans (Smith & Kirby), 1910.

Liptena praestans congoensis Schultze, 1823, Ergebn. 2te D. Zent. Afr. Exped. 1910-11, 1:1181.

Liptena praestans kamitugensis Dufrane, 1945, Bull. Annls Soc. R. ent. Belg. 81: 118.

Liptena rectifascia Hawker Smith, see durbania.

\*Liptena rochei Stempffer, 1951, Bull. Soc. ent. Fr. 1951: 66, fig.

\*Liptena rubromacula rubromacula Hawker Smith, 1933, Stylops 2:9.

\*Liptena rubromacula jacksoni Carpenter, 1934, Proc. R. ent. Soc. Lond. 9: 12.

Liptena sauberi Schultze, 1912. Fig. Aurivillius in Seitz, 1914–25.

Liptena semilimbata (Mabille), see simplicia.

\*Liptena septistrigata (Bethune Baker), 1903.

\*Liptena similis (Kirby), 1890. Fig. Smith & Kirby, 1892.

\*Liptena simplicia Möschler, 1888.

albula (H. H. Druce), 1888 (October); semilimbata (Mabille), 1890.

\*Liptena submacula Lathy, 1903.

Liptena subpunctata Bethune Baker, see bassae.

Liptena subsuffusa Hawker Smith, 1933, Stylops 2:7.

Liptena subundularis Staudinger, 1891. Fig. Smith & Kirby, 1892.

Liptena subvariegata subvariegata Smith & Kirby, 1890.

Liptena subvariegata aliquantum H. H. Druce, 1910.

Liptena tricolora Bethune Baker, 1915, Ann. Mag. nat. Hist (8) 16: 188.

\*Liptena tullia (Staudinger), 1891. Fig. Aurivillius, 1898.

\*Liptena tulliana Smith, 1901.

Liptena turbata (Kirby), 1890. Fig. Smith & Kirby, 1891.

\*Liptena undina (Smith & Kirby), 1894.

\*Liptena undularis Hewitson, 1866.

Liptena xantha (Smith), see xanthostola.

Liptena xanthis (Holland), see evanescens.

\*Liptena xanthostola (Holland), 1890. Fig. Aurivillius in Seitz, 1914–25. xantha (Smith), 1901.

Liptena xanthostola coomassiensis Hawker Smith, 1933, Stylops 2:8.

Liptena yukadumae Schultze, 1916, Arch. Naturgesch. 82 A 3:38. Fig. Schultze, 1923.

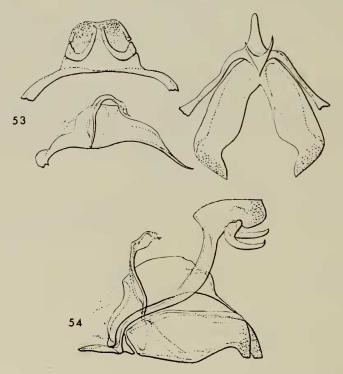
## LIPTENA (TETRARHANIS) Karsch

Tetrarhanis Karsch, 1893, Berl. ent. Z. 38: 217. Type-species: Liptena ilma Hewitson, 1873, by monotypy.

Lectiles Birket Smith, 1960, Bull. inst. fr. Afr. noire. (A) 22: 1968. Type-species Lectiles collitorum B. Smith by monotypy.

Eyes, palpi, antennae and legs as in other species of Liptena.

Wing venation (Text-fig. 246). On the fore wings vein 7 ends at the apex. On the hind



Figs 53, 54. Liptena (Tetrarhanis) ilma Hewitson, & genitalia.

wings veins 3 and 4 are narrowly separate at their points of origin, and vein 7 arises from the

upper angle of the cell and not from its anterior margin as in other species of Liptena.

Male genitalia (Text-fig. 53, ventral aspect; Text-fig. 54, lateral aspect). Uncus with posterior margin slightly depressed; subunci rather long, evenly curved; tegumen large; vinculum rather broad, with a short saccus; lower fultura strongly developed, surrounding the middle of the penis; valves oblong; penis of very characteristic form, narrow at the base, swollen and massive in the middle, ending in a slender point. Uncus and apices of valves pilose.

The male genitalia in *Tetrarhanis* are of a uniform type, except that in *L. schoutedeni* the tip of the penis is bifid. There is agreement here between divisions founded on wing venation and genitalia. I consider *Tetrarhanis* therefore to be a valid subgenus.

# LIST OF SPECIES OF Liptena (Tetrarhanis)

Liptena (Tetrarhanis) collitorum (Birket Smith), (as Lectiles sp.), see schoutedeni.

- \*Liptena (Tetrarhanis) diversa diversa Bethune Baker, 1904. Genitalia, Stempffer, 1964: 1236, fig.
- \*Liptena (Tetrarhanis) diversa ilala Riley, 1929, Trans. ent. Soc. Lond. 1929: 493.

\*Liptena (Tetrarhanis) etoumbi Stempffer, 1964: 1236, fig.

- \*Liptena (Tetrarhanis) ilma ilma Hewitson, 1873. Genitalia, Berger, 1954, Annls Mus. r. Congo belge, Zool. 1.
- \*Liptena (Tetrarhanis) ilma daltoni Poulton, 1929, Trans. ent. Soc. Lond. 1929: 493, fig.
- \*Liptena (Tetrarhanis) ilma lathyi Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1: 82, fig.
- \*Liptena (Tetrarhanis) ilma ugandae Stempffer, 1964: 1234, fig. Liptena (Tetrarhanis) laminiter Clench, 1965, Butt. Liberia: 30, figs.
- \*Liptena (Tetrarhanis) nubifera H. H. Druce, 1910. Genitalia, Stempffer, 1961: 43, fig.
- \*Liptena (Tetrarhanis) ogojae Stempffer, 1961: 44, fig.

\*Liptena (Tetrarhanis) onitshae Stempffer, 1962: 1146, fig.

- \*Liptena (Tetrarhanis) rougeoti Stempffer, 1954, Bull. Soc. ent. Fr. 1954: 92, fig.
- \*Liptena (Tetrarhanis) schoutedeni Berger, 1954, Annls Mus. R. Congo belge, Zool. 1, 1954: 308, fig.

Lectiles collitorum Birket Smith, 1960.

\*Liptena (Tetrarhanis) simplex Aurivillius, 1895. Genitalia, Stempffer, 1964: 1235.

\*Liptena (Tetrarhanis) souanke Stempffer, 1962: 1148, fig.

\*Liptena (Tetrarhanis) stempfferi stempfferi Berger, 1954, Annls Mus. R. Congo belge, Zool. 1, 1954: 307, fig.

\*Liptena (Tetrarhanis) stempfferi kigezi Stempffer, 1956: 8, fig.

\*Liptena (Tetrarhanis) symplocus Clench, 1965, Butt. Liberia: 302, figs.

## Genus MICROPENTILA Aurivillius

Micropentila Aurivillius, 1895, Ent. Tidsk. 16: 202; 1898: 281; 1920: 339. Type-species: Liptena adelgitha Hewitson, 1874, by original designation.

Eyes glabrous; palpi of medium length, shorter than in Liptena, the second segment clothed below with stiff hair or hair-scles; third segment subconical; antennae of medium length reaching beyond the tip of the discal cell, the club oval, short, sharply defined; legs black, ringed with white, the  $\beta$  fore tarsi short, unsegmented, finely spinose beneath.

Wing shape. Fore wing costa rather strongly curved, the outer margin convex; hind wing oval with a rather well marked anal angle.

Venation (Text-fig. 247). Fore wing with 12 veins; the stem of 7 + 8 + 9 arises from the upper angle of the cell or very shortly before it; hind wing, 3 and 4 free from lower angle of cell, or very shortly stalked, 7 a little before the upper angle of cell.

Male genitalia (Text-fig. 55). Uncus divided into two subtriangular processes with rounded extremities separated by a deep indentation; subunci long slender and curved; tegumen large; vinculum with a slender saccus; valves oblong and tapering evenly to a blunt apex; penis basally bulbous and sheathed by a small inferior fultura, then cylindrical, dilated and notched at the tip; uncus clothed in long hair, only the ends of the valves slightly pilose.

In collaboration with N. H. Bennett of the British Museum (N.H.) I have recently published a revision of the genus *Micropentila* (1965: 397–434, 31 figs, 4 plates), in which the genitalia of almost all the known species are figured, and to which reference should be made. Not all the species have genitalia like those of *M. adelgitha*. As a rule the uncus is not so deeply divided, the saccus is sometimes broadly spatulate, and the penis in particular shows many variations. According to the characters of the penis we have divided the genus into nine sections:—

- Section A Penis bulbous at base, excised apically:—
  adelgitha, subplagata, souanke
- Section B Penis long and bent; valves as in some Liptena:—fulvula
- Section C Penis long, cylindrical, bent at right angles near the base, its apex rolled back:—

  adelgunda (Text-fig. 56), bitjeana, dorothea, gabunica
- Section D Penis rather long, ending in a fine point:—
  brunnea (Text-fig. 57), victoriae, katerae, flavopunctata, jacksoni,
  bakotae, nigeriana, mpigi, fontainei, fuscula, ogojae, kelleana.

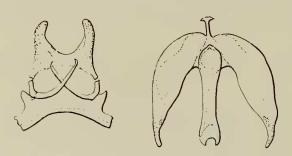


Fig. 55. Micropentila adelgitha (Hewitson), & genitalia.

Section E Penis shorter and stouter than in Section D :— alberta, mabangi

Section F Penis long, slender, strongly curved :— cingulum, ugandae

Section G Penis long, only gently curved:—
sankuru

Section H Penis short, its extremity bulbous and bifid:— katangana, cherereti

Section I Penis short, its tip broadly concave :— bunyoro.

In spite of the very uniform external facies throughout, the genus *Micropentila*, it would seem on this showing, has a rather mixed phylogeny.

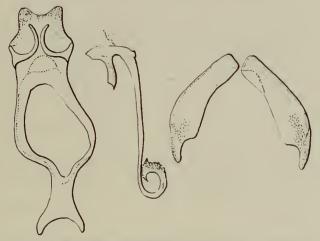


Fig. 56. Micropentila adelgunda (Staudinger), o genitalia.

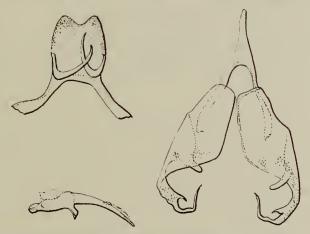


Fig. 57. Micropentila brunnea (Kirby), o genitalia.

# LIST OF SPECIES OF Micropentila

- \*Micropentila adelgitha (Hewitson), 1874. Fig. Smith & Kirby, 1892. Fig. genitalia, Stempffer & Bennett, 1965: 402.

  moneta (Mabille), 1890.
- \*Micropentila adelgunda (Staudinger), 1891. Fig. Smith & Kirby, 1892. Fig. genitalia, Stempffer & Bennett, 1965: 406.
- \*Micropentila alberta (Staudinger), 1891. Fig. Smith & Kirby, 1892. Fig. genitalia, Stempffer & Bennett, 1965: 424.
- \*Micropentila bakotae Stempffer & Bennett, 1965: 416, fig. and fig. genitalia.
- \*Micropentila bitjeana Stempffer & Bennet, 1965: 407, fig. and fig. genitalia.
- \*Micropentila brunnea (Kirby), 1887. Fig. Smith & Kirby, 1888. Fig. genitalia, Stempffer & Bennett, 1965: 410.

Micropentila brunnea centralis Bennett, 1966, Entomologist 99: 186, fig.

- \*Micropentila bunyoro Stempffer & Bennett, 1965: 432, fig. and fig. genitalia. Micropentila catocata Strand, 1914. Arch. Naturgesch. 80 A 2: 155.
- \*Micropentila cherereti Stempffer & Bennett, 1965: 431, fig. and fig. genitalia.
- \*Micropentila cingulum H. H. Druce, 1910. Fig. and fig. genitalia, Stempffer & Bennett, 1965: 425.
- \*Micropentila dorothea Bethune Baker, 1903. Fig. and fig. genitalia, Stempffer & Bennett, 1965: 407.
- \*Micropentila flavopunctata Stempffer & Bennett, 1965:414, fig. and fig. genitalia.
- \*Micropentila fontainei Stempffer & Bennett, 1965: 419, fig. and fig. genitalia.
- \*Micropentila fulvula Hawker Smith, 1933, Stylops 2: 9. Fig. and fig. genitalia, Stempffer & Bennett, 1965: 405.
- \*Micropentila fuscula Grose Smith, 1898. Fig. and fig. genitalia, Stempffer & Bennett, 1965: 420.
- \*Micropentila gabunica Stempffer & Bennett, 1965: 409, fig. and fig. genitalia.
- \*Micropentila jacksoni Talbot, 1937, Trans. R. ent. Soc. Lond. 86: 61. Fig. and fig. genitalia, Stempffer & Bennett, 1965: 415.
- \*Micropentila katangana Stempffer & Bennett, 1965: 430, fig. and fig. genitalia.
- \*Micropentila katerae Stempffer & Bennett, 1965: 412, fig. and fig. genitalia.
- \*Micropentila kelleana Stempffer & Bennett, 1965: 422, fig. and fig. genitalia.
- \*Micropentila mabangi Bethune Baker, 1904. Fig. and fig. genitalia, Stempffer & Bennett, 1965: 424.

Micropentila moneta (Mabille), see adelgitha.

- \*Micropentila mpigi Stempffer & Bennett, 1965: 418, fig. and fig. genitalia.
- \*Micropentila nigeriana Stempffer & Bennett, 1965: 417, fig. and fig. genitalia.
- \*Micropentila ogojae Stempffer & Bennett, 1965: 421, fig. and fig. genitalia.
- \*Micropentila sankuru Stempffer & Bennett, 1965: 428, fig. and fig. genitalia.
- \*Micropentila souanke Stempffer & Bennett, 1965: 403, fig. and fig. genitalia.
- \*Micropentila subplagata Bethune Baker, 1915, Ann. Mag. nat. Hist. (8) 16: 189. Fig. genitalia, Stempffer & Bennett, 1965: 402.

Micropentila triangularis Aurivillius, 1895. Fig. Aurivillius in Seitz, 1914-25.

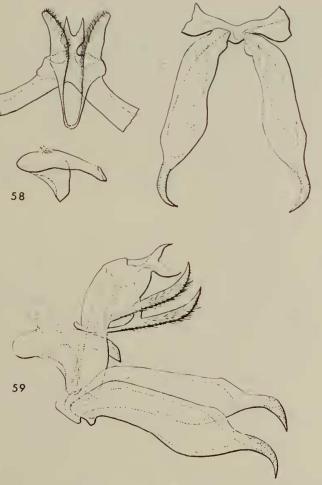
\*Micropentila ugandae Hawker Smith, 1933, Stylops 2:10. Fig. and fig. genitalia, Stempffer & Bennett, 1965:427.

\*Micropentila victoriae Stempffer & Bennett, 1965: 411, fig. and fig. genitalia.

### Genus ERESINOPSIDES Strand

Eresinopsides Strand, 1911, Arch. Naturgesch. 77: 193; Aurivillius, 1920: 340. Typespecies: Eresinopsides bichroma Strand, by original designation.

Eyes smooth; palpi long, projecting well beyond the frons, second segment oval, laterally compressed, clothed with large scales, third segment long, slender, cylindrical; antennae moderately long, delicate, with a well differentiated fusiform club; fore leg of  $\delta$  with tibia slightly shorter than femur, tarsus unsegmented, bearing fine hairs below; mid and hind legs with tibiae a little shorter than femora, slightly dilated.



Figs 58-59. Eresinopsides bichroma Strand, & genitalia.

Wing venation (Text-fig. 248). Fore wing with 11 veins only; 6 and 7 arise from a common stem from the upper angle of the cell; hind wing: 7 stalked on 6.

Male genitalia (Text-fig. 58, ventral view; Text-fig. 59, side view). Uncus bifid, shaped like a crescent with pointed horns; tegumen consists of a small, narrow dorsal strip; two large lobes, which probably represent the subunci, are attached to the tegumen by their middle portions, on either side of the uncus, and their hinder parts are united by a membrane, the whole of their surface being shortly pilose and the internal edge bearing long spines; vinculum wide, without saccus; lower fultura closely sheathing the penis; valves long and narrow, their distal portion falcate with a pointed apex; penis oblong, obliquely truncated at apex; uncus and apices of valves shortly pilose.

The genitalia of *E. bichroma* have a very peculiar structure; they in no way resemble those of species of *Eresina*, but are somewhat analogous to those of *Baliochila minima* Hawker-Smith.

# LIST OF SPECIES OF Eresinopsides

\*Eresinopsides bichroma bichroma Strand, 1911.
staphyla (Hulstaert), 1924 (as Pseuderesia sp.).
Eresinopsides bichroma jefferyi Stempffer, 1950, Revue fr. Ent. 17: 135.
Eresinopsides staphyla (Hulstaert), see bichroma.

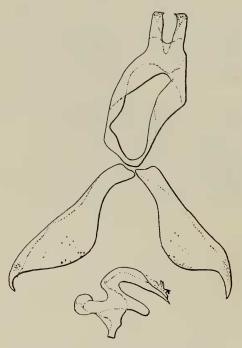


Fig. 60. Eresina corynetes (Kirby & Smith), & genitalia of type.

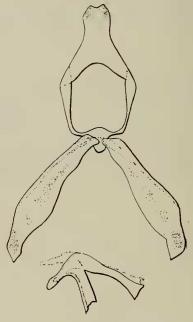


Fig. 61. Eresina toroensis Joycey & Talbot, & genitalia.

### Genus **ERESINA** Aurivillius

Eresina Aurivillius, 1898, Rhopalocera Aethiopica: 282; 1920: 341. Type-species: Durbania corvnetes Smith & Kirby, by original designation.

Eyes smooth; palpi long, extending beyond the frons, second segment bearing scales and bristly hairs, third segment rather long, subcylindrical; antennae of moderate length, barely half the length of costa, the club distinct, oval, flattened; legs short, scaly, anterior tarsi of  $\eth$  unsegmented, bearing delicate spines below; hind tibiae gently curving, thickened and somewhat compressed.

Wing shape. The hind margin of the hind wings is slightly scalloped.

Wing venation (Text-fig. 249 corynetes). Very characteristic: on the fore wing vein 6 arises from the upper angle of the cell, 7 arises from 6 and ends at the apex, 8 stalked on 7, 9 absent, 10 also branching from 6 about midway between the upper angle of cell and the origin of 7, 11 separate from the anterior margin of cell.

Wing venation in Eresina is not entirely uniform. In some species vein 10 arises nearer to the upper angle of the cell and sometimes actually from this angle, but even in these species the 3 genitalia are quite similar to those of the other species.

Male genitalia (Text-fig. 60). Uncus composed of two subrectangular processes, separated by a deep division, each bearing a sort of triangular crest at an oblique angle; no subunci; tegumen rather big; vinculum broad dorsally; inferior fultura with a short peduncle fused to the extreme base of the valves and tightly enclosing base of the penis; valves oblong, falcate at the apex (slightly asymmetric in the holotype of corynetes); penis doubly curved (S-shaped) with a very slender tip and distally widely open dorsally, vesica bearing two short spines and other smaller ones. Apex of uncus covered with rather long fine hair, extremity of the values lightly pilose and their ventral margins shortly but densely pilose.

In most species of *Eresina* the male armature is as in *corynetes*, i.e. with a deeply divided uncus and an S-shaped penis. In *toroensis* (Text-fig. 61) and *conradti* the uncus is only shallowly divided; in *crola*, *fusca* and *pseudofusca* the penis is simply curved and in *conradti* its extremity is bifid.

### LIST OF SPECIES OF Eresina

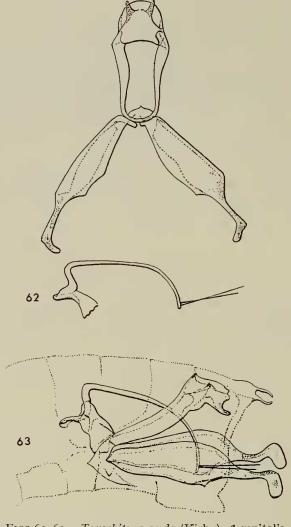
- \*Eresina bergeri Stempffer, 1956: 12, figs.
- \*Eresina bilinea Talbot, 1935, Entomologist's mon. Mag. 71: 73, figs.
- \*Eresina conradti Stempffer, 1956: 27, figs.
- \*Eresina corynetes (Smith & Kirby), 1890.
- \*Eresina crola Talbot, 1935, Entomologist's mon. Mag. 71: 73, figs.
- \*Eresina fontainei Stempffer, 1956: 18, figs.
- \*Eresina fusca (Cator), 1904. Fig. Stempffer, 1956.
- \*Eresina jacksoni Stempffer, 1961: 48, figs.
- \*Eresina katangana Stempffer, 1956: 15, figs.
- \*Eresina katera Stempffer, 1962: 1155, figs.
- \*Eresina likouala Stempffer, 1962: 1149, figs.
- \*Eresina maesseni Stempffer, 1956: 14, figs.
- \*Eresina masaka Stempffer, 1962: 1152, figs.
- \*Eresina pseudofusca Stempffer, 1961: 47, figs.
- \*Eresina rougeoti Stempffer, 1956: 19, figs.

- \*Eresina saundersi Stempffer, 1956: 17, figs.
- \*Eresina theodori Stempffer, 1956: 21, figs.
- \*Eresina toroensis Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1:83, figs.

# Genus TOXOCHITONA Stempffer

Toxochitona Stempffer, 1956, Annls Mus. R. Congo belge 49: 28. Type-species: Durbania gerda Kirby, 1890, by original designation.

Eresina Aurivillius, 1898: 283; 1920: 341 (pro parte).



Figs 62-63. Toxochitona gerda (Kirby), & genitalia.

Eyes, palpi, antennae and legs as in Eresina.

Wing venation (Text-fig. 250). Differs little from that of Eresina: on the fore wings vein 10

arises from the anterior margin of the cell very slightly before the upper angle.

Male genitalia (Text-figs 62, 63). Eighth tergite ends in a sort of small claw; uncus composed of two small finger-like lightly divergent processes; no subunci; tegumen rather large; pseudotergum (see Bayard, 1933, Bull. Soc. Fr. Microsc. 2(4): 83, 99) well developed but semi-membranous and translucent, so that it is difficult to locate its anterior margin precisely by observation under the microscope; inferior fultura pedunculate, fused to the base of the valves; valves oblong, abruptly narrowing just before the extremity which is bent at an angle like a foot; penis very specialized, strongly curved, slender and including (at its tip) two long needle-like spines.

The remarkable form of the penis in this genus suggested the generic name *Toxochitona*. The armature of *T. sankura* is very similar to that of *gerda*, differing only in the shape of the uncus and the apex of the valves. I do not know the armature of *T. vansomereni*, described on the basis of a unique female and I place it in this genus only on account of its external appearance.

#### LIST OF SPECIES OF Toxochitona

Toxochitona bitjensis (Bethune Baker), see gerda.

Toxochitona gerda gerda (Kirby), 1890. Fig. Smith & Kirby, 1892.

bitjensis (Bethune Baker), 1926.

Toxochitona gerda unicolor (Aurivillius), 1898.

\*Toxochitona sankuru Stempffer, 1961: 51, figs.

Toxochitona vansomereni (Stempffer), 1953, Annls Mus. R. Congo belge 27: 12.

## Genus ARGYROCHEILA Staudinger

Argyrocheila Staudinger, 1891, Dt. ent. Z. Iris 4: 215; Aurivillius, 1898: 284; 1920: 341. Type-species: Argyrocheila undifera Staudinger, by monotypy.

Eyes naked; palpi small, slender, scarcely reaching beyond the frons; antennae slender, short, hardly one-third the length of the costa, club distinct, short; legs annulated, black and white,  $\delta$  fore tarsus unsegmented.

Wing-shape; fore wing in undifera deeply lobed at the ends of veins 6, 5 and 4, with the biggest saliant at vein 4. In inundifera and bitje the outer margin is strongly convex but devoid of lobes.

Wing venation (Text-fig. 251). Fore wing with 12 veins; from the upper angle of the cell there arises a stem from which vein 6 branches, and this stem continues as the common stem of 7 + 8 + 9.

Male genitalia (Text-fig. 64). Uncus narrow, trapezoidal, with a slight notch in the distal edge; no subunci; tegumen much reduced; vinculum wide in its tergal portion, narrow in its sternal portion; inferior fultura fused to the base of the valves and sheathing the base of the penis; valves oblong, with two sharp teeth at the apex; penis shaped like an open S, the upper surface widely open at the base; uncus and apices of valves pilose.

The genital armature of *inundifera* (Text-fig. 65) differs from that of *undifera* in the shape of the uncus which is bifid and formed of two horn-shaped processes,

widely separated at their bases. The armature of bitje is very like that of inundifera. Although the species of Argyrocheila in their peculiar external appearance differ considerably from the species of allied genera, their male genitalia show that they are not very far from Eresina.

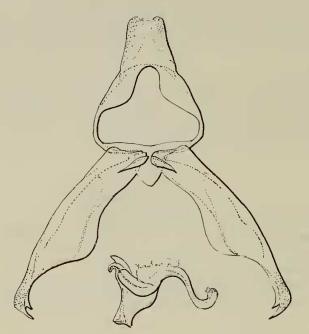


Fig. 64. Argyrocheila undifera Staudinger, & genitalia.

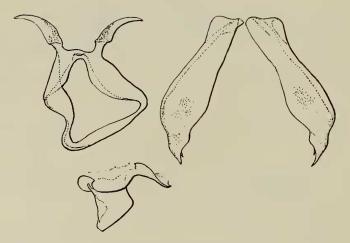


Fig. 65. Argyrocheila inundifera Hawker Smith, & genitalia.

# LIST OF SPECIES OF Argyrocheila

- \*Argyrocheila bitje Bethune Baker, 1915, Ann. Mag. nat. Hist. (8) 16: 187.
- \*Argyrocheila inundifera Hawker Smith, 1933, Stylops 2: 10.
- \*Argyrocheila undifera undifera Staudinger, 1891.
- \*Argyrocheila undifera ugandae Hawker Smith, 1933, Stylops 2: 10.

# Genus ASLAUGA Kirby

Aslauga Kirby, 1890, Ann. Mag. nat. Hist. (6) 6:261; Aurivillius, 1898:284; 1920:341 (pro parte); Bethune Baker, 1924:208. Type-species: Aslauga marginalis Kirby, 1890 (Liphyra vininga Hewitson, 1875), by selection by Bethune Baker, 1924, Trans. ent. Soc. London 1924:209.

Eyes naked; palpi divergent, obliquely erect, clothed with adpressed scales, third segment fairly long; antennae short and thick, segments not much longer than broad, club rounded;  $\Im$  fore tarsus composed of five distinct segments, with two claws on terminal segment, as in the  $\Im$ .

Wing shape: shape peculiar; fore wing slightly falcate, outer margin excised at extremity of vein 6, strongly convex at the extremities of veins 4 and 3; hind wing with anal margin concave between the ends of veins 1a and 1b, outer margin concave between 1b and 4, thus forming a blunt tooth at the end of 1b and an angle at the end of 4.

Wing venation (Text-fig. 252).

Male genitalia (Text-fig. 66). Dorsal elements small in relation to the other parts of the genitalia; uncus deeply excised at the apex; no subunci; tegumen with a median rounded protuberance on the anterior edge; vinculum very narrow, prolonged to form a strong saccus; lower fultura composed of two large triangular lobes with pointed apex; valves elliptical, their lower process short, upper process ending in a long, strong hook; penis massive, subcylindrical, tapering slightly in its external portion; vesica armed with numerous cornuti and strong spines; uncus and apices of valves pilose.

Aslauga lamborni (Text-fig. 67). The genitalia have been described by Bethune Baker somewhat inaccurately. Even if it is true that the uncus is only weakly

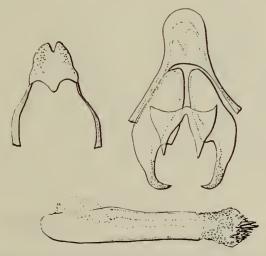


Fig. 66. Aslauga vininga (Hewitson), & genitalia.

excised, the manner of the attachment of the valves is the same as in *vininga* and the vinculum is similarly prolonged to form a stout saccus. The ends of the valves are obliquely truncate, with a serrate margin.

In aura and pandora the uncus is only weakly excised above and the penis is less elongate than in vininga and lamborni.

The genus Aslauga belongs to the sub-family Liphyrinae, sharply characterized by the five-segmented male fore tarsus, the morphology of its pupae and its larvae.

The early stages of A. vininga and A. lamborni were the subject of the remarkable observations of Dr. Lamborn (1914, Trans. ent. Soc. Lond., 1913: 446-7) and the pupae of the same species have been described by Eltringham (1922, Trans. ent. Soc. Lond. 1921: 473, pl. 12, figs 4 and 5) and by Bethune Baker (1924: 214-7, pls 17 to 24).

I give below only a brief summary of these researches. The caterpillars, seen from above, are oblong with the sides sloping downwards and outwards; the dorsal skin, which is tough and covered with rough tubercles, forms a kind of carapace beneath which the small head can be withdrawn; the ventral surface is flat. They feed on Coccidae and are cared for by ants of the genus *Crematogaster*, which erect around the caterpillars small shelters made of particles of bark and other vegetable debris. The carapace enables the caterpillars to withstand any casual attacks from the ants.

The chrysalis bears branched hairs of a complex structure; some are like the flower of a water-lily with a long pistil, others like small balls on a short stem. Eltringham calls these hairs "chitinanths".

The caterpillar of A. purpurascens Holland has been described by T. H. E. Jackson (1937, Trans. R. ent. Soc. Lond., 86: 207); it also has a kind of carapace and feeds on Membracidae.

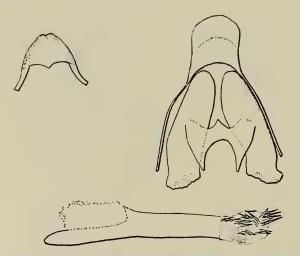


Fig. 67. Aslauga lamborni Bethune Baker, & genitalia

# LIST OF SPECIES OF Aslauga

\*Aslauga aura H. H. Druce, 1913.

Aslauga bella Bethune Baker, 1913.

Aslauga bitjensis Bethune Baker, 1924: 211.

\*Aslauga lamborni Bethune Baker, 1913.

Aslauga leonae Aurivillius, see vininga.

Aslauga marginalis Kirby, see vininga.

Aslauga marginata (Plötz), see vininga.

Aslauga modesta Schultze, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910-11, 1: 1192.

\*Aslauga pandora H. H. Druce, 1913.

Aslauga purpurascens purpurascens (Holland), 1890.

Aslauga purpurascens marginaria Talbot, 1937, Trans. ent. Soc. Lond. 86: 61, fig.

Aslauga purpurascens marshalli Butler, 1898.

Aslauga subfulvida (Holland), see vininga.

\*Aslauga vininga (Hewitson), 1875.

marginata (Plötz), 1880; marginalis Kirby, 1890; subfulvida (Holland), 1890 (as Epitola sp.); leonae Aurivillius, 1920.

### Genus PARASLAUGA Bethune Baker

Paraslauga Bethune Baker, 1924, Trans. ent. Soc. Lond. 1924: 207. Type-species: Aslauga kallimoides Schultze, by original designation.

Aslauga Kirby (partim); Aurivillius, 1920: 243.

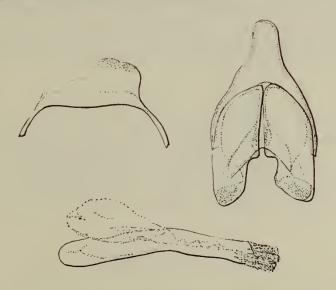


Fig. 68. Paraslauga kallimoides (Schultze), of genitalia.

Eyes smooth; palpi slightly divergent, protruding beyond the frons, second segment clothed with adpressed scales, third segment fairly long; antennae short and thick, segments hardly longer than broad, club cylindrical, poorly differentiated;  $\eth$  fore tarsi with five distinct segments densely clothed with short, very fine hair.

Wing shape peculiar; fore wing slightly falcate, outer margin deeply excised between the apex and the end of vein 4, then strongly convex as far as the much rounded internal angle; hind wing outer margin very convex between the apex and the end of vein 3, concave between veins 3 and 1b, and with a wide, spatulate tail at the end of vein 1b, inner margin concave between 1b and the end of 1a.

Wing venation (Text-fig. 253).

Male genitalia (Text-fig. 68). (I had at my disposal only one specimen, of which the abdomen had been dorsally mutilated by a parasite, hence I cannot accurately describe the uncus.) Uncus subrectangular with rounded angles (?); no subunci; tegumen narrow; vinculum prolonged to form a very large rounded saccus; lower fultura attached to the middle of the valves, which are oblong with rounded apices; penis elongate, subcylindrical, widely open on the dorsal surface of the internal portion, vesica armed with numerous cornuti, giving it a shagreened appearance; distal edge of uncus and apices of valves pilose.

# List of Species of Paraslauga

Paraslauga kallimoides (Schultze), 1912. cephien (H. H. Druce), 1913.

### Genus EULIPHYRA Holland

Euliphyra Holland, 1890, Psyche 5: 423; Aurivillius, 1898: 285; 1920: 343; Bethune Baker, 1924: 203. Type-species: Euliphyra mirifica Holland, selected by Hemming, 1964, Annot. Lep. 1: 132.

Eyes smooth; palpi rather short, second segment of medium length, third segment much reduced; antennae short, with a sharply differentiated cylindrical club;  $\delta$  fore tarsus with five distinct segments.

Wing shape peculiar; outer margin of fore wing forming a well marked angle at the extremity of vein 4, deeply excised between veins 4 and 2; hind wing produced at its anal angle, forming a kind of obtuse tail at the extremity of vein 1b.

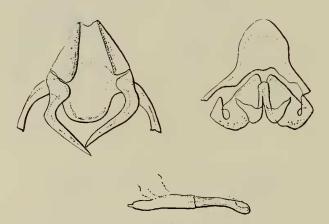


Fig. 69. Euliphyra mirifica Holland, & genitalia.

Wing venation (Text-fig. 254). The figure given by Bethune Baker (1924: 204) is not correct; he shows 13 veins in the fore wing.

Male genitalia (Text-fig. 69). Uncus trapezoidal, the posterior edge notched, the two side edges folded back on the inner surface; subunci long, robust, tapered at the apex and bent at an obtuse angle; anterior edge of tegumen strongly convex, projecting towards tergite 8; vinculum rather wide, prolonged below to form a large rounded saccus; inferior fultura composed of two lobes in the form of a crescent; valves moderately large, oblong, the apex of the upper process bent back; penis very small, elongate, subcylindrical with rounded apex; sides of uncus and distal portions of the valves bearing a few hairs.

The genitalia of *E. leucyana* are of the same type as those of *E. mirifica*, except that the subunci are bent in an acute angle. In both these species the genitalia resemble somewhat those of *Liphyra brassolis* Westwood.

Of all Ethiopian genera, Euliphyra comes nearest to the Indo-Malayan Liphyra, not only in the structure of its male genitalia and the segmented fore tarsi of the male but also in the morphology and ethology of its early stages. The caterpillar of E. mirifica has been described by Lamborn (Trans. ent. Soc. Lond. 1913: 450) and Eltringham (Trans. ent. Soc. Lond. 1913: 509 and ibid. 1921: 474). It is dorsally clothed in a tough carapace which protects it admirably from the attacks of ants (Oecophylla smaragdina longinosa Latreille), in the nest of which it lives. The conical head is borne on a sort of retractile neck. It does not seem to eat any vegetable food; the ants feed it by regurgitation and get nothing in exchange. Pupation takes place practically inside the skin of the last larval instar, and this is another analogy with Liphyra brassolis, in which, however, the chrysalis is completely enclosed in the cast-off skin.

# LIST OF SPECIES OF Euliphyra

Euliphyra hewitsoni Aurivillius, see mirifica.

- \*Euliphyra leucyana (Hewitson), 1874. Fig. Hewitson, 1878.
- \*Euliphyra mirifica mirifica Holland, 1890.

hewitsoni Aurivillius, 1898.

Euliphyra mirifica sjoestedti Aurivillius, 1895.

### Genus EGUMBIA Bethune Baker

Egumbia Bethune Baker, 1924, Ann. Mag. nat. Hist. (9) 14: 135. Type-species: Egumbia catori Bethune Baker, 1924—(Epitola ernesti Karsch, 1895) by original designation. Euliphyrodes Romieux, 1937, Mitt. schweiz. ent. Ges. 17: 120.

Bethune Baker, owing to the segmentation of the fore tarsi, mistook the type specimen of *catori* for a female, whereas it is really a male as shown by the genitalia. Karsch had previously described *ernesti* (1895) from a true female.

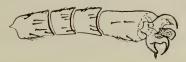


Fig. 70. Egumbia ernesti (Karsch) &, extremity of anterior tarsus.

 $\it Head$  small; eyes smooth; palpi protruding beyond the frons, slightly ascendant, second segment long, clothed below with white scales, third segment very short, blackish with a few white scales; antennae short, scarcely half the length of the costa, with a poorly differentiated fusiform club; thorax and abdomen robust, clothed below with white silky hair; legs clothed with yellow scales, tibiae shorter than the femora;  $\it 3$  fore tarsus (Text-fig. 70) five-segmented and bearing at the apex two strong claws.

Wing venation (Text-fig. 255).

Male genitalia (Text-fig. 71): dorsum (i.e. uncus and tegumen) folded over like a hood, the posterior edge rounded, with a shallow median depression, and a median rounded bulge on the anterior edge; no subcuni; vinculum rather narrow, prolonged backwards to form a large rounded saccus; lower fultra composed of two small lobes attached near the base of the valves, which are subtriangular with rounded apices; penis long, strong, subcylindrical, with its apex obliquely truncate; vesica bearing numerous small cornuti; uncus and apices of valves bearing long, fine hairs.

The genitalia somewhat resemble those of Aslauga vininga Hewitson, in the reduced dorsal elements, the absence of subunci, the large rounded saccus and the robust penis. This similarity, like the segmented fore tarsi of the male, shows that the genus Egumbia belongs to the Liphyrinae and not, as Bethune Baker supposed, to the Epitolinae. Karsch did not place ernesti in Epitola without some reserve.

The holotype of *Epitola ernesti* Karsch is a female. In 1895 Karsch did not know the male, which was only described in 1904 by Suffert (*Dt. ent. Z. Iris* 17:52). The two sexes of this species are very dissimilar in appearance, the male being silvery blue, the female almost pure white. When Bethune Baker received two specimens from Egumbe, in Northern Nigeria, he believed them to be females, though in fact they were males, as I have been able to ascertain by dissecting one of them. As it did not occur to Bethune Baker that his specimens could be Karsch's *Epitola ernesti*, he described them as *catori* and erected for them the new genus *Egumbia* at the same time. There is no doubt whatever that *catori* is the male of Karsch's *ernesti*.

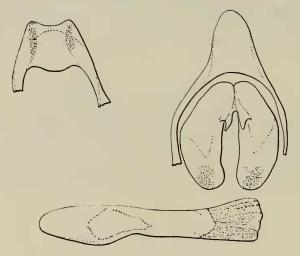


Fig. 71. Egumbia ernesti (Karsch), & genitalia.

Unfortunately I have not been able to examine the type of *katangana* Romieux, type-species of the genus *Euliphyrodes* Romieux, but judging by the description and figures published by this author, it seems to be certain that *katangana* is also a male of *ernesti*.

The synonymy of the names is indicated in the list of species given below.

## LIST OF SPECIES OF Egumbia

Egumbia catori Bethune Baker, see ernesti.

Egumbia ernesti (Karsch) (as Epitola sp.), 1895.

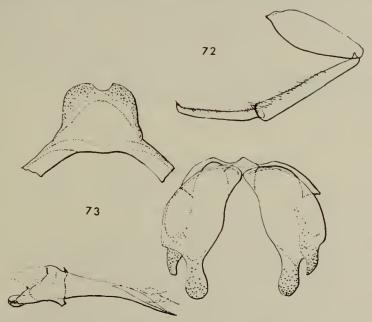
catori Bethune Baker, 1924; katangana (Romieux) (as Euliphyrodes sp.), 1937.

Egumbia katangana (Romieux), see ernesti.

### Genus TERATONEURA Dudgeon

Teratoneura Dudgeon, 1909, Proc. ent. Soc. Lond.: 50; Aurivillius, 1920: 344; Bethune Baker, 1924: 205. Type-species: Teratoneura isabellae Dudgeon, by monotypy.

Eyes smooth; palpi with second segment laterally compressed, much enlarged, oval, third segment much reduced, short, conical; antennae just under half the length of the costa, club poorly differentiated, expanding gradually, reddish brown with an orange apex; 3 fore leg (Text-fig. 72) with femur swollen, tibia long, tarsus unsegmented (in contradiction to Bethune Baker's description [1924:199]), clothed with scales, furnished below with two rows of short spines, and devoid of claws; mid leg swollen, clothed with scales; hind leg with flattened tibia and tarsus.



Figs 72-73. Teratoneura isabellae &, 72, anterior leg; 73, genitalia.

Wing shape peculiar; in the  $\Im$  the apex of the fore wing is almost rectangular, outer margin straight from the apex to the end of vein 4, concave between veins 4 and 2, inner angle much rounded; in the  $\Im$  the apex is rounded instead of angular; in both sexes the hind wing is almost quadrangular with a very concave fore margin.

Wing venation (Text-fig. 256). Fore wing with veins 2 and 3 arising from the lower edge of the cell and strongly curved towards the inner margin. Hind wing: vein 8 is very long, runs parallel to the costa and arises from the upper edge of the cell, not from the base of the wing,

as indicated in error by Bethune Baker (1924: 205).

Male genitalia (Text-fig. 73). Uncus crescentic, with a shallow notch in the posterior edge; no subunci; tegumen subtriangular; vinculum fairly wide; a small lower fultura sheathing the base of the penis; valves oblong, the two processes separated at the apex, the upper process with blunt apex, the lower lobed; penis elongate, slightly curved, the external portion tapering uniformly to the obliquely cut apex; uncus and apices of valves densely pilose.

The life-history of *T. isabellae* has been well described by Farquharson (see Eltringham, 1921, *Trans. ent. Soc. Lond.* 1921: 342 and 476, pl. 12, figs 7 to 9, 14 and 15). I give below a brief résumé of the paper. The caterpillar has the appearance of that of a species of Lymantriidae; all segments bear dorsal and lateral tubercles, and each tubercle is adorned with a tuft of long, fine, branched hairs; on segments 5 to 8 there are urticating spicules. The chrysalis, in the thoracic and dorsal regions, is covered with "chitinanths", which give it a mouldy appearance. The imago sucks up the secretion of certain *Coccidae*, after driving away the ants for which this secretion is the usual food.

The systematic position of *Teratoneura* is doubtful. Bethune Baker included it in the Liphyrinae, having erroneously endowed it with a segmented male fore tarsus, whereas, as I have said above, the male fore tarsus has the usual Lycaenid structure. The lymantriid aspect of the caterpillar and its mode of life indicate relationship with *Epitola*, *Hewitsonia*, etc.

#### LIST OF SPECIES OF Teratoneura

\*Teratoneura isabellae isabellae Dudgeon, 1909.

\*Teratoneura isabellae congoensis Stempffer, 1953, Annls Mus. R. Congo belge 46: 16.

### Genus IRIDANA Aurivillius

Iridana Aurivillius, 1920: 345. Type-species: Iris incredibilis Staudinger, by monotypy. Iris Staudinger, 1891, Dt. ent. Z. Iris 4: 141 (invalid homonym). Iridopsis Aurivillius, 1898: 286 (invalid homonym).

Eyes large, naked; palpi slightly turned upwards, protruding far beyond the frons, second segment long, thick, clothed below with short hairs, third segment one-third to one-quarter the length of the second, smooth, acuminate; antennae slender, two-thirds the length of the costa, club short; thorax long, very pilose; fore leg slender, smooth, tibia slightly longer than the femur, 3 tarsus unsegmented, about two-thirds the length of the tibia.

Wing venation (Text-fig. 257). Fore wing, vein 2 strongly recurved towards the inner margin; vein 3 also recurved but not so strongly; vein 7 is doubly curved and ends at the apex; vein 9 missing.

Male genitalia (Text-fig. 74). Tegumen subtriangular excised caudad; uncus represented by two lateral strips folded inwards; no subunci; vinculum fairly wide, prolonged into a long slender saccus; lower fultura pedunculate, sheathing the base of the penis; valves suboval, the two processes separate at the apex, the lower process notably longer than the upper; penis long, slightly curved, ending in a slender point, widely open on its upper surface; uncus and apices of valves pilose.

The male genitalia of all the known species of *Iridana* are so similar that they do not provide useful specific characters.

The larvae of *I. incredibilis* and *I. marina* have been observed by Farquharson (1921, Trans. ent. Soc. Lond. 1921: 357) and T. H. E. Jackson (1937, Trans. R. ent. Soc. Lond. 86: 208). The latter suggests that they feed on lichens among ants, though in captivity they were reared in the absence of ants. They are nocturnal, living and pupating in silken shelter tents in crevices on the bark of trees. They live in the neighbourhood of the nests of ants of the genus Crematogaster which ignore them, yet assure them of a kind of passive protection. They have no dorsal glands. The chrysalis of *I. incredibilis* has been described and figured by Eltringham (1921, Trans. ent. Soc. Lond. 1921: 477), that of marina by Jackson (l.c.: 209).

### LIST OF SPECIES OF Iridana

Iridana ansorgei (Smith), see incredibilis.

\*Iridana bwamba Stempffer, 1964: 1258, fig.

Iridana euprepes (H. H. Druce), 1905.

- \*Iridana exquisita (Smith), 1898. Fig. Smith & Kirby, 1901.
- \*Iridana gabunica Stempffer, 1964: 1246, fig.
- \*Iridana ghanana Stempffer, 1964: 1249, fig.
- \*Iridana hypocala Eltringham, 1929, Trans. ent. Soc. Lond. 1929: 494 (fig.  $\mathcal{Q}$ ). Fig. Stempffer,1964: 1243.

  magnifica Hawker Smith, 1933.

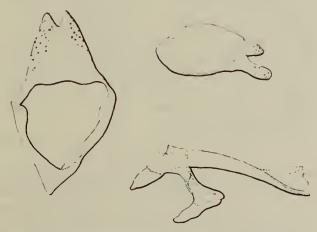


Fig. 74. Iridana incredibilis (Staudinger), & genitalia.

- \*Iridana incredibilis (Staudinger), 1891. ansorgei (Smith), 1898.
- \*Iridana jacksoni Stempffer, 1964: 1252, fig.
- \*Iridana katera Stempffer, 1964: 1255, fig.

Iridana magnifica Hawker Smith, see hypocala.

- \*Iridana marina Talbot, 1935, Entomologist's mon. Mag. 71:74 ( $\circlearrowleft$ ); Stempffer, 1964: 1241 ( $\circlearrowleft$ ), fig.
- \*Iridana nigeriana Stempffer, 1964: 1247, fig.

\*Iridana obscura Stempffer, 1964: 1259, fig.

- \*Iridana perdita (Kirby), 1890. Fig. Smith & Kirby, 1891 ( $\mathcal{Q}$ ); Stempffer, 1964: 1239 ( $\mathcal{J}$ ), fig.
- \*Iridana rougeoti Stempffer, 1964: 1244, fig.
- \*Iridana tororo Stempffer, 1964: 1253, fig.
- \*Iridana unyoro Stempffer, 1964: 1262, fig.

### Genus **DELONEURA** Trimen

Deloneura Trimen, 1868, Trans. ent. Soc. Lond. 1868: 81; Aurivillius, 1898: 287; 1920: 346; Murray, 1935: 58; Swanepoel, 1953: 189. Type-species: Deloneura immaculata Trimen, by monotypy.

Of the type-species there are only three known specimens, all females, captured by J. H. Bowker on the Bashee River, Kaffraria and preserved in the S. African Museum, Cape Town. I have been unable to examine them and must be content to reproduce Trimen's very full description.

Head wide, flattened anteriorly, clothed with scales above; eyes prominent, smooth; palpi naked, ascending, widely divergent, second segment much swollen, third segment rather short, slender, pointed; antennae of moderate length, robust, gradually becoming incrassate, the apex slightly curved outwards; thorax short, robust, smooth, sparsely clothed anteriorly with scales and posteriorly with fine hairs; legs very robust, smooth, hairless, femur and tibia in all about the same length.

Trimen considered that among his three specimens there were two males and one female, but in fact all three specimens are females. In consequence he wrongly concluded that the fore tarsus is segmented in both sexes, an error repeated by Bethune Baker (1924: 202). In 1906 Trimen, when describing *D. millari*, recognized that the segments of the male fore tarsi in *Deloneura* were fused together, short, and clothed below with fine spines. I have been able to verify this in *D. millari dondoensis* and *D. millari sheppardi*.

Wing venation. Text-fig. 258 is of D. immaculata, after Aurivillius (1898: 287); Text-fig. 259 is of D. millari.

The shape of the wings and their venation in the species of *Deloneura* have sometimes been drawn inaccurately; in Aurivillius' drawings of *D. immaculata* the inner margin of the fore wing is shown as straight, whereas Trimen says it is convex basally. Murray (1935: 58) repeats Trimen's description, but his fig. 6 of *D. millari* shows only eleven veins on the fore wing, vein 9 having been omitted.

Bethune Baker (1924: 202) also says "there are but eleven veins on the fore wing". My personal observation of *D. millari* agrees completely with the characters given by Trimen in his description of the genus, viz.: fore wing with twelve veins and inner margin basally convex.

Male genitalia of *D. millari* (Text-fig. 75). Uncus composed of two subtriangular, apically rounded lobes, separated by a deep depression of the posterior margin and united to the tegumen by a translucid zone; subunci long, strong, curved, tapering evenly to the apex; tegumen triangular; vinculum fairly wide; lower fultura consisting of a simple fold of the lower edge of the valves which are oblong with rounded apices; penis elongate, curved, with a slender base, the upper surface bearing at the beginning of the external portion two rounded expansions separated by a longitudinal groove, the apical portion slender and widely open to allow the passage of the vesica; uncus bearing long, fine hairs; valves almost bare except the upper process near its apex.

The male genitalia of *D. millari* are closely allied to those of *Epitola posthumus* Fabricius. In spite of big differences in venation and external appearance, it seems to me that the genus *Deloneura* should be included in the Epitolinae.

### LIST OF SPECIES OF Deloneura

Deloneura barca (Smith), 1901.

Deloneura immaculata Trimen, 1868.

Deloneura innesi van Son, 1949, Occ. Pap. natn. Mus. Sth. Rhod. 2 (15): 259.

Deloneura millari millari Trimen, 1906.

\*Deloneura millari dondoensis Pennington, 1953, Jl ent. Soc. sth. Afr. 16: 102, fig.

\*Deloneura millari sheppardi Stevenson, 1934, Occ. Pap. natn. Mus. Sth. Rhod. 3: 15, fig.

Deloneura subfusca Hawker Smith, 1933, Stylops 2: 10.

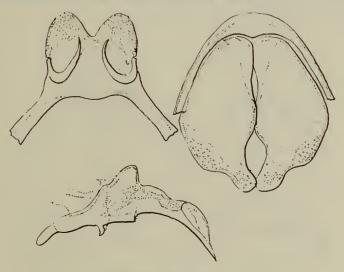


Fig. 75. Deloneura millari Trimen, & genitalia.

## Genus EBEPIUS Hemming

Ebepius Hemming, 1964, Annot. Lep. (4): 141. Type-species: Poultonia ochrascens Neave, by monotypy.

Poultonia Neave, 1904, Novit. zool. 11: 336 (invalid homonym).

Deloneura Trimen (partim); Aurivillius, 1920: 347.

Head short and wide; eyes smooth; palpi not short as stated by Neave in his description, but protruding considerably beyond the frons, second segment much swollen, third segment frail, acuminate; antennae half as long as the costa, gradually becoming thicker from the base to the poorly differentiated club; thorax robust; 3 fore tarsus unsegmented, bearing below a double row of short spines.

Wing venation (Text-fig. 260). There are several inaccuracies in the figure given by Neave (l.c.: 336); on the fore wing vein 3 arises before, and not from the lower angle of the cell, and vein 9 which branches out of 7, is omitted. On the fore wing in the 3 a secondary sexual character is present in the form of an ochreous line below and alongside the swelling in vein 1.

Male genitalia (Text-fig. 76). Uncus bilobed, the lobes subtriangular with rounded apices and separated by a median depression; subunci slender, curved; tegumen subtriangular, connected to the uncus by a very slightly chitinized zone; vinculum narrow, prolonged to form a short saccus; the small lower fultura secures the penis to the base of the valves; valves oblong with rounded apices; the penis resembles that of some species of Epitola (e.g. post-humus), narrow at the base with, on its dorsal surface, rounded expansions separated by a deep longitudinal groove, thence gradually tapering to a pointed apex; lobes of uncus and apices of valves pilose.

I do not see in the wing venation or in the genital armature any important character permitting the separation of *Ebepius* from *Deloneura*. This was also the opinion of Aurivillius (1920: 346).

The egg, caterpillar and chrysalis of *P. ochrascens* have been described by T. H. E. Jackson (1937, *Trans. R. ent. Soc. Lond.* 86: 209). The caterpillar is black, clothed with long, black hair, and resembles the caterpillar of certain Lymantriidae. It lives among ants of the genus *Crematogaster*, which seem to avoid it. It probably feeds on bark or on micro-fungi.

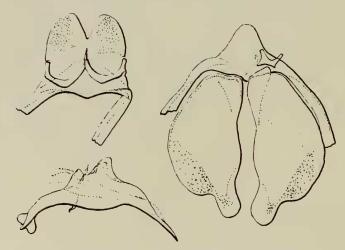


Fig. 76. Ebepius ochrascens (Neave), of genitalia.

## LIST OF SPECIES OF Ebepius

\*Ebepius ochrascens ochrascens (Neave), 1904.

\*Ebepius ochrascens littoralis (Talbot), 1935, Entomologist's mon. Mag. 71:74.

### Genus BATELUSIA H. H. Druce

Batelusia H. H. Druce, 1910, Proc. zool. Soc. Lond. 1910: 367; Aurivillius 1920: 347. Typespecies: Batelusia zebra H. H. Druce, by original designation.

Eyes large and naked; palpi ascending, distinctly protruding beyond the frons, second segment long, clothed below with adpressed scales, third segment short, slender, acuminate; antennae slender, white annulated; thorax slender; legs clothed with scales, of fore tarsus very short, unsegmented, hind tibia swollen.

Wing shape: short and broad, fore wing with costa and outer margin strongly convex; hind wing oval with blunt anal angle.

Wing venation (Text-fig. 261). Fore wing with only 11 veins, vein 9 being absent.

Druce in his original description compared the generus *Batelusia* with *Powellana*; he noted that in the fore wing veins 10 and 11 are free and do not have a common stem, but he did not mention the absence of vein 9, which however is missing in the specimens that I have examined.

Male genitalia (Text-fig. 77). Uncus crescentic with a rounded distal edge; subunci long, strongly curved, swollen in the middle and then tapering to a sharp pointed apex; tegumen triangular, somewhat heavily sclerotized; vinculum rather narrow, prolonged to form a tapering saccus; lower fultura fused to the base of the valves, closely sheathing the internal portion of the penis; valves oblong, ending in a small rounded process; penis robust, external portion ending in an obliquely cut apex; on the dorsal surface of the external portion there are two large, rounded expansions, analogous to those found in Epitola posthumus but much larger; uncus densely pilose, but valves bare except for the upper edge and the distal process.

The genitalia of zebra show close relationship to those of species of Epitola, Phytala, etc.

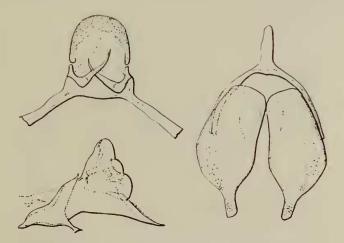


Fig. 77. Batelusia zebra Druce, & genitalia.

#### LIST OF SPECIES OF Batelusia

\*Batelusia zebra H. H. Druce, 1910.

#### Genus TUMEREPEDES Bethune Baker

Tumerepedes Bethune Baker, 1913, Ann. Mag. nat. Hist. (8) 11:564. Type-species: Tumerepedes flava Bethune Baker, by original designation.

Tumerepes Aurivillius, 1920: 564 (unjustified emendation).

Eyes smooth; palpi fairly short and yet protruding beyond the frons, third segment almost as long as the first two together, smooth, cylindrical and ending in a point; antennae short, two-fifths the length of the costa, thickening gradually up to the poorly differentiated club, segments of antenna distinct, scarcely longer than broad; legs with femora and tibiae swollen and equal in length, mid femora not so swollen as fore and hind femora.

As the unique holotype is the only known specimen, and a  $\mathcal{P}$ , I have been unable to examine the fore tarsus and genitalia of a  $\mathcal{J}$ .

Wing-venation (Text-fig. 262).

## LIST OF SPECIES OF Tumerepedes

Tumerepedes flava Bethune Baker, 1913.

#### Genus NEAVEIA H. H. Druce

Neaveia H. H. Druce, 1910, Proc. zool. Soc. Lond. 1910: 364; Aurivillius 1920: 348. Typespecies: Neaveia lamborni H. H. Druce, by original designation.

Head wide; eyes prominent, naked; palpi shining black, fairly long, turned upwards, slightly divergent, second segment long, swollen, clothed below with scales and bearing on the inner surface pale brown hairs, third segment short, slender, acuminate; antennae slender with ovoid club; thorax rather robust; legs: tibia shorter than femur, 3 fore tarsus unsegmented, spinose below, hind tibia slightly swollen.

Wing venation (Text-fig. 263).

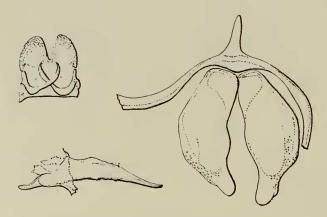


Fig. 78. Neaveia lamborni Druce, & genitalia.

Male genitalia (Text-fig. 78). Uncus composed of two subtriangular lobes with rounded lateral margins separated by a deep median depression; subunci long, curved, dilated in the middle; tegumen subtriangular; vinculum rather narrow, prolonged to form a slender saccus; lower fultura sheathing the base of the penis and fused to a fold of the base of the valves; valves oblong with rounded apices, shaped as in *Epitola*; penis elongate, with a slender obliquely truncate apex, the lower surface of the external portion bearing a few fine spines; uncus densely pilose, only a few hairs on the upper edge of the distal portion of the valves.

### LIST OF SPECIES OF Neaveia

\*Neaveia lamborni lamborni H. H. Druce, 1910.

Neaveia lamborni orientalis Jackson, 1962, Bull. Br. Mus. nat. Hist. (Ent.) 12: 158, figs.

## Genus PSEUDONEAVEIA Stempffer

Pseudoneaveia Stempsfer, 1964: 1265. Type-species: Pseudoneaveia jacksoni Stempsfer, by original designation.

Eyes naked; second segment of palpi swollen, brown-scaled; third segment acuminate blackish, brown-tipped; stem of antennae weakly white-ringed, club weak, blackish; tarsus of  $\eth$  fore leg unsegmented, spinose beneath.

Wing venation (Text-fig. 264).

Male genitalia (Text-fig. 79, in profile). Uncus composed of two subtriangular lobes, rounded apically folded back and fused at right angles to the tegumen; subunci absent; vinculum narrow, prolonged to form a short saccus; lower fultura ensheathing the base of the penis; valves subrectangular with a slightly recurved finger-like apex; penis elongate, the distal portion gently curved, the sharp-pointed apex dorsally widely open; uncus densely hairy, the upper margin of the valves also hairy, but less densely, towards the apex.

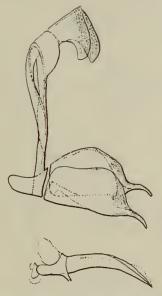


Fig. 79. Pseudoneaveia jacksoni Stempffer, & genitalia.

The dorsal (tergal) part of the genitalia in *P. jacksoni* is profoundly different from that of other Epitolinae. In these the uncus in general crescentic, more or less excised caudad, bounding an oval-shaped tegumen and, besides, provided with well-developed subunci. It is because of this very special condition that I have been led to erect the genus *Pseudoneaveia*.

### LIST OF SPECIES OF Pseudoneaveia

\*Pseudoneaveia jacksoni Stempffer, 1964: 1265.

#### Genus **EPITOLINA** Aurivillius

Epitolina Aurivillius, 1895, Ent. Tidsk. 16: 205; 1898: 287; 1920: 348. Type-species: Teriomima dispar Kirby, by monotypy.

Eyes naked; palpi long, protruding far beyond the frons, second segment slightly swollen, clothed below with adpressed scales, third segment long, slender, acuminate; segments of antennae long and thin, club ovoid, well differentiated; thorax clothed ventrally with long, silky brown hairs; legs black and white annulated; of fore tarsus unsegmented, hind tarsi swollen.

Wing venation (Text-fig. 265). Vein 9 very long and branching from 7 near its base.

Male genitalia (Text-fig. 8o). Uncus shaped like a segment of a circle, the apex forming a rounded weakly obtuse angle; subunci long, curved, the median portion swollen and with its lower edge forming an acute angle, tapering at the apex; tegumen fairly large; vinculum rather narrow, with a much reduced saccus; valves oblong, widened at the base, the lower process ending in a blunt point; penis elongate, gently curving, the apex obliquely cut to a sharp point; uncus and upper process of valves densely and finely pilose.

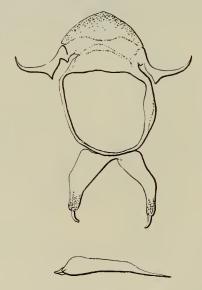


Fig. 80. Epitolina dispar (Kirby), & genitalia.

The genitalia of *E. catori* are of the same type as those of *E. dispar*, but the posterior margin of the uncus forms an acute angle apically, the subunci are less swollen medially, and the valves are shorter and wider.

*Epitolina* is scarcely distinguishable from *Epitola* unless by its feeble stature and the longer vein 9 of the fore wings.

### LIST OF SPECIES OF Epitolina

- \*Epitolina catori catori Bethune Baker, 1904.
- \*Epitolina catori ugandae Jackson, 1962: 159, figs.
- \*Epitolina dispar (Kirby), 1887, fig. Smith & Kirby, 1888. mnestra (Möschler), 1888.
  - Epitolina dispar f. dubia (Kirby), 1890.
- \*Epitolina dispar f. cordelia (Kirby), 1890.
- \*Epitolina dispar f. melissa (H. H. Druce), 1888.

Epitolina mnestra (Möschler), see dispar.

### Genus STEMPFFERIA Jackson

Stempfferia Jackson, 1962: 157. Type-species: Stempfferia carcassoni Jackson, 1962, by original designation.

Eyes faintly pubescent; palpi protruding beyond the frons, black, the second segment laterally compressed, third segment cylindrical, acuminate; antennae half as long as fore wing costa, black above, ringed with white below, club slightly swollen; legs not swollen, fore tarsi of the  $\eth$  unsegmented, pilose beneath.

Sexual dimorphism is considerable: on the upperside the  $\beta$  is clear blue, the  $\varphi$  slightly yellowish white.

Wing venation (Text-fig. 266). In the  $\eth$  fore wing vein 10 is weakly stalked on vein 7, and vein 11 arises from the same point as vein 7; in the  $\heartsuit$  veins 10 and 7 are also connate, and vein 11 arises very shortly before them.

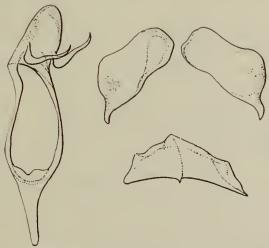


Fig. 81. Stempfferia carcassoni Jackson, of genitalia of type.

Male genitalia (Text-fig. 81). Uncus in the form of a crescent, the distal margin not excised; subunci long, curved, swollen at the bend, ending in a sharp point; tegumen subtriangular; vinculum rather narrow, prolonged to form a long triangular saccus which bears long coremata apically. Lower fultura very reduced; valves subquadrangular, the upper process ending in a gently curved point. Penis short, very robust, the apex obliquely truncate and widely open dorsally. Uncus and distal part of the upper process of the valves covered with long fine hair.

## LIST OF SPECIES OF Stempfferia

\*Stempfferia carcassoni Jackson, 1962: 157, figs.

### Genus PHYTALA Westwood

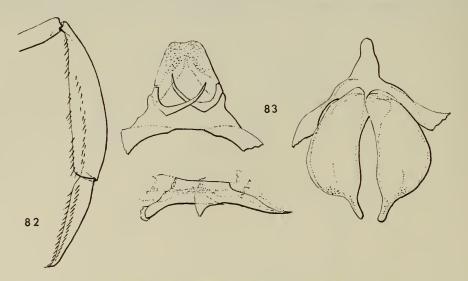
Phytala Westwood 1851, Gen. Diurn. Lep. plate 77; Aurivillius, 1898: 288; 1920: 348. Type-species: Phytala elais Westwood (l.c.), by monotypy.

Eyes naked; palpi scarcely projecting beyond the frons, second segment long, swollen, clothed with adpressed scales, third segment very short, ovoid; antennae a little over half the length of the costa, club very gradual, poorly differentiated, fusiform; of fore leg (fig. 82) with tibia swollen slightly longer than the femur, tarsus unsegmented, tapering evenly, tibia and tarsus bearing below two rows of spines; mid and hind legs with similarly swollen tibiae, longer than the femora and bearing two rows of spines.

Secondary sexual character in the  $\delta$  a patch of blackish brown hair-scales at the base of the inner margin of the fore wing.

Wing venation (Text-fig. 267). Fore wing with vein 11 short and uniting with 12 (this is not an entirely constant character; in a specimen of *P. elais catori* which I examined, veins 11 and 12 run close together but remain separate right to the costa).

Male genitalia (Text-fig. 83). Uncus crescentic, the posterior edge with a shallow notch; subunci long, curved, slightly dilated in the middle and from there gradually tapering to the apex; tegumen triangular; vinculum fairly broad, prolonged to form a stout saccus; the small lower fultura fused to a fold of the base of the valves and closely sheathing the base of the



Figs 82-83. Phytala elais catori Bethune Baker 3, 82, fore leg; 83, genitalia.

penis; valves oblong, ending in a slightly recurved process with rounded apex; penis elongate, subcylindrical, slightly curved, the apex obliquely truncate and widely open; uncus and median and distal parts of the valves all very pilose.

I have examined the genitalia of eight other species besides P. elais, namely P. vansomereni, P. hyettoides, P. intermixta, P. henleyi, P. schultzei, P. reducta, P. benitensis and P. rezia.

The genitalia of all these species are all much alike and of a type common to most of the Epitolinae, but the dorsal surface of the penis bears rounded expansions which are not present in *P. elais*. This last species is rather isolated in the genus *Phytala* both in its larger size and in the shape of its hind wings, in which the margin is slightly angled.

## LIST OF SPECIES OF Phytala

Phytala aequatorialis Jackson, 1964: 62, figs.

\*Phytala benitensis (Holland), 1890. Fig. Jackson, 1964.

Phytala elaidina Strand, see elais catori.

\*Phytala elais elais Westwood, 1851.

Phytala elais catori Bethune Baker, 1903.

elais f. elaidina Strand, 1920.

\*Phytala elais ugandae Jackson, 1964: 60, figs.

\*Phytala henleyi (Kirby), 1890. Fig. Smith & Kirby, 1892.

Phytala hyetta (Hewitson), 1873. Fig. Hewitson, 1878. Phytala hyettina Aurivillius, 1897. Fig. Aurivillius, 1898.

- \*Phytala hyettoides (Aurivillius), 1895. Fig. Aurivillius, 1920.
- \*Phytala intermixta Aurivillius, 1897. Fig. Aurivillius, 1898.

Phytala nigrescens Jackson, 1964: 67, figs.

Phytala obscura Schultze, 1915, Arch. Naturgesch. 81, A, 12: 142.

\*Phytala reducta Aurivillius, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910-11, 1: 1197. Fig. Jackson, 1964.

\*Phytala rezia (Smith & Kirby), 1893.

- \*Phytala schultzei Aurivillius, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910-11, 1: 1196, fig.
- \*Phytala vansomereni Jackson, 1964: 60, figs.

#### Genus EPITOLA Westwood

Epitola Westwood, 1851, Gen. Diurn. Lep. pl. 68, fig. 5; Aurivillius, 1898: 289; 1920: 349. Type-species: Epitola elion Westwood, 1851, l.c., by monotypy (Papilio posthumus Fabricius, 1793).

Eyes large, smooth; palpi ascending, protruding well beyond the frons, clothed with adpressed scales, second segment robust, much swollen, third segment short, slender, acuminate; antennae long and fine with a gradually swollen ovoid club;  $\delta$  fore leg with femur somewhat swollen, tibia long and slender, tarsus short, unsegmented, finely spinose below.

Wing venation (Text-fig. 268). Fore wing with 11 and 12 not confluent, 10 and 11 arising free from anterior border of cell; hind wing cell produced at its lower angle, the lower discocellular being very oblique.

The venation is not entirely constant in all the species of *Epitola*. In *carcina*, *leonina* and *zelza* vein 10 arises as a branch of 7 not far from its origin; in other species vein 11 and 12 are sometimes in contact.

Male genitalia (Text-fig. 84, A, B). Uncus subtriangular, with rounded apex, joined to the tegumen by a semimembranous piece which is translucid under the microscope; subunci long, gently curved, swollen in the middle, tapering towards the apex; tegumen large; vinculum rather large and prolonged to form a long saccus, which is directed towards the genital orifice and bears at the apex a tuft of long black scales; lower fultura fused to the base of the valves and sheathing the base of the penis; valves oblong with widely rounded apices. Uncus clothed with an abundance of long, fine hairs, upper process of valves pilose in its distal third.

The penis is variable. I have dissected the genitalia of six specimens of "posthumus", all externally very similar. In four specimens (Text-fig. 84, B) the structure agrees with the figures given by Talbot (1921, Bull. Hill Mus. Witley 1: 1, pl. 8, fig. 7) that is to say that on the middle of the dorsal surface there are two rounded expansions separated by a deep longitudinal groove and the ventral surface bears only a few very fine spines; and the distal part tapers regularly and is widely open dorsally. In the other two specimens the general shape of the penis is the same (Text-fig. 84A), and bears the same dorsal expansions but on the ventral surface there is moreover in the middle a small protuberance which bears irregular teeth followed by fine spines. I do not know whether we are here dealing with individual variations or a pair of sibling species, but the second hypothesis seems to be the more probable.

The genus *Epitola* is numerous in species. I have studied the genitalia of only a limited number of them, since they rarely furnish specific characters. The dorsal elements and the valves are very much alike in all those I have examined. I have only found appreciable differences in the form of the penis. The dorsal expansion of the penis in *posthumus* recurs, strongly developed, in *urania* (c.f. Talbot l.c. fig.

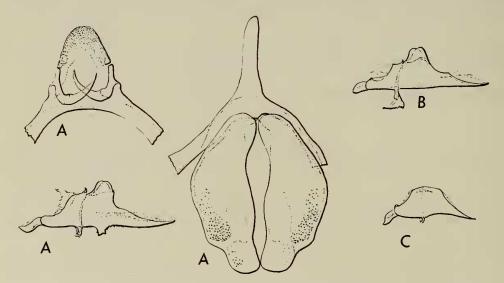


Fig. 84. Epitola spp., & genitalia, (A, B) E. posthumus (Fabricius), (C) E. cercene Hewitson.

10), less so in decellei and ceraunia. It does not occur in the following species:—
hewitsoni (Text-fig. 85), stempfferi, crowleyi, conjuncta, carcina, marginata, cercene,
moyambina, cercenoides, iturina, staudingeri and leonina.

The pupae of *E. hewitsoni*, *E. miranda*, *E. concepcion*, *E. carcina* and *E. ceraunia* have been described and figured by Eltringham (1922, *Trans. ent. Soc. Lond.* 1921: 473–5, pl. 12, figs 1, 3, 13, 18). The chrysalis of *E. hewitsoni* is quite remarkable, as it bears tubercles armed with strong, curved spines.

# LIST OF SPECIES OF Epitola

Epitola adolphifrederici Schultze, 1911. Fig. Seitz, 1920.

Epitola alba Jackson, 1962: 155, figs.

Epitola albomaculata Bethune Baker, 1903 (♂); 1904 (♀). Fig. Seitz.

Epitola ammon Joicey & Talbot, see semibrunnea.

Epitola azurea Jackson, 1962: 148, figs.

Epitola badia Kirby, see zelza.

Epitola badura Kirby, 1890. Fig. Smith & Kirby, 1891.

Epitola batesi H. H. Druce, see cercenoides.

Epitola bella Aurivillius, see iturina.

Epitola belli Hewitson, see posthumus.

Epitola bwamba Jackson, 1964: 73, figs.

\*Epitola carcina Hewitson, 1873. Fig. Hewitson, 1878.

kholifa Bethune Baker, 1904.

Epitola carilla Roche, 1954: 495, figs.

Epitola catuna Kirby, 1890. Fig. Smith & Kirby, 1892.

mus Suffert, 1904.

**Epitola catuna carpenteri** Bethune Baker, 1921, Trans. ent. Soc. Lond. 1921:  $462 \ (3)$ ; Jackson, 1962: 144, fig. (9).

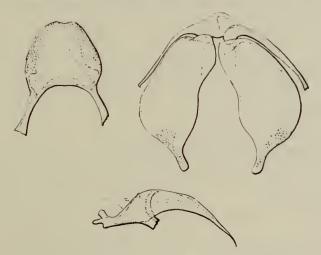


Fig. 85. Epitola hewitsoni Mabille, & genitalia.

Epitola cephena cephena Hewitson, 1873. Fig. Hewitson, 1878 (♀).

doleta Kirby, 1890 (3); leonina Bethune Baker, 1903; leonensis Bethune Baker, 1904.

**Epitola cephena entebbeana** Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) **17**: 392 (♂); Jackson, 1962: 146, fig. (♀).

\***Epitola ceraunia** Hewitson, 1873. Fig. Hewitson, 1878 (♂). dewitzi Kirby, 1887 (♀).

\***Epitola cercene** Hewitson, 1873. Fig. Hewitson, 1878 (3). versicolor Kirby, 1887 (2).

\*Epitola cercenoides Holland, 1890.

batesi H. H. Druce, 1910.

Epitola ciconia Kirby, see leonina Staudinger.

Epitola coerulea Jackson, 1962: 140, figs.

Epitola concepcion Suffert, 1904. Fig. Aurivillius in Seitz, 1920.

Epitola congoana Aurivillius, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910-11, 1: 1202.

\*Epitola conjuncta conjuncta Smith & Kirby, 1893.

Epitola conjuncta budduana Talbot, 1937, Trans. ent. Soc. Lond. 86: 62, fig.

**Epitola convexa** ♂ Roche, 1954, l.c.: 496, figs. (♂); Jackson, 1962: 131, figs. (♀).

\*Epitola crowleyi E. M. Sharpe, 1890.

hewitsoni Staudinger, 1889 (invalid homonym).

Epitola cyanea Jackson, 1964: 71, figs.

Epitola daveyi Roche, 1954: 499, figs.

\*Epitola decellei Stempffer, 1956: 30, figs.

Epitola dewitzi Kirby, see ceraunia.

Epitola doleta Kirby, see cephena.

Epitola dolorosa Roche, 1954: 498, figs.

*Epitola dorothea* Bethune Baker, 1904. Fig. Jackson, 1962.

Epitola dubia Jackson, 1964: 70, figs.

**Epitola dunia** Kirby, 1887. Fig. Smith & Kirby, 1889 (3); Jackson, 1962: 141, figs. (\$\time\$).

Epitola elion Westwood, see posthumus.

**Epitola elissa** Grose Smith, 1898. Fig. Smith & Kirby, 1902 (3); Jackson, 1962: 139, (2), figs.

oniensis Bethune Baker, 1913.

Epitola falkensteini Dewitz, see hewitsoni.

Epitola flavoantennata Roche, 1954: 495, figs.

**Epitola gerina** Hewitson, 1878 (3); Jackson, 1964:68, figs. (9).

**Epitola ghesquierei** Roche, 1954: 498, figs. (♂); Jackson, 1962: 145, figs. (♀). nigeriae Jackson, 1962.

**Epitola goodi** Holland, 1890. Fig. Smith & Kirby, 1892 ( $\mathfrak{P}$ ); Jackson, 1964: 69, figs. ( $\mathfrak{F}$  &  $\mathfrak{P}$ ).

\*Epitola hewitsoni Mabille, 1877. Fig. Aurivillius in Seitz, 1920. falkensteini Dewitz, 1879.

Epitola hewitsoni Staudinger, see crowleyi.

Epitola hewitsonioides Hawker Smith, 1933, Stylops 2: 11. Fig. Jackson, 1964.

Epitola ikoya Roche, 1954: 497, figs.

**Epitola insulana** Aurivillius, 1923, Ergebn. 2te D. Zent. Afr. Exped. 1910–11, 1: 1203, fig. (♂); Jackson, 1962: 133, fig. (♀).

**Epitola intermedia** Roche, 1954: 497, fig. (3); Jackson, 1964: 74, fig. (9).

\*Epitola iturina Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1: 85, fig. bella Aurivillius, 1923.

Epitola jacksoni Roche, see mara.

Epitola kamengensis Jackson, 1962: 150, fig.

Epitola katerae Jackson, 1962: 149, fig.

Epitola katharinae Poulton, 1929, Trans. ent. Soc. Lond. 77: 494, fig.

Epitola kholifa Bethune Baker, see carcina.

Epitola lamborni Bethune Baker, 1921, Trans. ent. Soc. Lond. 1921: 461.

Epitola leonensis Bethune Baker, see cephena.

Epitola leonina Bethune Baker, see cephena.

\*Epitola leonina Staudinger, 1888; Jackson, 1962: 136, fig. (\$). ciconia Kirby, 1892.

**Epitola liana** Roche, 1954: 500, fig. (3); Jackson, 1962: 138, fig. ( $\mathcal{P}$ ).

Epitola maculata Hawker Smith, 1926, Revue zool. afr. 14: 240. pulchra Jackson, 1964.

\*Epitola magnifica Jackson, 1964, Ann. Mag. nat. Hist. (13) 7:699, fig.

**Epitola mara** Talbot, 1935, Entomologist's mon. Mag. 71:75 (3); Jackson, 1962:135, fig. ( $\mathfrak{P}$ ).

jacksoni Roche, 1954.

**Epitola marginata marginata** Kirby, 1887. Fig. Smith & Kirby, 1889; Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1: 85, fig. (Ω).

Epitola marginata umbratilis Holland, 1980.

**Epitola mengoensis** Bethune Baker, 1906 (3). Fig. Aurivillius in Seitz, 1920 ; Jackson, 1962 : 153, fig. (9).

Epitola mercedes Suffert, 1904. Fig. Jackson, 1962.

**Epitola miranda miranda** Staudinger, 1889. Fig. Smith & Kirby, 1893 (3); Jackson, 1962: 126, fig.  $(\mathfrak{P})$ .

**Epitola miranda vidua** Talbot, 1935, Entomologist's mon. Mag. **71**: 75 (3); Jackson, 1962: 127, fig.  $(\mathfrak{P})$ .

Epitola mirifica Jackson, 1964, : 72, fig.

\*Epitola moyambina Bethune Baker, 1903, (3); Jackson, 1962: 130, fig. (2).

Epitola mpangensis Jackson, 1962: 149, figs.

Epitola mus Suffert, see catuna.

Epitola nigeriae Jackson, 1962, see ghesquierei.

Epitola nigra Bethune Baker, 1903.

Epitola nigrovenata Jackson, 1962: 136, figs.

Epitola nitide H. H. Druce, 1910.

Epitola obscura Hawker Smith, 1933, Stylops 2: 11.

Epitola oniensis Bethune Baker, see elissa.

Epitola orientalis Roche, 1954: 499, fig.

Epitola ouesso Jackson, 1962: 147, fig.

Epitola pinodes H. H. Druce, 1890. Fig. Smith & Kirby, 1891.

Epitola pinodoides Smith & Kirby, 1893.

\*Epitola posthumus (Fabricius), 1793. Fig. Hewitson, 1878. elion Westwood, 1851 (3); belli Hewitson, 1874 (2).

Epitola pseudoconjuncta Jackson, 1962: 153, figs.

Epitola pulchra Jackson, 1964, see maculata.

Epitola pulverulenta Dufrane, 1953, Bull. Annls Soc. R. ent. Belg. 89: 51.

Epitola rileyi Audeoud, 1936, Bull. Soc. lépidopt. Genève 7, 5 : 186, fig.

Epitola semibrunnea Bethune Baker, 1916, Ann. Mag. nat. Hist. (8) 17:378. ammon Joicey & Talbot, 1921.

\*Epitola staudingeri staudingeri Kirby, 1890. Fig. Aurivillius in Seitz, 1920 (3); Jackson, 1962: 134, fig. (4).

Epitola staudingeri aequatorialis Jackson, 1962: 135, figs.

Epitola staudingeri gordoni H. H. Druce, 1903.

\*Epitola stempfferi Jackson, 1962: 127, fig.

Epitola subalba Bethune Baker, see zelica.

Epitola subargentea Jackson, 1964, Ann. Mag. nat. Hist. (13) 7:698, fig.

Epitola subcoerulea Roche, 1954: 498, figs.

Epitola subgriseata Jackson, 1964: 72, figs.

Epitola sublustris Bethune Baker, 1904.

**Epitola tumentia** H. H. Druce, 1910 (3); Jackson, 1962: 137, fig. (9).

**Epitola uniformis** Kirby, 1887 (3). Fig. Smith & Kirby, 1889; Jackson, 1962: 133, figs. (9).

versicolor (3) Kirby, 1887.

\*Epitola urania urania Kirby, 1887 (3). Fig. Smith & Kirby, 1889; Jackson, 1962: 128, figs. (2).

**Epitola urania tanganikensis** Joicey & Talbot, 1921, Bull. Hill Mus. Witley **1**: 86, fig. (♂); Jackson, 1962: 129, fig. (♀).

Epitola versicolor Kirby, 3 see uniformis, \$\varphi\$ see cercene.

**Epitola vinalli** Talbot, 1935, Entomologist's mon. Mag. **71**: 75 (3); Jackson, 1962: 154, figs. ( $\mathcal{P}$ ).

**Epitola virginea** Bethune Baker, 1904 ( $\mathcal{P}$ ); Roche, 1954:501, fig. ( $\mathcal{J}$ ).

Epitola viridana viridana Joicey & Talbot, 1921, Bull. Hill. Mus. Witley 1:84, figs. (♂). Fig. ♀ (as orientalis ♀) Jackson, 1962:143.

**Epitola viridana radiata** Bethune Baker, 1926, *Ann. Mag. nat. Hist.* (9) **17**: 393. **Epitola zelica** Kirby, 1890 (♂). Fig. Smith & Kirby, 1892; Jackson, 1964: 69, fig. (♀).

subalba Bethune Baker, 1915.

Epitola zelza Hewitson, 1873. Fig. Hewitson, 1878. badia Kirby, 1887.

## Genus NEOEPITOLA Jackson

Neopitola Jackson, 1964, Bull. Br. Mus. nat. Hist. (Ent.) 15: 78. Type-species: Epitola barombiensis Kirby, 1890, by original designation.

Epitola Westwood (partim): Aurivillius, 1898: 293; 1920: 358.

Eyes smooth; palpi reaching beyond the frons, the second segment broad, laterally compressed, third segment short, slender, acuminate; antennae less than half the length of costa, annulated black and white, club distinct and flattened; legs black with a few white scales, fore tarsi of 3 unsegmented, tibiae of hind legs swollen.

Wing venation (Text-fig. 269). On the fore wing vein 11 arises from 10 and is very close to, but not touching, vein 12.

Male genitalia (Text-fig. 86). Uncus crescentic, its posterior margin slightly depressed dorsally; subunci rather stout, curved near the base; tegumen suboval; vinculum rather broad, prolonged as a long triangular saccus; lower fultura attached to a fold of the base of the valves, tightly enclosing the base of the penis; valves oblong, obliquely truncate at the apex, and with the upper process bearing a kind of subtriangular harpe clothed in small short spines; penis elongate, the dorsal and ventral margins of the external part deeply excised, the whole resembling a shallow sickle complete with handle and curved blade; uncus densely pubsecent, the distal fourth of valves slightly hairy.

## LIST OF SPECIES OF Neoepitola

Neoepitola barombiensis (Kirby), 1890. Fig. Smith & Kirby, 1892.

#### Genus AETHIOPANA Bethune Baker

Aethiopana Bethune Baker, 1915, Ann. Mag. nat. Hist. (8) 16: 191. Type-species: Papilio honorius Fabricius, by original designation.

Epitola Westwood (partim); Aurivillius, 1898: 291; 1920: 351.

Eyes large, smooth; palpi ascending, protruding far beyond the frons, clothed with adpressed scales, second segment robust, much swollen, third segment short, slender, acuminate; antennae

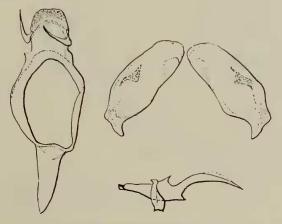


Fig. 86. Neoepitola barombiensis (Kirby), o genitalia.

long, slender, with a swollen club; 3 fore leg with tibia long, thin, tarsus short, unsegmented. Secondary sexual characters. On the fore wing in the 3 there is a large androconial dull blackish brown patch situated between the upper edge of the cell, vein 5 and the costa, and vein 1 is swollen and underlined by long brown hairs.

Wing venation (Text-fig. 270). Fore wing 8, 9, and 10 are stalked on 7 which arises from the upper margin of the cell well before its upper angle, 11 free; hind wing cell drawn out at the

lower angle.

Male genitalia (Text-fig. 87): uncus subtriangular, united to the tegumen by a slightly sclerotized translucent area; subunci long, curved, tapering towards the apex; tegumen trapezoidal; vinculum broad and prolonged to form a long saccus; lower fultura tightly sheathing the base of the penis; valves oblong and with rounded apices as in Epitola; penis long, slightly curved, tapering gradually to its apex, and devoid of the dorsal expansions of Phytala and some species of Epitola; uncus densely clothed with many long, fine hairs; a few short hairs on the distal portion of the valves.

The caterpillar and chrysalis of *A. honorius* have been described and figured by Eltringham (1922, *Trans. ent. Soc. Lond.* 1921: 474, pl. 12, figs 16 and 17). The caterpillar bears on each segment four tubercles adorned with tufts of fine spines and long, delicate, branched hairs. The chrysalis bears dorsal and lateral tubercles furnished with recurved spines and chitinanths on its abdominal segments.

# LIST OF SPECIES OF Aethiopana

\*Aethiopana honorius honorius (Fabricius), 1793. teresa (Hewitson), 1869.

Aethiopana honorius ab. & coarctata Hulstaert, 1924, Revue zool. afr. 12: 118. Aethiopana honorius divisa (Butler), 1901.

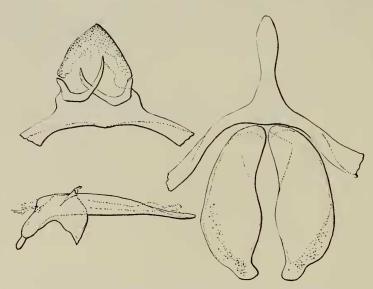


Fig. 87. Aethiopana honorius honorius (Fabricius), & genitalia.

## Genus HEWITSONIA Kirby

Hewitsonia Kirby, 1871, Cat. Diurn. Lep.: 426, nom. nov. pro Corydon Hewitson, 1869 (Ill. Diurn. Lep. Suppl.: 1), praeocc.; Aurivillius, 1898: 294; 1920: 359. Type-species: Corydon boisduvali Hewitson, 1869, by monotypy.

Eyes large, glabrous; palpi long, ascending, distinctly protruding beyond the frons, second segment robust, clothed below with adpressed scales, third segment slender, acuminate; antennae long, slender, with a poorly differentiated, gradually swollen club;  $\vec{o}$  fore tibia long, its unsegmented tarsus finely spinose below.

Wing venation (Text-fig. 271). On the hind wing the lower discocellular is straight, not concave as in *Epitola* so that the cell is not drawn out at its lower angle.

Male genitalia (Text-fig. 88): uncus crescentic, its posterior margin very slightly excised, joined to the tegumen by a lightly sclerotized zone, which appears translucent under the microscope; subunci long, fairly robust, curved, tapering gradually to the apex; tegumen subtriangular; vinculum moderately broad and prolonged to form a long saccus bearing at its tip a tuft of long black scales; valves oblong with rounded apices as in *Epitola*; penis robust, gently curved, ending in a sharp point, widely open dorsally; uncus densely clothed with long, fine hair, apex of the valves bearing short hairs.

I have examined the male genitalia of Hewitsonia similis, H. kirbyi and H. magdalenae. Those of H. bitjeana have been figured by Joicey and Talbot (1921, Bull. Hill Mus. Witley 1, pl. 8, fig. 3). These four species are all very similar to boisduvalii. In a recent work Jackson (1964: 3) has figured the genitalia of H. mittoni. In this the dorsal structures and the valves are clearly of the Hewitsonia type, but the penis shows the rounded dorsal expansions which are present in Phytala and certain species of Epitola.

The caterpillar and chrysalis of *H. intermedia* have been described by T. H. E. Jackson (1937, *Trans. R. ent. Soc. Lond.* 86: 209). The caterpillar feeds on lichens. It resembles that of some species of Lymantriidae; fore part of body wider than hind part, dorsum smooth, sides and extremity fringed with long, fine hairs, and on each segment some shorter, thicker light brown silky hairs.

The chrysalis of H. similis has been described and figured by Eltringham (1922, Trans. ent. Soc. Lond. 1921: 478, pl. 12, fig. 2).

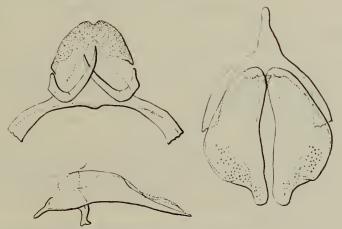


Fig. 88. Hewitsonia boisduvali boisduvali Hewitson, of genitalia.

#### LIST OF SPECIES OF Hewitsonia

Hewitsonia beryllina Schultze, see bitjeana.

Hewitsonia bitjeana Bethune Baker, 1915, Ann. Mag. nat. Hist. (8) 16: 190. beryllina Schultze 1916.

\*Hewitsonia boisduvali boisduvali (Hewitson, 1869).

Hewitsonia boisduvali ab. virilis Aurivillius, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910–11, 1: 1209.

Hewitsonia boisduvali ab. ♀ albifascia Hulstaert, 1924, Revue zool. afr. 12: 118. Hewitsonia boisduvali borealis Schultze, 1916, Arch. Naturgesch. 81, A, 11: 111. Hewitsonia boisduvali congoensis Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1: 87, fig.

Hewitsonia boisduvali nigeriensis Jackson, 1962: 160, figs.

Hewitsonia crippsi (Stoneham), 1933, Bull. Stoneham Mus. 17: 1.

Hewitsonia intermedia intermedia Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1: 86, fig.

Hewitsonia intermedia gomensis Dufrane, 1953, Bull. Annls Soc. R. ent. Belg. 89: 52.

\*Hewitsonia kirbyi Dewitz, 1879. preussi Staudinger, 1890.

\*Hewitsonia magdalenae Stempffer, 1951, Bull. Soc. ent. Fr. 56: 67, fig.

Hewitsonia mittoni Jackson, 1964: 77, figs.

Hewitsonia preussi Staudinger, see kirbyi.

\*Hewitsonia similis similis Aurivillius, 1891.

Hewitsoni similis ugandae Jackson, 1962: 160, figs.

#### Genus POWELLANA Bethune Baker

Powellana Bethune Baker, 1908, Proc. zool. Soc. Lond. 1908: 114; Aurivillius, 1920: 360. Type-species: Powellana cottoni Bethune Baker, by original designation.

Eyes naked; palpi protruding far beyond the frons, second segment slightly swollen, clothed with adpressed scales, third segment fairly long, acuminate; antennae slender, with a poorly differentiated, gradually swollen, fusiform club;  $\delta$  fore leg with long slender tibia, short unsegmented tarsus, bearing fine spines below.

Wing venation (Text-fig. 272). Vein II branches from 10 near its base, a rare condition which occurs also in Neoepitola.

Male genitalia (Text-fig. 89). Uncus crescentic, the hind edge evenly rounded; subunci long, slender, curving, swollen in the distal half, then tapering; tegumen triangular; vinculum narrow, prolonged to form a long saccus; lower fultura fused to a fold of the base of the valves, closely sheathing the base of the penis; valves oblong, ending in a small rounded process; penis robust, elongate, slightly curved, the apex obliquely cut and widely open; uncus and distal edge of valves densely pilose.

The genitalia of P. cottoni closely resemble those of most of the species of Epitola.

### LIST OF SPECIES OF Powellana

\*Powellana cottoni Bethune Baker, 1908.

weberi (Holland), 1913; virginea Birket Smith, 1960.

Powellana virginea Birket Smith, see cottoni.

Powellana weberi (Holland), see cottoni.

### Genus MEGALOPALPUS Röber

Megalopalpus Röber, 1886, Dt. ent. Z. Iris 1:51; Aurivillius, 1898:300; 1923:361. Typespecies: Megalopalpus simplex Röber, by original designation.

M. simplex was described as coming from Borneo, very probably the result of an error in labelling, for, as far as I know, it has never been found again in the Oriental Region, whilst it is common in Africa from the Gold Coast, Liberia, Nigeria, Cameroon, Gaboon and Congo to Uganda.

Head small; eyes smooth; palpi very long, even longer than in the Indo-Malayan genera Miletus and Allotinus, ascending, laterally compressed, clothed with short, adpressed hairs, third segment slightly longer than the second; antennae slender, less than half as long as the costa, club fusiform, very slightly swollen; thorax slender; abdomen long, protruding far beyond the anal angle; 3 fore leg with tibia slightly shorter than the femur, tarsus very long, unsegmented, pubescent and with a single claw; mid and hind legs with tibiae slightly shorter than femora, first tarsal segment very long; no terminal claws.

In the shape of the hind wings the sexes show a marked difference. The outer margin in the 3 has a slight angle at the end of vein 5, which in the 9 is much more prominent.

Wing venation (Text-fig. 273): fore wing with 11 veins, vein 9 being absent; hind wing with a short precostal vein which arises from vein 8 near its base.

Male genitalia (Text-fig. 90): uncus composed of two enormous triangular lobes with sharp pointed apices, fused at their base to the tegumen and separated by a very deep groove; subunci fairly robust, regularly curved and tapering to the apex; tegumen well developed with the

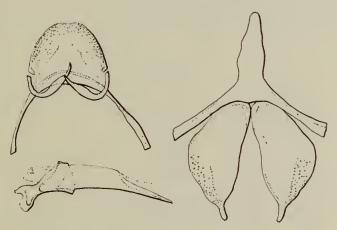


Fig. 89. Powellana cottoni Bethune Baker, & genitalia.

posterior edge divided to form two hooks; in the normal position the tegumen is folded over and the two lobes of the uncus face one another, the straight edges of the groove being dorsal; vinculum wide dorsally, narrow below; no lower fultura; valves much reduced compared with the dorsal structures, finger-shaped with a small, sharp tooth on the inferior edge near the apex; penis subcylindrical, elongate, very slender; uncus with a felt-like covering of fine hairs, distal portion of the valves pilose.

The male genitalia of M. zymna and M. metaleucus are very similar to those of simplex.

Bethune Baker (1914, Trans. ent. Soc. Lond. 1914: 317, pl. 58, figs 9 and 9a) figured the genitalia of "Megalopalpus gigas", the name occurring only in the explanation of the plate. The genitalia figured correspond exactly with those of M. simplex.

The early stages of *M. zymna* have been described by W. A. Lamborn (1914, *Trans. ent. Soc. Lond.* 1913: 458). The caterpillar is protected from attack by ants by a coriaceous skin bearing tubercles tipped with coarse hairs; it is carnivorous and feeds on Jassidae and Membracidae (Hemiptera). Its mode of life is the same as that of the caterpillar of *Gerydus chinensis* Felder and demonstrates the close relationship of *Megalopalpus* to the Indo-Malayan genera *Gerydus*, *Allotinus* etc.

The shape of the palpi, the venation and the genitalia indicate that *Megalopalpus* should be included in the sub-family Miletinae (= Gerydinae), as Aurivillius and Bethune Baker have already pointed out.

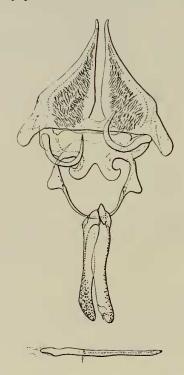


Fig. 90. Megalopalpus simplex Röber, & genitalia.

## LIST OF SPECIES OF Megalopalpus

Megalopalpus angulosus Grünberg, 1910.

Megalopalpus bicoloria (Capronnier), see simplex.

Megalopalpus gigas Bethune Baker, 1914, Trans. ent. Soc. Lond. 1914, explanation of pl. 58, see simplex.

\*Megalopalpus metaleucus Karsch, 1893. Fig. Aurivillius in Seitz, 1923.

Megalopalpus similis (Kirby), see simplex.

\*Megalopalpus simplex Röber, 1886.

bicoloria (Capronnier), 1889; similis Kirby, 1890; gigas Bethune Baker, 1914.

\*Megalopalpus zymna (Westwood), 1851.

Megalopalpus zymna f. pallida Aurivillius, 1923.

### Genus LACHNOCNEMA Trimen

Lachnocnema Trimen, 1887, S. African Butterflies 2: 233; Aurivillius, 1898: 301; 1923: 362; Murray, 1935: 55; Pinhey, 1949: 97; Swanepoel, 1953: 191. Type-species: Papilio bibulus Fabricius, 1793, selected by Hemming, 1960, Annot. Lep. 1: 11.

Head small, pilose; eyes densely hairy; palpi long, ascending, first and second segments clothed below with stiff hair, third segment long, acuminate; antennae short, thick, with a poorly differentiated, cylindrical, blunt-tipped club; thorax short, very hairy; legs short, robust, femora and tibiae clothed with scales and dense long, woolly hair which conceals the basal portion of the tarsi; tarsi short, robust, scaly, with a few short bristles, spinose below; of fore tarsus segmented like the other tarsi, but rather smaller and very hairy.

Wing venation (Text-fig. 274).

Male genitalia (Text-fig. 91): uncus subrectangular, the lateral angles rounded, and slightly folded back; subunci very long, bent at right-angles at one-third of their length, tapering gradually to an apex which bears a distinct hook; tegumen reduced to a narrow band; vinculum rather narrow, prolonged to form a spatulate saccus; inferior fultura well developed, like a furca, arising from the base of the valves; valves elongate with broadly rounded apices; to the upper process is articulated distally a long, finger-like process bent in the shape of an open V; the lower margin of the valves bears, about midway, a broad triangular expansion with rounded apex, and, midway beyond this a weak blunt tooth; penis elongate, fairly robust, slightly curved, tapering gradually to an obtuse apex; vesica bears fine cornuti which give it a shagreened appearance; uncus almost bare, distal portion of valves densely and finely pilose.

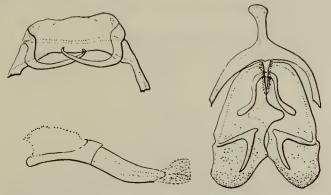


Fig. 91. Lachnocnema bibulus (Fabricius), & genitalia.

The genitalia of *L. durbani* Trimen and *L. magna* Aurivillius closely resemble those of *L. bibulus*.

The caterpillar of *L. bibulus* has been frequently described:—Lamborn (1914, *Trans. ent. Soc. Lond.* 1913: 470); Farquharson (*ibid.*, 1921: 388); Jackson (1937, *Trans. R. ent. Soc. Lond.* 86: 210); Cripps and Jackson (*ibid.*, 1940, 89: 449–453), and Pinhey (1949: 97.)

The caterpillar lives and pupates in ants' nests, to which it has been carried by the ants. It feeds partly on the frass of Jassidae and Membracidae (Hemiptera), partly on plant sap and partly on food regurgitated by the ants. Lamborn (1913) noted that in captivity the caterpillar devoured numerous Jassidae.

### LIST OF SPECIES OF Lachnocnema

\*Lachnocnema bibulus (Fabricius), 1793. Fig. Staudinger, 1887. delegorguei (Boisduval), 1847; emperamus (Snellen), 1872.

Lachnocnema brimo Karsch, 1893. Fig. Aurivillius in Seitz, 1923.

sudanica Aurivillius, 1905; obliquisigna Hulstaert, 1924; rectifascia Hulstaert, 1924.

Lachnocnema busoga Bethune Baker, 1906.

Lachnocnema delegorguei (Boisduval), see bibulus.

Lachnocnema disrupta Talbot, 1935, Entomologist's mon. Mag. 71: 76, figs.

Lachnocnema divergens Gaede, 1915. Fig. Aurivillius in Seitz, 1923.

\*Lachnocnema durbani Trimen, 1887. Fig. Aurivillius in Seitz, 1923.

Lachnocnema emperamus (Snellen), see bibulus.

Lachnocnema exiguus Holland, 1890. Fig. Aurivillius in Seitz, 1923.

Lachnocnema luna H. H. Druce, see reutlingeri.

\*Lachnocnema magna Aurivillius, 1895. Fig. Aurivillius in Seitz, 1923. umbra (Smith), 1901; niveus H. H. Druce, 1910.

Lachnocnema niveus H. H. Druce, see magna.

Lachnocnema obliquisigna Hulstaert, see brimo.

Lachnocnema rectifascia Hulstaert, see brimo.

Lachnocnema reutlingeri Holland, 1892. Fig. Aurivillius in Seitz, 1923. luna H. H. Druce, 1910.

Lachnocnema sudanica Aurivillius, see brimo.

Lachnocnema umbra (Smith) see magna.

#### Genus **DEUDORIX** Hewitson

Deudorix Hewitson, 1863, Ill. Diurn. Lep., Lycaenidae 1:16. Type-species: Dipsas epijarbas Moore (an Indian species), by original designation.

Eyes shortly and densely pilose; palpi scarcely protruding beyond the frons, second segment long, laterally compressed, clothed with adpressed scales, third segment short, slender, acuminate; antennae two-thirds the length of the costa, club elongate, fusiform; thorax very

robust, especially in the 3, clothed below with long white hair; 3 fore leg, femur clothed with long white hair, tibia shorter than the femur, tarsus unsegmented, finely spinose below.

Wing shape: hind wing oval, apex rounded, outer margin showing a slight salient at the end of vein 3, vein 2 prolonged to form a short filiform tail, a small rounded lobe between the end of vein 1b and the anal angle, abdominal margin slightly excised between the lobe and the end of vein 1a. The presence of this lobe renders easy the recognition of the species of Deudorix.

Wing venation (Text-fig. 275). The venation in the figure given by Murray (1935: 60) is incorrect; the filiform tail is shown at the end of vein 1b when in fact it is at the end of vein 2.

Male genitalia (Text-fig. 92): uncus composed of two lateral lobes separated by the rounded depression of the hind edge of the tegumen; subunci long, strong, bent in an acute angle, suddenly narrowed a little before the apex, and with a small apophysis on the lower side at the level of the bend; tegumen very large, hood-shaped; vinculum narrow with a small round saccus; no lower fultura; valves small compared with the other structures, broadly fused together in their lower oval halves, distally consisting of a slightly recurved process which has an obliquely truncated apex, about midway the upper edges are folded inwards and connected to each other by a membrane; penis elongate, subcylindrical, widely open dorsally and proximally, widening apically; vesica with a group of cornuti ending in a single more robust apical spine; uncus and middle part of valves pilose.

The description given above is based on the type-species alone, *Deudorix epijarbas*. Aurivillius included in the genus *Deudorix* all the Ethiopian Lycaenidae whose hind wings have the shape of those of *D. epijarbas* (i.e. with a lobe near the anal angle and a filiform tail at the end of vein 2) and whose venation agrees, at least on the whole, with that of *D. epijarbas*. But among these species there are some that differ from the type-species of *Deudorix*, either by the more rounded shape of the fore wings or by some venational detail, or by the presence in the male of conspicuous secondary sexual characters. Taking into account these differences, Karsch and H. H. Druce have erected the following genera for African species of *Deudorix* (sensu Aurivillius), *Hypomyrina* Druce, *Actis* Karsch, *Kopelates* Druce, *Hypokopelates* Druce, *Pilodeudorix* Druce, and *Diopetes* Karsch. Most modern authors make use of these genera and they also assign to the Indo-Malayan genus *Virachola* Moore certain species included in *Deudorix* by Aurivillius. To test the validity of the abovementioned genera I have examined their type-species and give the results under the appropriate genera (See also p. 108).

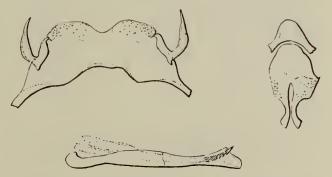


Fig. 92. Deudorix epijarbus (Moore), & genitalia.

### LIST OF SPECIES OF Deudorix

There is no African species that agrees precisely in all morphological characters with *Deudorix epijarbas*, and so would fall into typical *Deudorix*.

#### Genus HYPOMYRINA H. H. Druce

Hypomyrina H. H. Druce, 1891, Ann. Mag. nat. Hist. (6) 7:364. Type-species: Myrina nomenia Hewitson, by original designation.

Deudorix Hewitson (partim); Aurivillius, 1898; 306; 1923; 365.

Eyes shortly and densely pilose; palpi moderately long, clothed with adpressed scales, third segment slender, acuminate, two-thirds the length of the second segment; antennae long, slender, white annulated, club fusiform; legs black and white annulated,  $\delta$  fore leg with tibia shorter than the femur, tarsus unsegmented, clothed below with stiff hair; no secondary sexual characters in the  $\delta$ .

Wing venation (Text-fig. 276).

Male genitalia (Text-fig. 93): uncus composed of two rounded lobes separated by a shallow depression of the hind edge of the tegumen; subunci long, curved, fairly robust, tapering gradually to the apex; tegumen very large, hood-shaped; vinculum fairly broad, without saccus; inferior fultura absent; valves oblong, fused together basally for about two-fifths of their length, apex pointed, the two upper processes being connected on their inner sides by a narrow band which surrounds the penis; penis elongate, subcylindrical; vesica with numerous cornuti and enclosing a long cuneus which is slightly recurved; uncus densely pilose, a few hairs on the distal portions of the valves.

# LIST OF SPECIES OF Hypomyrina

Hypomyrina acares (Karsch), see nomenia.

Hypomyrina nomenia nomenia (Hewitson), 1874. Fig. Hewitson, 1878. acares (Karsch), 1893.

\*Hypomyrina nomenia nomion (Staudinger), 1891.

Hypomyrina nomenia fournierae Gabriel, 1939, Ruwenzori Exp. 1934–35, 3: 74, fig.

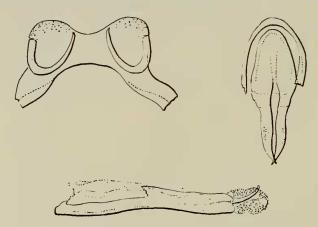


Fig. 93. Hypomyrina nomenia (Hewitson), & genitalia.

### Genus ACTIS Karsch

Actis Karsch, 1895, Ent. Nachr. 21: 315. Type-species: Actis mimeta Karsch, selected by Hemming, 1960, Annot. Lep. 1:8.

Deudorix Hewitson (partim); Aurivillius, 1898: 306; 1923: 366.

Eyes large, densely but shortly pilose; palpi rather long, ascending, second segment long, laterally compressed, with long hairs below at its base, third segment short and slender; antennae long, slender, white annulated, club fusiform;  $\delta$  fore tarsus unsegmented.

Wing shape. Inner border of fore wing slightly lobed in basal third. The 3 has the following secondary sexual characters: on the underside of the base of the fore wing there is a patch between vein 1 and the lower edge of the cell, and on the inner margin a small brush of tawny hairs lying along the surface of the wing and directed towards the aforementioned patch; on the upper side of the hind wing there is a small clear silky patch between vein 8 and the origin of vein 7 and partly covered by the small lobe of the inner margin of the fore wing.

Wing venation (Text-fig. 277). Fore wing with 11 veins; 10 and 11 free, from the upper edge

Male genitalia (Text-fig. 94). Uncus composed of two rounded lobes separated by the median depression of the hind edge of the tegumen; subunci rather long, slender, curving; tegumen very large, hood-shaped; vinculum moderately wide with an indistinct saccus; valves very elongate, the lower process ending in a point, the upper process much shorter and folded inwards to envelope the penis, which is robust, dilated distally, its apex crowned with fine, erect spines; vesica with many cornuti and enclosing an enormous cuneus. Uncus clothed with long fine hair, valves slightly hairy apically.

The male genitalia of Actis ula are similar.

#### LIST OF SPECIES OF Actis

\*Actis mimeta mimeta Karsch, 1895, (3). Fig. Aurivillius in Seitz, 1923. perigrapha (Karsch), 1895 (2).

Actis mimeta unda (Gaede), 1915.

Actis perigrapha (Karsch), see mimeta.

\*Actis ula Karsch, 1895.

Actis ula ab. nigrostriata (Aurivillius), 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910-11, 1: 1215.

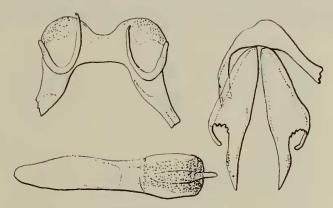


Fig. 94. Actis mimeta mimeta Karsch, & genitalia.

### Genus KOPELATES H. H. Druce

Kopelates H. H. Druce, 1891, Ann. Mag. nat. Hist. (6) 7:364. Type-species: Kopelates virgata H. H. Druce, by original designation.

Deudorix Hewitson (partim); Aurivillius, 1898: 307; 923: 366.

Eyes shortly and densely pilose; palpi protruding only slightly beyond the frons, second segment long and robust, clothed below with white adpressed scales, third segment slender, acuminate; antennae long, slender, white annulated, with an elongate fusiform club; legs black and white annulated, 3 fore tarsus unsegmented.

Wing shape: fore wing inner margin with a slight lobe near its base. Male secondary sexual characters: on the underside of fore wing there is a patch of modified scales at the base between vein I and the lower edge of the cell, and on the inner margin a brush of adpressed brown hairs directed towards the above patch; on the upperside of the hind wing there is a very small creamy white spot, with yellowish centre, near the origin of vein 8.

Wing venation (Text-fig. 278). Fore wing with 11 veins; 10 free, from the upper edge of the cell; 11, to a great extent, confluent with 12.

Male genitalia (Text-fig. 95). Very similar to those of Actis perigrapha Karsch, except that the uncal lobes are not so prominent and the hind margin of the tegumen between them has only a very shallow median depression.

The genus *Kopelates* differs from the genus *Actis* only by the confluence of veins 11 and 12 in the fore wing.

## LIST OF SPECIES OF Kopelates

Kopelates gracilis (Staudinger), see virgata.

\*Kopelates virgata H. H. Druce, 1891 (April). Fig. (as gracilis) Staudinger, 1891. gracilis (Staudinger), 1891 (July).

#### Genus HYPOKOPELATES H. H. Druce

Hypokopelates H. H. Druce, 1891, Ann. Mag. nat. Hist. (6) 7:364. Type-species: Hypolycaena mera Hewitson, by original designation.

Deudorix Hewitson (partim); Aurivillius, 1898: 306; 1923: 366.

Eyes shortly and densely pilose; palpi very slightly protruding beyond the frons, second segment clothed below with white adpressed scales, third segment slender, acuminate, shorter than that of the species of *Hypomyrina*; antennae long, slender, white annulated, with a

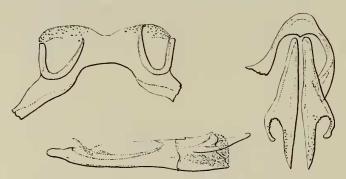


FIG. 95. Kopelates virgata Druce, & genitalia.

gradually swollen fusiform club; thorax clothed below with long, silky white hairs; & fore leg femur clothed with long white hairs, tibia shorter than the femur, tarsus unsegmented, bearing below small stiff bristles.

Wing shape: fore wing inner margin slightly lobed in its basal half. Male secondary sexual characters: on the underside of the fore wing there is a tuft of long black hairs on the inner margin at the level of its convexity; on the upperside of the hind wing there is a matt glandular patch on the origin of vein 7.

Wing venation (Text-fig. 279). Fore wing with 11 veins; 10 and 11 free, from the upper

edge of the cell; vein 8 absent.

Male genitalia (Text-fig. 96). Uncus composed of two lobes separated by the shallow rounded depression of the posteior edge of the tegumen; subunci long, rather slender, strongly curved, with a very shallow apical hook; tegumen very large hood-shaped,; vinculum narrow with an indistict saccus; no lower fultura; valves narrow, their lower edges widely fused together basally, their upper processes connected in the middle by a narrow band which surrounds the penis; penis elongate, wide open on the upper surface proximally; vesica bearing many cornuti, which give it a shagreened appearance, and enclosing a long cuneus; uncus and distal parts of the valves pilose.

I have examined the male genitalia of all the Hypokopelates. They are very similar to those of H. mera, described above. In most cases they do not even present useful specific characters.

# LIST OF SPECIES OF Hypokopelates

\*Hypokopelates anetia (Hulstaert), 1924, Revue zool. afr. 12: 120.

- \*Hypokopelates anetta Talbot, 1935, Entomologist's mon. Mag. 71: 76, fig.
- Hypokopelates angelita angelita (Suffert), 1904. Fig. Aurivillius in Seitz, 1923.
- \*Hypokopelates angelita makala (Bethune Baker), 1908.
- \*Hypokopelates angelita schultzei (Aurivillius), 1907.
- \*Hypokopelates aruma aruma (Hewitson), 1873. Fig. Hewitson, 1878.
- \*Hypokopelates aruma simplex (Schultze), 1917.
- \*Hypokopelates azurea Stempffer, 1964: 1280, figs.
- \*Hypokopelates canescens Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1: 90, figs.
- \*Hypokopelates catori (Bethune Baker), 1903, (3); Stempffer, 1964: 1280 (2).
- \*Hypokopelates cobaltina Stempffer, 1964: 1271, figs.

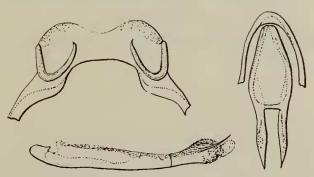


Fig. 96. Hypokopelates mera (Hewitson), & genitalia.

- \*Hypokopelates eleala (Hewitson), 1865.
- \*Hypokopelates elealodes (Bethune Baker), 1908.

mariana (Hulstaert), 1924.

Hypokopelates feminina Hulstaert, see kafuensis.

Hypokopelates fusca (Aurivillius), 1922 (? = canescens Joicey & Talbot, 1921).

Hypokopelates genuba (Hewitson), see otraeda.

\*Hypokopelates infuscata Stempffer, 1964: 1275, figs.

\*Hypokopelates ituri (Bethune Baker), 1908.

- \*Hypokopelates ituri f. lineosa Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1:89.
- \*Hypokopelates kafuensis (Neave), 1910.

feminina (Hulstaert), 1924.

Hypokopelates kallipygos Birket Smith, 1960, Bull. Inst. fr. Afr. noire 22: 974, fig.

\*Hypokopelates leonina (Bethune Baker), 1904.

\*Hypokopelates marginata Stempffer, 1962: 1157, figs.

Hypokopelates mariana (Hulstaert), see elealodes Bethune Baker.

\*Hypokopelates mera (Hewitson), 1873. Fig. Hewitson, 1878.

Hypokopelates mera f. kinumbensis Dufrane, 1945, Bull. Annls Soc. R. ent. Belg. 81: 120.

\*Hypokopelates moyambina (Bethune Baker), 1904.

\*Hypokopelates nyanzana Stempffer, 1957, Bull. Inst. fr. Afr. noire 19: 214, fig.

\*Hypokopelates obscura Bethune Baker, 1913.

- \*Hypokopelates otraeda Hewitson, 1863. genuba (Hewitson), 1875.
- \*Hypokopelates otraeda f. modesta Talbot, 1935, Entomologist's mon. Mag. 71:77.
- \*Hypokopelates petersi Stempffer & Bennett, 1956, Bull. Inst. fr. Afr. noire 23: 507, figs.
- \*Hypokopelates tenuivittata Stempffer, 1951, Bull. Soc. ent. Fr. 56: 119, fig.
- \*Hypokopelates ugandae Talbot, 1935, Entomologist's mon. Mag. 71:77.

\*Hypokopelates ultramarina Stempffer, 1964: 1272, figs.

\*Hypokopelates viridis Stempffer, 1964: 1269, figs.

#### SPECIES INCERTA

Hypokopelates rava (Holland), 1892. (? aruma Hewitson ♀).

#### Genus **PILODEUDORIX** H. H. Druce

Pilodeudorix H. H. Druce, 1891, Ann. Mag. nat. Hist. (6) 7:366. Type-species: Pilodeudorix barbatus H. H. Druce, 1891 (camerona Plötz, 1880), by original designation.

Deudorix Hewitson (partim); Aurivillius, 1898:307; 1921:369.

Eyes shortly and densely pilose; palpi not protruding beyond the frons, second segment clothed below with erect bristles and scales, third segment slender, acuminate; antennae long,

slender, with an elongate fusiform club; & fore leg, femur clothed with long white hairs, tibia shorter than the femur, tarsus unsegmented, clothed below with stiff hair.

Wing shape. Fore wing inner margin slightly lobed in its basal portion. Male secondary sexual characters: on underside of fore wing in the basal half a silky, shining patch which is silvery between veins 1 and 2 and violet-black between veins 2 and 3; on the inner margin a tuft of long black hairs directed towards this patch; on upperside of hind wing a small whitish spot near the origin of vein 8 and, along vein 2, a tuft of very long black hairs directed towards the anal margin.

Wing venation (Text-fig. 280). Fore wing with 11 veins; veins 10 and 11 free.

Male genitalia (Text-fig. 97). Uncus composed of two lobes separated by the rounded depression of the posterior edge of the tegumen; subunci long and bent about mid-length; tegumen very large, hood-shaped; vinculum narrow; no lower fultura; valves oblong, fused at the base, distal half digitate, upper edges folded in and connected by a membrane which surrounds the penis; penis elongate, subcylindrical, the external portion much shorter than the internal portion; vesica enclosing an enormous cuneus; uncus and apices of valves pilose.

The male genitalia of all the species of *Pilodeudorix* that I have examined are very similar to those of *camerona*, described above.

The caterpillars of *P. camerona* and *P. diyllus* live among ants on flowers of *Pterocarpus esculentus* Schumach (Leguminosae), see Farquharson, 1922, *Trans. ent. Soc. Lond.* 1921: 381.

### LIST OF SPECIES OF Pilodeudorix

\*Pilodeudorix ankoleensis Stempffer, 1953, Annls Mus. R. Congo belge 27: 21. camerona ugandae Stempffer, 1946.

Pilodeudorix barbatus H. H. Druce, see camerona.

Pilodeudorix bemba (Neave), 1910.

\*Pilodeudorix caerulea caerulea (H. H. Druce), 1890. Fig. Aurivillius in Seitz, 1921.

hollandi Ehrman, 1894 (Argiolaus).

\*Pilodeudorix caerulea obscurata (Trimen), 1891.

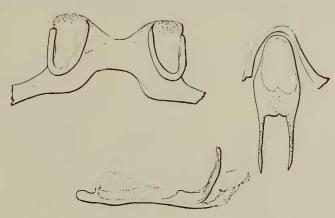


Fig. 97. Pilodeudorix camerona (Plötz), & genitalia.

\*Pilodeudorix camerona camerona (Plötz), 1880. Fig. Aurivillius in Seitz, 1921. barbatus H. H. Druce, 1891; nobilis (Staudinger), 1891.

Pilodeudorix camerona katanga Clench, 1966, Jl N.Y. ent. Soc. 73: 178.

Pilodeudorix camerona ugandae Stempffer, see ankoleensis.

Pilodeudorix congoana (Aurivillius), 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910–11, 1: 1218.

Pilodeudorix debilis (Staudinger), see divllus.

- \*Pilodeudorix diyllus diyllus (Hewitson), 1878. debilis (Staudinger), 1891.
- \*Pilodeudorix diyllus orientalis Stempffer, 1957, Bull. Inst. fr. Afr. noire 19: 217.

Pilodeudorix hollandi Ehrman, 1894, see caerulea.

\*Pilodeudorix kohli (Aurivillius), 1921 (3); Stempffer, 1962: 1159,  $(\mathfrak{P})$ . Pilodeudorix nobilis (Staudinger), see camerona.

Pilodeudorix simplex (Staudinger), see zela.

- \*Pilodeudorix zela zela (Hewitson), 1869. simplex (Staudinger), 1891.
- \*Pilodeudorix zela zeloides (Butler), 1901.
- \*Pilodeudorix zelomina (Rebel), 1914.

### Genus DIOPETES Karsch

Diopetes Karsch, 1895, Ent. Nachr. 21: 317. Type-species: Deudorix deritas Hewitson, by original designation.

Deudorix Hewitson (partim); Aurivillius, 1898: 307; 1921: 371.

Eyes shortly and densely hairy; palpi moderately long, scarely protruding beyond the frons, second segment clothed below with long, erect scales, third segment slender with a blunt apex; antennae long, slender, with a fusiform club; 3 fore tarsus unsegmented.

Wing shape. Fore wing outer margin very convex, inner margin with a lobe in its basal half. Male secondary sexual characters: on the underside of the fore wing, level with the lobe of the inner margin, there is a strong tuft of black lying close against the wing; on the upperside of the hind wing there is a large matt yellowish rounded spot between the upper edge of the cell and

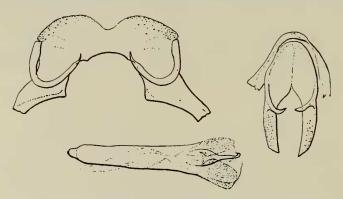


Fig. 98. Diopetes deritas (Hewitson), of genitalia.

the origin of vein 8, covering the origin of vein 7, and visible as a slight circular prominence on the underside; in the normal position of the wings it is to a great extent covered by the lobe of the inner margin of the fore wing.

Wing venation (Text-fig. 281). Fore wing with 11 veins; 10 and 11 free, from the upper edge of the cell.

Male genitalia (Text-fig. 98). Uncus composed of two flattened lobes separated by the shallow depression of the posterior margin of the tegumen; subunci fairly robust, long, bent at about mid-length; tegumen very large, hood-shaped; vinculum wide dorsally, narrow ventrally; no lower fultura; valves oblong with pointed apices, the lower edge of each straight, the upper edge convex with a small median process which is normally folded down over the inner suface, the two processes being united by a delicate membrane which surrounds the penis as in the preceding genera; penis elongate, widely open in the doral surface of the inner portion; the vesica bears numerous cornuti giving it a shagreened appearace, and encloses an enormous cuneus which has a hook-shaped, recurved apex; uncus and apices of valves pilose.

The male genitalia of all the species of *Diopetes* examined closely resemble those of *D. deritas*.

## LIST OF SPECIES OF Diopetes

- \*Diopetes aucta Karsch, 1895.
- \*Diopetes aurivilliusi Stempffer, 1954, Bull. Soc. ent. Fr. 59: 106.
- \*Diopetes bwamba Stempffer, 1962: 1161, figs.
- \*Diopetes catalla Karsch, 1895, (3). Fig. Aurivillius in Seitz, 1921; fig.  $\varphi$ , Stempffer, 1954, Bull. Soc. ent. Fr. 59: 105.
- \*Diopetes corruscans (Aurivillius), 1897. Fig. Aurivillius in Seitz, 1921.
- \*Diopetes deritas (Hewitson), 1874. Fig. Hewitson, 1878.
- \*Diopetes fumata Stempffer, 1954, Bull. Soc. ent. Fr. 59: 107, fig.
- \*Diopetes kedassa H. H. Druce, 1910.

Diopetes laticlavia Clench, 1965, Jl N.Y. ent. Soc. 73: 180.

Dioptes nirmo Clench, 1965, l.c.: 179.

Diopetes pasteon H. H. Druce, 1910.

\*Diopetes pseudoderitas Stempffer, 1964: 1283, fig.

Diopetes sadeska Clench, 1966, l.c.: 179.

\*Diopetes violetta (Aurivillius), 1897. Fig. Aurivillius in Seitz, 1921.

#### Genus VIRACHOLA Moore

Virachola Moore, 1881, Lep. Ceylon 1: 104. Type-species: Deudorix perse Hewitson, 1862, (Indo-Malayan species), by original designation.

Deudorix Hewitson (partim); Aurivillius, 1898: 308; 1921: 373; Murray 1935: 60; Pinhey, 1949: 98; Swanepoel, 1953: 159.

Eyes shortly and densely hairy; palpi hardly protruding beyond the frons, second segment shorter than in species of Deudorix, laterally compressed, clothed below with adpressed scales, third segment very short with rounded apex; antennae about two-thirds the length of the costa, club gradually swollen, fusiform; thorax very robust, hairy above and below;  $\delta$  fore leg with femur hairy, tibia shorter than femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing in the  $\delta$ , apex very pointed, outer margin straight, even slightly concave between the ends of veins 3 and 1, inner margin lobed in its basal half; in the  $\circ$  the apex is less pointed and the outer margin more convex, the inner margin not lobed. Male

secondary sexual characters: on underside of fore wing there is a tuft of long hairs on the inner margin on a level with the lobe; on the upper side of the hind wing there is a matt glandular patch below vein 8 and covering the origins of veins 6 and 7.

Wing venation (Text-fig. 282). Fore wing with 11 veins; 10 and 11 free, from the upper

edge of the cell.

Male genitalia (Text-fig. 99). Uncus composed of two lobes separated by the rounded depression of the posterior margin of the tegumen; subunci long, robust, bent at an acute angle, suddenly narrowed in the apical quarter of their length, and on the lower edge, level with the bend, there is a small pointed apophysis; tegumen large, hood-shaped; vinculum rather narrow with a small round saccus; no lower fultura; valves small compared with the other parts, their proximal halves broadly fused together along their lower margins, the distal third consisting of two finger-like processes which are slightly recurved, their upper edges connected on the inner side by a membrane; penis elongate, subcylindrical, slightly dilated at the apex; vesica bearing a series of cornuti; uncus and distal portions of the valves pilose.

I have been able to examine the male genitalia of nearly all the African species of Virachola. Except in some details, they are of the same type as those of V. perse, described above. The subunci carry a small apophysis in caliginosa, odana, galathea, dinomenes, dinochares, lorisona, diocles, dariaves, wardii and batikeli. The subunci of the other species lack this apophysis. The valves are short, blade-shaped and fused together for almost their entire length in livia, suk, ecaudata, caliginosa, vansoni, penningtoni and antalus. In the other species they taper more and their general shape recalls the valves of the preceding genera.

The early stages of several species of *Virachola* have been described, see Aurivillius, 1921: 375; Murray, 1935: 61–3; Farquharson, 1921, *Trans. ent. Soc. Lond.* 1921: 377–8; Jackson, 1937, *Trans. R. ent. Soc. Lond.* 86: 211; Pinhey, 1949: 98–99. As a general rule the caterpillars of species of *Virachola* live in the pods of Mimosaceae and Papilionaceae; the caterpillar of *V. antalus* seems polyphagous as it has been found on *Crotalaria capensis*, *Canavalia ensiformis* and *Acacia stenocarpa*. The caterpillar of *V. jacksoni* on the other hand feeds on the young leaves of *Loranthus usuiensis* and that of *V. dinochares* on the fruits of *Syzygium cordatum*.

At this point it is opportune to examine the group of Ethiopian genera erected by the subdivision of the genus *Deudorix*. To me it does not seem that their characteristics taken as a whole allow a clear-cut separation.

Eyes, palpi and antennae are closely alike in all the genera. In wing shape, the fore wing of the male is triangular with a straight costa, apex pointed, outer margin almost straight in *Pilodeudorix* and *Virachola* (although in *V. odana*, *V. diocles* and

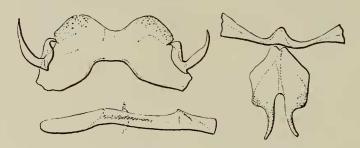


Fig. 99. Virachola perse (Hewitson), & genitalia.

V. galathea the apex is less pointed and the outer margin slightly convex); costa slightly convex, apex gently rounded, outer margin more or less convex in species of Hypomyrina and Hypokopelates; outer margin distinctly convex in species of Actis, Kopelates and Diopetes. In all the genera the hind wing is lobed between the end of vein 1b and the anal angle, tailed at the end of vein 2, and the outer margin is more or less angular at the end of vein 3. However, a single exception is provided by V. ecaudata (dohertyi B. Baker) which lacks a tail. Male secondary sexual characters are present in all the genera except Hypomyrina.

Venation, except for some minor details, is identical in all the genera, with the exception of *Kopelates*, where veins II and I2 of the fore wing are anastomosed, but I do not think that much stress should be laid on this character because I have found

the same peculiarity in a specimen of Hypokopelates eleala Hewitson.

The male genitalia are of a uniform type in all the genera; however, in species of *Virachola* the penis is generally more elongate and the subunci shorter and more robust, often bearing a small apophysis on the lower edge.

None of these characters seems to me to be of real generic value. I agree with Aurivillius, who concluded that all these subdivisions of *Deudorix* were at most subgenera or even only simple groups of species.

#### LIST OF SPECIES OF Virachola

Virachola alticola (Aurivillius), see lorisona.

Virachola anta Trimen, see antalus.

Virachola antalus antalus (Hopffer), 1855. Fig. Hopffer, 1862; Stempffer, 1938, Mission Omo 4: 179 (genitalia).

anta (Trimen), 1862; gambius (Mabille), 1885.

Virachola antalus kitobolensis (Strand), 1912.

Virachola badhami (Carcasson), 1961, Occ. Pap. Coryndon meml Mus. 7: 19, fig. Virachola baronica (Ungemach), 1932, Mem. Soc. Sci. nat. phys. Maroc 82, fig. Virachola batikeli (Boisduval), 1833.

licinia (Mabille), 1878; derona (Smith), 1891.

Virachola batikeli ab. tsiphana (Boisduval), 1833.

Virachola batikelides (Holland), 1920, Bull. Am. Mus. nat. Hist. 43: 221, fig.

Virachola bimaculata (Hewitson), see lorisona.

\*Virachola caliginosa (Lathy), 1903. Fig. genitalia, Gifford, 1963, Entomologist 96:46.

Virachola chalybeata (Joicey & Talbot), 1926, Entomologist 59: 225.

\*Virachola dariaves (Hewitson), 1877. Fig. Hewitson, 1878.

Virachola derona (Smith), see batikeli.

\*Virachola dinochares dinochares (Smith), 1887. Figs. Monteiro, 1891; Stempffer, 1938, Mission Omo 4: 179, (genitalia).

licinia (Trimen), nec Mabille, 1887.

\*Virachola dinochares rhodesiensis (Stevenson), 1937, Occ. Pap. natn. Mus. Sth. Rhod. 6: 21.

\*Virachola dinomenes (Smith), 1887.

\*Virachola diocles diocles (Hewitson), 1869.

Virachola diocles vosseleri (Strand), 1911.

Virachola diomedes Jackson, 1965 Ann. Mag. nat. Hist. (13) 8:528, fig.

Virachola diopolis (Hewitson), see wardii.

Virachola dohertyi (Bethune Baker), see ecaudata.

\*Virachola ecaudata (Gifford), 1963, Entomologist, 96: 43. dohertyi (Bethune Baker), 1905 (homonym).

Virachola edwardsi Gabriel, 1939, Ruwenzori Exp.: 74, fig.

\*Virachola galathea (Swainson), 1821–22.

Virachola gambius Mabille, see antalus.

- \*Virachola jacksoni Talbot, 1935, Entomologist's mon. Mag.  $71:206\ (3)$ ; Passkewsky, 1937, Bull. Soc. ent. Fr.  $42:106\ (9)$ .
- \*Virachola kayonza Stempffer, 1956: 31, figs.

Virachola licinia (Mabille), see batikeli.

Virachola licinia (Trimen), nec Mabille, see dinochares.

- \*Virachola livia (Klug), 1834. Figs. Aurivillius in Seitz, 1921; Stempffer, 1938, Mission Omo 4: 178, (genitalia).
- \*Virachola lorisona lorisona (Hewitson), 1863 (♂).

bimaculata (Hewitson), 1874 ( $\mathcal{P}$ ), fig. Hewitson, 1878; alticola (Aurivillius), 1923 ( $\mathcal{P}$ ).

Virachola lorisona f. albifrons, Stempffer, 1948, Revue fr. Ent. 15: 185.

Virachola lorisona f. immaculata, (Hawker Smith), 1928, Revue Zool. Bot. afr. 16: 215.

Virachola lorisona f. obliterata, (Hawker Smith), 1928, Revue Zool. Bot. afr. 16: 215.

Virachola lorisona coffea Jackson, 1965, l.c.: 529, fig.

- \*Virachola lorisona sesse Stempffer & Jackson, 1962, Proc. R. ent. Soc. Lond. (B) 31: 34.
- \*Virachola magda Gifford, 1963, Entomologist 96: 46.

Virachola nicephora (Hulstaert), 1924, Revue zool. afr. 12: 122. Fig. genitalia, Gifford, 1963, Entomologist 96: 44.

\*Virachola odana (H. H. Druce), 1887. Fig. Seitz, 1921 (3), Bethune Baker, 1921 (2).

\*Virachola penningtoni (van Son), 1949, Annls Transv. Mus. 21:213, fig.

Virachola renidens (Mabille), 1884. Fig. Lathy, 1926, Lepidoptera 2:35.

Virachola rutila (Mabille), see wardii.

Virachola rutilans (Mabille), see wardii.

- \*Virachola suk Stempffer, 1948, Revue fr. Ent. 15: 186, fig.
- \*Virachola vansomereni Stempffer, 1951, Bull. Soc. ent. Fr. 56: 120, fig. Fig. Stempffer, 1952, Bull. Soc. ent. Fr. 57, pl. 1.
- \*Virachola vansoni (Pennington), 1948, J. ent. Soc. sth. Afr. 10: 165, fig. Fig. genitalia, Gifford, 1963, Entomologist 96: 44.
- \*Virachola wardii (Mabille), 1878.

diopolis (Hewitson), 1878; rutila (Mabille), 1878; rutilans (Mabille), 1885.

#### Genus MYRINA Fabricius

Myrina Fabricius, 1807, Illiger Mag. 6: 286; Aurivillius, 1898: 310; 1922: 377; Murray, 1935: 65; Pinhey, 1949: 99; Swanepoel, 1953: 169. Type-species: Papilio alcides Cramer, selected by Kirby, 1870, J. linn. Soc. Lond. (Zool.) 10: 500.

Eyes naked; frons clothed with scales and short hairs; palpi very long, parallel, densely clothed with adpressed scales and hair, second segment long and robust, slightly ascending, third segment conical, horizontal; antennae short, thick, without a distinct club, bluntended; thorax long, robust, the patagiae long and hairy;  $\beta$  fore leg strong, femur robust, hairy, tibia shorter than femur, tarsus unsegmented, finely spinose below; mid and hind legs robust, femora hairy, tibiae shorter than femora, tarsi longer than tibiae, spiny beneath, the metatarsus half the length of the whole tarsus.

Wing shape. Hind wing oval, drawn out at the anal angle, outer margin convex, a long spatulate tail at the end of vein 1b slightly twisted, and with a lobe at the base on its abdominal margin.

Wing venation (Text-fig. 283).

Male genitalia (Text-fig. 100). Uncus shaped like a wide flattened triangle with the posterior edge as its base, this edge bearing in the middle three pointed processes, the whole resembling a trident; subunci long, curved, rather slender; tegumen large; in the natural position uncus and tegumen together take the shape of a hood; vinculum narrow ventrally, without a saccus; inferior fultura shaped like a furca with long, curved arms; valves oblong, small compared with the dorsal structures, their upper process with a serrated apex, and folded inwards; penis elongate, slightly curving, open dorsally in its internal portion, external portion very short; uncus and distal portion of the valves finely pilose.

As I have pointed out (1943, Ann. Soc. ent. Fr. 1942: 117) the male genitalia of M. ficedula are identical with those of M. silenus and it is probable that ficedula is a subspecies of silenus.

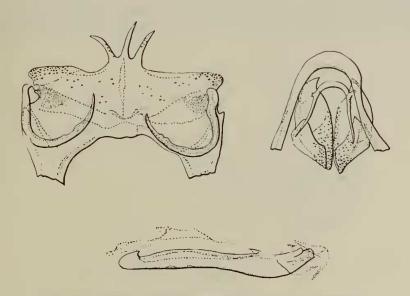


Fig. 100. Myrina silenus silenus (Fabricius), & genitalia.

M. dermaptera, male genitalia (Text-fig. 101). Uncus well developed, trapezoidal, posterior edge deeply notched; subunci long, curved, uniformly tapered; tegumen large, united to the uncus by a lightly chitinized zone; vinculum broad; lower fultura with small curved arms; valves small compared with the dorsal structures, oblong, the edge of the lower process slightly serrated before the apex which has the shape of an open hook; penis as in M. silenus; a few hairs on the uncus and on the distal portion of the valves. The male genitalia of M. subornata are identical with those of M. dermaptera; again I think that they are perhaps two races of the same species. The armature of M. sharpei differs from that of dermaptera only in its shorter and more curved penis.

The early stages of *Myrina* are fairly well known. The caterpillars of *M. silenus* and *M. subornata* have been described by Farquharson (1911, *Trans. ent. Soc. Lond.* 1911: 99); Lamborn (1913, *Trans. ent. Soc. Lond.* 1913: 472) and by Jackson (1937, *Trans. R. ent. Soc. Lond.* 86: 212); those of *M. ficedula* and *M. dermaptera* by Murray (1935: 14, 66, fig. 18) and Pinhey (1949: 101). They feed on wild and cultivated *Ficus*; they possess dorsal glands and are tended by ants of the genera *Camponotus* and *Pheidole*.

## LIST OF SPECIES OF Myrina

Myrina alcides (Cramer), see silenus.

Myrina anettae Fleury, 1924, Bull. Soc. ent. Fr. 1924: 161.

\*Myrina dermaptera dermaptera (Wallengren), 1857. Fig. Hewitson, 1863. Myrina dermaptera nyasae Talbot, 1935, Entomologist's mon. Mag. 71: 78.

\*Myrina silenus silenus (Fabricius), 1775. Fig. Aurivillius in Seitz, 1922. alcides (Cramer), 1776.

Myrina silenus ab. corax (Cramer), 1781.

\*Myrina silenus fidecula Trimen, 1879. Fig. Aurivillius in Seitz, 1922.

Myrina silenus nzoiae Stoneham, 1937, Bull. Stoneham Mus. 34: 2.

\*Myrina sharpei sharpei Bethune Baker, 1906. Fig. Aurivillius in Seitz, 1922.

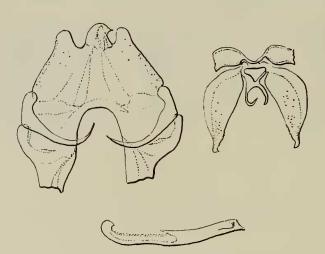


Fig. 101. Myrina dermaptera (Wallengren), & genitalia.

\*Myrina sharpei fontainei Stempffer, 1961: 54, fig.

\*Myrina subornata subornata Lathy, 1903.

Myrina subornata nuba Talbot, 1935 Entomologist's mon. Mag. 71: 78.

#### Genus PSEUDALETIS H. H. Druce

Pseudaletis H. H. Druce, 1888, Entomologist's mon. Mag. 24: 259; Aurivillius, 1898: 311; 1922: 378. Type-species: Pseudaletis agrippina H. H. Druce, by original designation.

Eyes naked; palpi extremely small, hardly visible, clothed with scales; antennae rather short, robust, swelling gradually to a poorly differentiated club; thorax robust; abdomen long, in the  $\varphi$  apically swollen and bearing a dense tuft of hairs, which are probably of use to cover the eggs when laid;  $\varphi$  fore tarsus unsegmented.

Wing shape. Fore wing proportionally more developed than the hind wing; hind wing oval,

produced at the anal angle, a delicate tail at the end of each of the veins 2 and 1b.

Wing-venation (Text-fig. 284). The venation is not entirely uniform throughout the genus. In *clymenus* veins 3 and 4 of the hind wing are briefly stalked and 7 arises from the upper angle of the cell.

Male genitalia (Text-fig. 102). Uncus composed of four digitate processes with rounded apices; no subunci; tegumen subrectangular, in its normal position hood-shaped, so that the two side processes of the uncus are below the middle processes when the genitalia in situ are looked at from the side; vinculum wide, prolonged to form a rounded saccus; lower fultura composed of two subtriangular pieces fused to the middle of the valves; valves oblong with blunt apices, the upper processes being connected in the middle of the inner surface by a membrane which lies above the penis; penis very swollen, bulbous at the base, the external part subcylindrical and bearing on its dorsal surface an irregular prominence; vesica with many fine cornuti; genitalia almost bare, just a few hairs on the uncus and the apices of the valves.

Owing to the scarcity of specimens of *Pseudaletis* in most European collections, I have had no opportunity of examining more than the six species indicated below. In all these species the male genitalia are the same as in *agrippina*. The processes forming the uncus vary a little in size and are sometimes tapered, as in *clymenus*, but

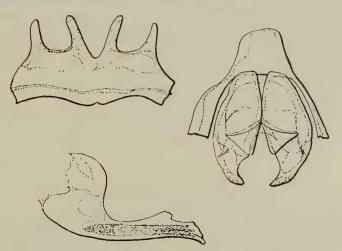


Fig 102. Pseudaletis agrippina Druce, & genitalia.

the valves and penis are closely similar. The genus *Pseudaletis* seems very homogeneous. Although the dorsal structures are very characteristic, the form of the penis, the lower fultura and the valves seems to indicate an affinity with the group containing *Aphnaeus*, *Spindasis* etc. Indeed, Druce, who had not examined the genitalia, began his generic description with the words "allied to *Spindasis*". The view held by Aurivillius, who placed *Pseudaletis* amongst the undoubted Theclinae, seems to me to be erroneous.

#### LIST OF SPECIES OF Pseudaletis

\*Pseudaletis agrippina H. H. Druce, 1888. Fig. Staudinger, 1891 (as tricolor).

Pseudaletis angustimargo Hawker Smith, 1926, Revue zool. afr. 14: 240.

Pseudaletis antimachus (Staudinger), 1888.

Pseudaletis arrhon H. H. Druce, 1913.

Pseudaletis batesi H. H. Druce, 1910.

Pseudaletis busoga van Someren, 1939, Jl E. Africa Uganda nat. Hist. Soc. 14: 174, fig.

Pseudaletis catori Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 393.

\*Pseudaletis clymenus (H. H. Druce), 1885. Fig. Holland, 1893. zebra Holland, 1891.

Pseudaletis clymenus subangulata Talbot, 1935, Entomologist's mon. Mag. 71: 115, figs.

Pseudaletis dardanella Riley, 1922, Entomologist 55: 251.

\*Pseudaletis leonis (Staudinger), 1888. Fig. Aurivillius in Seitz, 1922. trifasciata Sharpe, 1890.

Pseudaletis lusambo Stempffer, 1961: 61, fig.

Pseudaletis mazanguli Neave, 1910.

- \*Pseudaletis nigra nigra Holland, 1892. Fig. Holland, 1893. occidentalis Bethune Baker, 1926.
- \*Pseudaletis nigra fontainei Stempffer, 1961: 60, figs. Pseudaletis occidentalis Bethune Baker, see nigra.
- \*Pseudaletis richardi Stempffer, 1952, Mém. Inst. fr. Afr. noire 19: 146, fig.

Pseudaletis spolia Riley, 1922, Entomologist 55: 250.

Pseudaletis tricolor (Staudinger), see agrippina.

Pseudaletis trifasciata Sharpe, see leonis.

\*Pseudaletis ugandae Riley, 1928, Entomologist 61: 187.

Pseudaletis zebra Holland, see clymenus.

#### Genus OXYLIDES Hübner

Oxylides Hübner, 1816, Verzeichnis: 77. Type-species: Hübner, without designating a type-species, included two species in his genus Oxylides, viz: Papilio celmus Cramer and P. faunus Drury. Of these, Papilio faunus was selected as type of the genus Oxylides by Scudder, 1875, Proc. Am. Acad. Arts Sci. 10: 234.

Oxylides Hübner (partim); Aurivillius, 1898: 312; 1922: 380.

Eves naked; palpi only slightly protruding beyond the frons, second segment laterally compressed and clothed below with adpressed scales, third segment slender, acuminate; antennae slender, about half as long as the costa, white-annulated, club very slightly swollen and with pointed apex; thorax moderately robust; abdomen short, clothed ventrally with silky, white hairs; legs slender, black and white-annulated, of fore tarsus unsegmented.

Wing shape. Fore wing short and broad, costa very convex, outer margin very convex, inner margin straight; hind wing outer margin slightly toothed at the end of veins 6, 5 and 4; three unequal tails, one each at the ends of veins 3, 2 and 1b, the one at vein 2 much longer than the others.

Wing venation (Text-fig. 285). Fore wing with only 10 veins.

Mule genitalia (Text-fig. 103). Uncus composed of two subtriangular lobes separated by the depression of the posterior margin of the tegumen; subunci extremely robust, only slightly curved, and bearing on the lower edge of its distal third a well marked tooth; tegumen rather large; vinculum broad dorsally, narrow ventrally and prolonged to form a long tapering saccus; lower fultura composed of a small lamella with rounded apex; valves small, oblong, with slightly spatulate apices; penis elongate, slightly curved, inner part swollen, outer part tapering gradually; a few hairs on the uncus and apices of the valves.

The male genitalia of bella and gloveri are very similar to those of faunus.

## LIST OF SPECIES OF Oxylides

\*Oxylides bella Aurivillius, 1898.

\*Oxylides faunus faunus (Drury), 1773.

hesiodus (Fabricius), 1793.

\*Oxylides faunus albata (Aurivillius), 1895.

feminina Sharpe, 1904.

Oxylides faunus ab. Q caerulescens Aurivillius, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910-11, 1:1121.

Oxylides feminina Sharpe, see faunus albata.

Oxylides gloveri Hawker Smith, 1929, Bull. Hill Mus. Witley 3: 234.

Oxylides hesiodus (Fabricius), see faunus.

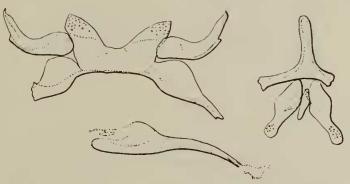


Fig. 103. Oxylides faunus faunus (Drury), & genitalia.

#### Genus SYRMOPTERA Karsch

Syrmoptera Karsch, 1895, Ent. Nachr. 21: 308. Type-species: Syrmoptera melanomitra Karsch, by original designation.

Oxylides (partim); Aurivillius (nec Hübner), 1898: 312; 1922: 381.

Eyes naked; palpi distinctly protruding beyond the frons, second segment laterally compressed, clothed with large white scales, third segment fairly long, acuminate; antennae about half as long as the costa, black and white-annulated, each segment much longer than wide, a well differentiated cylindrical club; thorax clothed below with white silky hairs; legs: tibia and tarsi black and white-annulated,  $\delta$  fore leg with femur clothed with white hairs, tibia shorter than femur, tarsus unsegmented.

Wing shape. Fore wing more triangular than in Oxylides, the apex less rounded; hind wing with three long slender tails, one each at the ends of veins 3, 2 and 1b, those at 2 and 1b subequal, the one at the end of 3 decidedly the shortest.

Wing venation (Text-fig. 286). Fore wing with only 10 veins.

Male genitalia (Text-fig. 104). Uncus composed of two subtriangular lobes; subunci very robust, curved, without apical hook, on the lower edge in the distal third with a strong, blunt tooth; tegumen very large, posterior edge with a rounded depression; vinculum fairly broad prolonged to form a large saccus; lower fultura representated by a small lamella hollowed out at the apex; valves much reduced in size, broadly fused to the vinculum and ending in a small rounded lobe; penis elongate, proximally swollen and flask-shaped, cylindrical and slightly curved externally with a slightly dilated apex; a few hairs on the uncus and on the apices of the valves.

The male genitalia of *S. homeyeri*, *S. amasa*, *S. nivea* and *S. bonifacei* are almost identical with those of *S. melanomitra*. Only the form of the valves differs from one to another.

The genus *Syrmoptera* is very close to the genus *Oxylides*, the only difference being in the shape and colour of the wings.

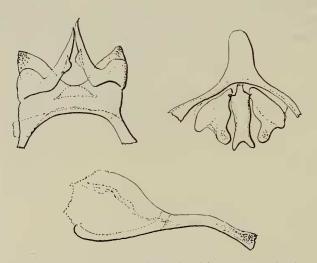


Fig. 104. Syrmoptera melanomitra Karsch, & genitalia.

## LIST OF SPECIES OF Syrmoptera

- \*Syrmoptera amasa (Hewitson), 1869, (2). Fig. Stempffer, 1961: 56 (3).
- \*Syrmoptera bonifacei Stempffer, 1961:57.
- \*Syrmoptera homeyeri (Dewitz), 1879. mixtura Hulstaert, 1924.
- \*Syrmoptera melanomitra Karsch, 1895. Fig. H. H. Druce, 1910.

Syrmoptera mixtura Hulstaert, see homeyeri.

\*Syrmoptera nivea Joicey & Talbot, 1924, Bull. Hill Mus. Willey 1:544. Syrmoptera nivea Q-f. androgyna Joicey & Talbot, 1924, l.c.:545.

#### Genus HYPOLYCAENA Felder

Hypolycaena Felder, 1862, Wien. ent. Monats. 6: 293. Type-species: Myrina sipylus Felder, 1860, selected by Scudder, 1875.

Hypolycaena Felder (partim); Aurivillius, 1898; 313; 1922: 381; 1923: 385; Murray, 1935: 70; Pinhey, 1949: 101; Swanepoel, 1953: 180.

Zeltus de Nicéville, 1890, Butt. India 3: 19, 399.

Eyes densely clothed with short hair; palpi long, ascending, slightly divergent, the second segment laterally compressed and clothed with long erect scales, third segment almost as long as the second, delicate, with sharp apex; antennae long and slender, white-ringed, each segment much longer than wide, club elongate, well defined; of fore leg rather long, the femur clothed in long white hair, tibia and tarsus also white-ringed, the latter unsegmented; mid and hind legs with femora clothed in long white hair, tibiae white-ringed and shorter than the femora, first segment of tarsus very long.

Wing shape. The shape of the hind wing is not uniform. According to it the species may be arranged in three groups, as follows:

- a. tail at vein 1b only slightly longer than that at vein 2; outer margin angled at vein 3: H. auricostalis, H. philippus, H. pachalica.
- b. tail at 1b much longer than tail at 2; outer margin angled at vein 3:
  H. sipylus, II. hatita, II. nigra, II. buxtoni, II. amanica, H. jacksoni.
- c. tail at 1b very long, sometimes longer than the wing; a short tail at vein 3: II. antifaunus, II. lebona, II. dubia, II. naara, II. liara.

For the Indo-Malayan *H. etolus* Fabricius, which has a very long tail at 1b, de Nicéville erected the genus *Zeltus*, which has been used by some authors for such African species as *lebona* and *antifaunus* which have very long tails. But as Aurivillius quite rightly pointed out (1898: 313), *naara* and *liara* are intermediate in this respect between *etolus* and the species of the *philippus* group. It seems to me therefore that no useful purpose is served by cutting up the genus *Hypolycaena*, which is homogeneous both in wing venation and male genitalia.

Butler (1887: 572) introduced the generic name *Tatura* initially for *lebona* only. Later (1896: 123) he also included the African species *buxtoni* and *caeculus*. He gave no generic description. Nevertheless this action, "indicating" a recognizable known species, taken at that date, renders the name *Tatura* available. However, the name has been used by no subsequent author, and *lebona* (its type-species) cannot be separated from other species of *Hypolycaena* either on venation or male genitalia, so it is here treated as a subjective synonym of *Hypolycaena*.

Wing venation (Text-fig. 287, Hypolycaena sipylus). Fore wing with only 10 veins.

Male genitalia (Text-fig. 105). Uncus formed of two roughly oval lobes; subunci long and slender, evenly curved; tegumen formed by a simple enlargement of the vinculum; in the natural position the tegumen and uncus form a hood-like structure; vinculum very wide, with a short rounded saccus; inferior fultura absent; valves small in relation to the dorsal elements, elongate with a slightly recurved tip, tapering, the upper margin bearing a rounded expansion, which represents the second point of attachment, and joined to the vinculum by a membrane; penis elongate, basally swollen, dilated distally, vesica enclosing very small cornuti which give it a shagreened appearance; uncus and distal portion of valves pilose.

The male genitalia of the African species of *Hypolycaena* are of the same pattern as those of *H. sipylus*, which is an Indo-Malayan species. The valves are always rather small in relation to the dorsal elements, but they are generally less elongate and more massive than in *sipylus*; the lobes of the uncus are oval, with rounded apices; the subunci are of the *sipylus* type in *pachalica*, *antifaunus*, *lebona* and *liara*; in *naara* they bear a small apophysis at the point of maximum curvature, and the apophysis is much more developed in *hatita*, *buxtoni*, *amanica*, *jacksoni*, *nigra*, *kadiskos*, *philippus*, and *auricostalis*. However, I do not attach any considerable taxonomic importance to the presence or absence of this apophysis, which also occurs in certain species of *Deudorix*. I figure it only in *antifaunus* (Text-fig. 106) and *nigra* (Text-fig. 107).

As I have pointed out in my original description, ogadenensis has male genitalia of a type very different from those of the other species of this genus. I have only placed this species in the genus Hypolycaena temporarily, on the basis of its wing venation, until I can study further material.

The larvae of nigra, philippus and lebona have been described by Lamborn (1911, Proc. ent. Soc. Lond. (C), and 1913, Trans. ent. Soc. Lond. 1913: 473), by Jackson (1937, Trans. R. ent. Soc. Lond. 86: 213) and by Pinhey (1949: 101). They are onisciform and are tended by ants of the genera Pheidole and Camponotus.

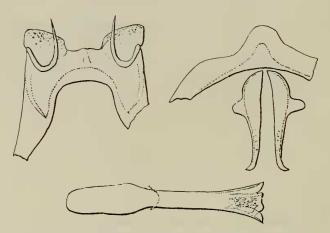


Fig. 105. Hypolycaena sipylus (Felder), & genitalia.

## LIST OF SPECIES OF Hypolycaena

\*Hypolycaena amanica Stempffer, 1951, Bull. Soc. ent. Fr. 56: 123, fig.

\*Hypolycaena antifaunus antifaunus (Doubleday & Hewitson), 1852.

Hypolycaena antifaunus ab. latefasciata Dufrane, 1953, Bull. Annls Soc. R. ent. Belg. 89: 52.

Hypolycaena antifaunus latimaculata (Joicey & Talbot), 1921, Bull. Hill Mus. Witley 1: 94, fig.

Hypolycaena aureolineata Bethune Baker, see pachalica.

\*Hypolycaena auricostalis auricostalis (Butler), 1897. sebasta Hulstaert, 1924.

Hypolycaena auricostalis frommi Strand, 1911. Fig. Aurivillius in Seitz, 1922.

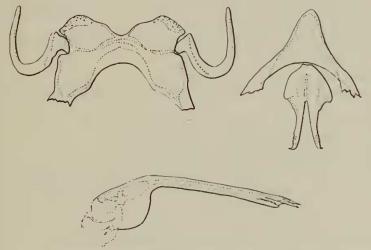


Fig. 106. Hypolycaena antifaunus (Doubleday & Hewitson), & genitalia.

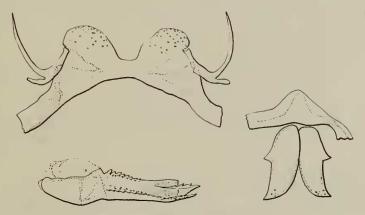


Fig. 107. Hypolycaena nigra Bethune Baker, of genitalia.

Hypolycaena bitjeana Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 394.

\*Hypolycaena buxtoni buxtoni Hewitson, 1874.

seamani Trimen, 1874.

**Hypolycaena buxtoni** Q-f. **divisa** Talbot, 1935, Entomologist's mon. Mag. **71**: 116, fig.

\*Hypolycaena buxtoni rogersi Bethune Baker, 1924, Ann. Mag. nat. Hist. (9) 14:131.

\*Hypolycaena buxtoni spurcus Talbot, 1929, Bull. Hill Mus. Witley 3: 141.

\*Hypolycaena condamini Stempffer, 1956, Mém. Inst. fr. Afr. noire, 48: 207, fig.

Hypolycaena dubia Aurivillius, 1895. Fig. Aurivillius in Seitz, 1922.

Hypolycaena erylus Trimen, see philippus.

\*Hypolycaena hatita hatita Hewitson, 1865.

\*Hypolycaena hatita japhusa Riley, 1921, Trans. ent. Soc. Lond. 1921: 246, fig.

\*Hypolycaena hatita ugandae Sharpe, 1904.

\*Hypolycaena jacksoni Bethune Baker, 1906. Fig. Joicey & Talbot, 1921.

\*Hypolycaena kadiskos H. H. Druce, 1890.

\*Hypolycaena lebona lebona Hewitson, 1865.

Hypolycaena lebona ab. anomale Dufrane, 1953, Bull. Annls R. Soc. ent. Belg. 89: 52.

Hypolycaena lebona ab. splendens Dufrane, 1953, l.c.: 52.

\*Hypolycaena lebona caerulea Aurivillius, 1895.

\*Hypolycaena lebona scintillans Aurivillius, 1895.

\*Hypolycaena liara liara H. H. Druce, 1890. Fig. Aurivillius in Seitz, 1922. naara Karsch, 1893 (nec Hewitson, 1873); symmacha Hulstaert, 1924.

\*Hypolycaena liara obscura Stempffer, 1947, Bull. Soc. ent. Fr. 52: 37.

Hypolycaena liara plana Talbot, 1935, Entomologist's mon. Mag. 71: 116, figs.

\*Hypolycaena naara Hewitson, 1873. Fig. Aurivillius in Seitz, 1922.

Hypolycaena naara (Karsch) (nec Hewitson, 1873), see liara.

\*Hypolycaena nigra Bethune Baker, 1913. Fig. Aurivillius in Seitz, 1922.

\*Hypolycaena ogadenensis Stempffer, 1946, Revue fr. ent. 13: 10, fig.

Hypolycaena orejus (Hopffer), see philippus.

\*Hypolycaena pachalica Butler, 1888. aureolineata Bethune Baker, 1906.

\*Hypolycaena philippus philippus (Fabricius), 1793. Fig. Hewitson, 1865. orejus (Hopffer), 1855; erylus (Trimen), 1866.

\*Hypolycaena philippus ramonza (Saalmuller) 1878.

Hypolycaena philippus ramonza ab. vittigera Mabille, 1879.

Hypolycaena schubotzi Aurivillius, 1923, Ergebn. 2te D. Zent. Exp. 1910-11, 1: 1213.

Hypolycaena seamani Trimen, see buxtoni.

Hypolycaena sebasta Hulstaert, see auricostalis.

Hypolycaena similis Dufrane, 1945, Bull. Annls Soc. R. ent. Belg. 81: 118.

Hypolycaena symmacha Hulstaert, see liara.

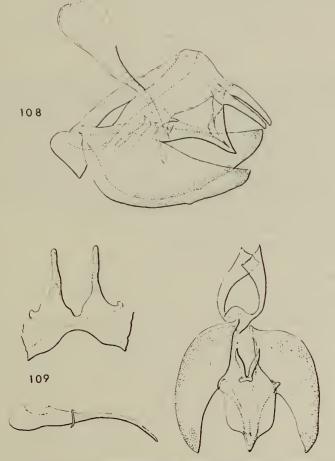
#### Genus DAPIDODIGMA Karsch

Dapidodigma Karsch, 1895, Ent. Nachr. 21: 310; Aurivillius, 1898: 318; 1923: 389. Typespecies: Papilio hymen Fabricius, 1775 (Papilio liger Cramer, 1782) by original designation.

Eyes smooth; vertex of head clothed with erect hairs; palpi protruding well beyond the frons, second segment very long, laterally compressed and clothed with short hairs, third segment reduced, acuminate; antennae, which are a little less than half the length of the costa, with short segments gradually getting stouter and ending in a poorly differentiated cylindrical club; of fore tarsus unsegmented and clothed with very long hairs almost up to its extremity.

Wing shape. Hind wing oval, produced at the anal angle, outer margin angled at the end of vein 4, concave between veins 4 and 3, and with three delicate tails, all about equal in length, at the ends of veins 3, 2 and 1b, a small anal lobe. Male secondary sexual character, a large very pale sulphur-yellow androconial patch covering the whole cell and the bases of the interspaces 2 to 5.

Wing venation (Text-fig. 288).



Figs 108-109. Dapidodigma hymen (Fabricius), & genitalia.

Male genitalia (Text-fig. 108, lateral view; Text-fig. 109, ventral view). Uncus composed of two long digitate processes with rounded apices; no subunci; tegumen deeply notched on its posterior margin; vinculum fairly broad; valves oblong, the upper edges almost straight, the lower ones convex; lower fultura composed of two small divergent arms fused basally to the base of the valves; in addition, there is an offshoot, which I consider an upper fultura, articulated at about half-way along the upper edge of the valves, where it is slightly notched, connected with the edge of the tegumen and entirely surrounding the penis; penis very elongate, swollen at the base, the external portion tapering regularly and slightly curving; there are a few fine, short hairs on the uncus; the valves are densely hairy, especially on the lower edges and apices.

The male genitalia of D. demeter Clench are identical with those of D. hymen.

# LIST OF SPECIES OF Dapidodigma

\*Dapidodigma demeter demeter Clench, 1961, Ann. Carneg. Mus. 36: 64, figs. Dapidodigma demeter nuptus Clench, 1961, l.c.: 65, fig.

\*Dapidodigma hymen (Fabricius), 1715. Fig. Cramer, 1779, (as liger).

#### Genus IOLAUS Hübner

When I undertook the revision of the "Iolaus complex", in collaboration with Neville Bennett, I found myself faced with considerable difficulties. The group is so remarkably uniform in external appearance that its species are recognisable at a glance; the upperside is nearly always blue, the underside pure white with a pattern of markings reduced to delicate postdiscal and antemarginal lines, and the hind wing carries delicate tails at the ends of veins 2 and 1b. But in respect of venation and male genitalia this uniformity is entirely lacking.

The genera hitherto widely employed within the *Iolaus* group have been based in the main on the venation, and can be grouped as follows:—-

- A. Fore wing with 10 veins in both sexes:—

  Hemiolaus, Stugeta, Pseudiolaus.
- B. Fore wing with 11 veins in both sexes:—

  Tanuetheira, Aphniolaus, Iolaus (sensu stricto), Epamera, Sukidion.
- C. Fore wing with 12 veins in the male, 11 in female:—

  Trichiolaus, Argiolaus.

Some of these genera seem to me to have been founded on characters of secondary importance, for example, *Trichiolaus* on the hairiness of the eyes, *Tanuetheira* on the great length of the tail, *Aphniolaus* on its yellow coloration; and it is impossible on any sound criterion to separate *Iolaus* (sensu stricto) from *Epamera*.

However, examination of the morphology of the male genitalia demonstrates that the great majority of species forming the *Iolaus* group have an armature of Thecline type, that is to say, with the dorsal elements forming a kind of hood with well developed subunci. Yet some others (e.g. *Argiolaus*) display a very different structure: in the *silas* group, the uncus is divided into two long sharp points, the subunci are rudimentary and the tegumen reduced; *parasilanus* has a pseudotergum

quite distinct from the tegumen; and in *catori* the very large dorsal elements are distally serrate and devoid of subunci.

Had we based our classification solely on the male genitalia, it would have been possible to devise genera wholly homogeneous in respect of this character, yet decidedly heterogeneous in respect of the characters of the venation and external features. We therefore adopted a system which is pragmatic rather than strictly logical, by retaining the genus *Iolaus* as a kind of "super-genus", by erecting subgenera for the species having genitalia profoundly different from the main bulk of the species, and by retaining the generic names hitherto in use, but treating them as subgenera. In this way it has resulted that within each subgenus there is both uniformity of venation and uniformity of genitalia. These characters are indicated in each of the subgenera adopted.

### IOLAUS (HEMIOLAUS) Aurivillius

Hemiolaus Aurivillius, 1923; 386; Pinhey, 1949; 102. Type-species; Iolaus caeculus Hopffer, 1855, designated by Stempffer & Bennett, 1958; 1254.

Hypolycaena Felder (partim); Aurivillius, 1898: 316; Murray, 1935: 69; Swanepoel, 1953: 180.

Eyes smooth; from reddish with two white side stripes and a dark depressed median line; palpi reaching well beyond the from, third segment slender, acuminate; antennae thin, 4/7th length of costa, each segment much longer than wide; club gently swollen; legs as in Hypolycaena, 3 fore leg unsegmented.

Wing shape. Fore wing inner margin of 3 with a large lobe near the base; hind wing outer margin excised between the ends of veins 3 and 4, a delicate tail at the end of vein 2 and a much longer one at the end of 1b, a small lobe at the anal angle. Male secondary sexual characters; on the upperside of the hind wing a gleaming bronze patch covers the base of the wing and the greater part of the cell, the upper part of this patch being covered by the lobe of the fore wing; between the lower edge of the cell and the basal part of vein 1b there is a tuft of long adpressed hairs facing towards the costa, and this is also partially covered by the lobe of the fore wing; on the underside of the fore wing, along the inner margin, a nacreous zone, but no hair-tuft.

Wing venation (Text-fig. 289). Fore wing with only 10 veins.

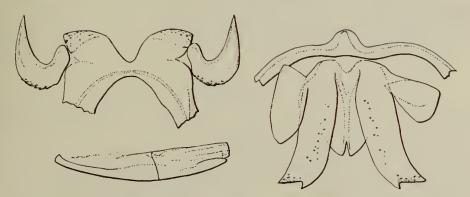


Fig. 110. lolaus (Hemiolaus) coeculus coeculus Hopffer, of genitalia.

Male genitalia (Text-fig. 110). Uncus composed of two suboval lobes slightly angled apically and separated by the deep depression of the posterior edge of the tegumen; subunci curved, very stout in the middle, the lower edge bearing short spines in its basal third; tegumen large, hood-shaped; vinculum narrow dorsally and having a weak saccus; lower fultura strongly developed and with two wide flattened arms which are almost wholly fused together; valves composed of two lobes, the upper one oval, semi-membranous and, on this account, often distorted when the genitalia are mounted for the microscope, the lower process shaped as an elongate rectangle with the apex excised; penis elongate, widely open dorsally, distally clubshaped; uncus and apices of valves pilose.

The male genitalia of the other species of *Hemiolaus* are very similar to those of *caeculus*. The shape of the subunci furnishes good specific characters.

## LIST OF SPECIES OF Iolaus (Hemiolaus)

Iolaus (Hemiolaus) ceres (Hewitson), 1865. Fig. genitalia, Stempffer & Bennett, 1958.

maryra mabillei Aurivillius, 1923.

Iolaus (Hemiolaus) cobaltina (Aurivillius), 1898. Fig. Aurivillius in Seitz, 1923; fig. genitalia, Stempffer, 1958.

coeculus (Mabille), nec Hopffer, 1855.

Iolaus (Hemiolaus) coeculus coeculus Hopffer, 1855. Fig. Hopffer, 1862; fig. genitalia, Stempffer, 1938, Mission Omo 4: 184.

Iolaus (Hemiolaus) coeculus f. dolores (Suffert), 1904. Fig. Aurivillius in Seitz, 1923.

Iolaus (Hemiolaus) coeculus f. obscurus (Suffert), 1904.

Iolaus (Hemiolaus) coeculus ab. duponti Dufrane, 1953, Bull. Annls Soc. R. ent. Belg. 89:53.

Iolaus (Hemiolaus) coeculus littoralis Stempffer, 1953, Annls Mus. R. Congo belge 27: 25.

Iolaus (Hemiolaus) coeculus (Mabille), nec Hopffer, see cobaltina.

Iolaus (Hemiolaus) margites (Mabille), 1899.

Iolaus (Hemiolaus) maryra maryra (Mabille), 1855. Fig. genitalia, Stempffer & Bennett, 1958.

Iolaus (Hemiolaus) maryra mabillei Aurivillius, see ceres.

Iolaus (Hemiolaus) varnieri (Stempffer), 1943, Ann. Soc. ent. Fr. 1942: 118. Figs. Stempffer & Bennett, 1958.

# IOLAUS (STUGETA) H. H. Druce

Stugeta H. H. Druce, 1891, Ann. Mag. nat. Hist. (6) 8: 149; Aurivillius, 1898: 317; 1923: 387; Murray, 1935: 71; Pinhey, 1949: 102; Swanepoel, 1953: 179. Type-species: Iolaus bowkeri Trimen, by original designation.

Eyes smooth; palpi protruding far beyond the frons, second segment long, clothed with adpressed scales, third segment much shorter, slender, cylindrical, acuminate; antennae robust, each segment almost as broad as long, club gradually swollen, cylindrical, not well differentiated; legs long and slender,  $\delta$  fore leg with femur hairy, tibia shorter than femur, tarsus unsegmented, finely spinose below.

Wing shape. Hind wing oval, somewhat produced at the anal angle, outer margin slightly undulating between the ends of veins 6 to 3, a delicate tail at the end of vein 2, a much longer one at the end of vein 1b, a small lobe at the anal angle. Abdominal margin excised between the lobe and the end of vein 1a. No secondary male sexual characters.

Wing venation (Text-fig. 290). Fore wing with only 10 veins.

Male genitalia (Text-fig. 111). Uncus composed of two subtriangular lobes with rounded apices; subunci long, evenly curved, tapering gradually to a blunt apex; tegumen large only slightly notched on its posterior margin; vinculum broad above, narrower below; lower fultura articulated to the base of the valves and composed of two curved arms connected at their apices by a membrane, thus forming a complete ring around the penis; valves subrectangular, widened at the apices and bearing on their distal edge two stout teeth of unequal length; penis elongate, curved, the basal portion widely open, the external portion tapering regularly to a blunt apex; vesica with two long, sharp-pointed spines; uncus and distal portion of the valves hairy.

Stugeta bowkeri is remarkable in that some of its geographical races show sharp differences in the terminal contour of the valves (see Stempffer & Bennett, 1958, 20: 1266, figs 14, 15). The male genitalia of the other species of the subgenus are very close to those of bowkeri.

The early stages of S. bowkeri have been described by Murray (1935: 72) and Pinhey (1949: 103); those of S. marmorea olalae by Jackson (1937, Trans. R. ent. Soc. Lond. 86: 213).

## LIST OF SPECIES OF Iolaus (Stugeta)

\*Iolaus (Stugeta) bowkeri bowkeri Trimen, 1864. Fig. Trimen, 1866.

\*Iolaus (Stugeta) bowkeri f. caerulea Stempffer, 1947, Bull. Soc. ent. Fr. 52: 37.

\*Iolaus (Stugeta) bowkeri ethiopica Stempsfer & Bennett, 1958: 1266.

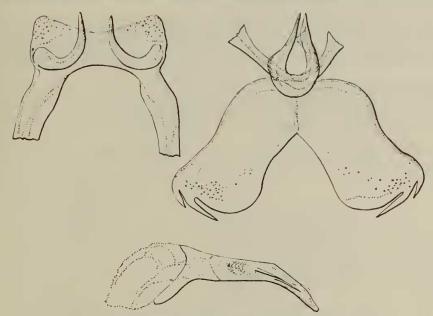


Fig. 111. Iolaus (Stugeta) bowkeri bowkeri Trimen, & genitalia.

- \*Iolaus (Stugeta) bowkeri kedonga van Someren, 1939, Jl E. Africa Uganda nat. hist. Soc. 14: 175, figs.
- \*Iolaus (Stugeta) bowkeri maria Suffert, 1904. Fig. H. H. Druce, 1907; fig. genitalia, Stempffer & Bennett, 1958.
- \*Iolaus (Stugeta) bowkeri mombase Butler, 1901.
- \*Iolaus (Stugeta) bowkeri nyanzana Wichgraft, 1911.
- \*Iolaus (Stugeta) bowkeri nyasana Talbot, 1935, Entomologist's mon. Mag. 71: 116, fig.
- \*Iolaus (Stugeta) bowkeri occidentalis Stempffer & Bennett, 1958: 1268.
- \*Iolaus (Stugeta) bowkeri subinfuscata Grünberg, 1910.
- \*Iolaus (Stugeta) carpenteri Stempffer, 1946, Revue fr. Ent. 13:11. Fig. genitalia, Stempffer & Bennett, 1958.
- \*Iolaus (Stugeta) marmorea marmorea (Butler), 1866. Fig. Aurivillius in Seitz, 1923; fig. genitalia, Stempffer & Bennett, 1958.
- \*Iolaus (Stugeta) marmorea olalae Stoneham, 1933, Bull. Stoneham Mus. 17: 2.
- \*Iolaus (Stugeta) mimetica Aurivillius, 1916. Fig. Stempffer & Bennett, 1958.
- \*Iolaus (Stugeta) somalina Stempffer, 1946, Revue fr. Ent. 13: 13, fig. (2); 1954, Bull. Soc. ent. Fr. 59: 109, fig. (3) and fig. genitalia.

# IOLAUS (PSEUDIOLAUS) Riley

Pseudiolaus Riley, 1928, Novit. 2001. 34: 392; Swanepoel, 1953: 179. Type-species: Pseudiolaus poultoni Riley, by original designation.

Eyes naked; palpi protruding well beyond the frons, second segment rather slender, clothed with adpressed scales, third segment one-third the length of the second, subcylindrical, apex blunt; antennae three-sevenths the length of the costa, club gradually swollen and one-third the total length of the antenna; legs unicolorous white, 3 fore tarsus unsegmented.

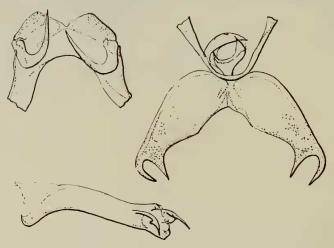


Fig. 112. Iolaus (Pseudiolaus) poultoni Riley, of genitalia.

Wing shape. Fore wing inner margin with a large basal lobe; hind wing outer margin with a marked angle at the end of vein 3, two delicate tails 4–5 mm. long, at the ends of veins 2 and 1b, a small anal lobe. Male secondary sexual characters: on the upperside of the hind wing there is an androconial patch, about 3 mm. wide, at the origin of vein 7, surrounded by a white pearly zone which extends into the cell; on the underside of the fore wing, level with the lobe of the inner margin, there is a tuft of long hairs directed towards the androconial patch on the hind wing.

Wing venation (Text-fig. 291). Fore wing has only 10 veins.

Male genitalia (Text-fig. 112). Uncus composed of two lateral strips separated by the depression of the margin of the tegumen; subunci strongly curved, very robust at the base and then tapering uniformly; tegumen large and hood-shaped; vinculum rather broad above, narrow below; lower fultura pedunculate and bearing a complete ring which encircles the penis; valves oblong, apex divided into two sharp-pointed teeth; penis elongate, robust, slightly curved; vesica enclosing two strong recurved spines; uncus and distal portion of valves densely hairy.

The male genitalia of *lulua* are very similar to those of *poultoni*.

The subgenus *Pseudiolaus* is easily distinguished from the subgenus *Stugeta* by the more slender antennae, and in the male by the presence of secondary sexual characters; the general appearance and wing markings of *Pseudiolaus* remind one of the subgenus *Epamera*.

## LIST OF SPECIES OF Iolaus (Pseudiolaus)

- \*Iolaus (Pseudiolaus) lulua Riley, 1944, Entomologist 77: 28. Fig. genitalia, Stempffer & Bennett, 1958.
- \*Iolaus (Pseudiolaus) poultoni Riley, 1928, Novit. zool. 34: 392, figs (3). Fig.  $\varphi$ , Talbot, 1935, Entomologist's mon. Mag. 71: 117.

## IOLAUS (TRICHIOLAUS) Aurivillius

Trichiolaus Aurivillius, 1898: 317; 1923: 389. Type-species: Hypolycaena mermeros Mabille, designated by Stempffer & Bennett, 1958: 1276.

Eyes densely hairy; palpi extending considerably beyond the frons, second segment very long and clothed with adpressed scales, third segment very short, conical; antennae with a gradually swollen, fusiform club; thorax clothed below with grey silky hairs;  $\beta$  fore leg, femur clothed with long grey hairs, tibia as long as the femur, tarsus unsegmented, robust, finely spinose below.

Wing shape. Hind wing oval, somewhat produced at the anal angle, outer margin slightly angular at the end of vein 4, a short pointed tail at the end of vein 3, a longer delicate tail at the end of vein 2, a still longer tail at the end of vein 1b, a lobe at the anal angle. Male secondary sexual characters absent.

Wing venation (Text-fig. 292, 3). Fore wing, 3 with 12 veins, 9 11.

Male genitalia (Text-fig. 113). Uncus composed of two large semicircular lobes; subunci long and curving; teguinen rather large, but narrow in the median area; vinculum broad dorsally, narrow ventrally; inferior fultura like a furca with divergent arms; valves oblong with rounded apices; penis elongate, widely open on the upper surface of almost the whole of the internal portion, the external portion very short with bulbous apex; vesica appearing shagreened; lobes of uncus and distal portion of valves hairy.

The male genitalia of I. (Trichiolaus) argentarius are almost the same as those of I. (Trichiolaus) mermeros, only the valves are a little narrower.

LIST OF SPECIES OF Iolaus (Trichiolaus)

\*Iolaus (Trichiolaus) argentarius Butler, 1879. Fig. Oberthur, 1916 (as leucoceros).

leucoceros (Oberthur), 1916.

Iolaus (Trichiolaus) leucoceros (Oberthur), see argentarius.

\*Iolaus (Trichiolaus) mermeros (Mabille), 1878. Fig. Mabille, 1887.

## IOLAUS (TANUETHEIRA) H. H. Druce

Tanuetheira H. H. Druce, 1891, Ann. Mag. nat. Hist. (6) 8:148. Type-species: Papilio timon Fabricius, by original designation.

Iolaus Hübner (partim); Aurivillius, 1898: 322; 1923: 391.

Eyes smooth; palpi extending well beyond the frons, second segment long, laterally compressed, clothed below with white adpressed scales and above with black scales, third segment rather short and similarly clothed with white and black scales; antennae about half as long as the costa, with a gradually swollen fusiform club which has a reddish apex; thorax clothed below with long white hairs; all tarsi black and white-annulated, 3 fore tarsus unsegmented.

Wing shape. Fore wing inner margin with slight lobe near the base; hind wing outer margin slightly angular at the end of vein 4, a short triangular tail at the end of vein 3, a long delicate tail at the end of vein 2, a broad tail about 2 mm. wide and 18-20 mm. long at the end of vein 1b, a small lobe at the anal angle. Male secondary sexual characters: on the underside of the fore wing, in the middle of the inner margin, there is a tuft of long black hairs; on the upperside of the hind wing there is a scaly patch at the base of vein 7, surrounded by a wide, glossy, satin-like area which covers a large part of the cell and costal area.

Wing venation (Text-fig. 293). Fore wing with 11 veins in both sexes. H. H. Druce was in error, as Aurivillius noted, in attributing 12 veins to the 3.

Male genitalia (Text-fig. 114). Uncus composed of two lobes with rounded apices separated by the shallow depression of the margin of the tegumen; subunci curved, fairly long with thickened base; tegumen large, subrectangular, in its normal position hood-shaped; vinculum rather broad with a very indistinct saccus; lower fultura shield-shaped, fused to the base of the valves, which are oblong with the apex slightly recurved and a serrated lower edge; penis

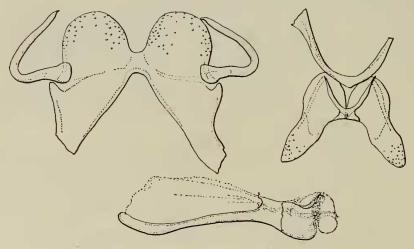


Fig. 113. Iolaus (Trichiolaus) mermeros (Mabille), & genitalia.

long, very robust, slightly swollen at the base with a cylindrical external portion; vesica enclosing two enormous lanceolate spines; uncus lobes clothed with long, fine hairs, apex and lower edge of valvae pilose.

The caterpillar of *T. timon* has been described by Farquharson (1922, *Trans. ent. Soc. Lond.* **1922**: 361). It lives on the flowers of *Loranthus incanus* Schumach (Loranthaceae), a parasite of *Funtumia elastica* (Preuss) Stapf (Apocynaceae).

## LIST OF SPECIES OF Iolaus (Tanuetheira)

- \*Iolaus (Tanuetheira) timon timon (Fabricius), 1787. Fig. Butler, 1870.
- \*Iolaus (Tanuetheira) timon promethus H. H. Druce, 1891. Fig. genitalia, Stempffer & Bennett, 1958.
- \*Iolaus (Tanuetheira) timon congoensis Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1: 90, fig. Fig. genitalia, Stempffer & Bennett, 1958.
- Iolaus (Tanuetheira) timon orientius Hulstaert, 1924, Rev. zool. afr. 12: 126. Fig. genitalia, Stempffer & Bennett, 1958.

### IOLAUS (ARGIOLAUS) H. H. Druce

Argiolaus H. H. Druce, 1891, Ann. Mag. nat. Hist. (6) 8:143; Pinhey, 1949:103. Typespecies: Iolaus silas Westwood, 1852, by original designation.

Iolaus Hübner (partim); Aurivillius, 1898: 324; 1923: 396; Murray, 1935: 74; Swanepoel, 1953: 173.

Eyes smooth; palpi long, parallel, second segment long, slightly ascending, laterally compressed, clothed below with closely packed white scales, third segment short, porrect, with blunt tip; antennae a little longer than half the costa, rather robust, the club cylindrical, not well defined; tibia of 3 fore leg shorter than femur, tarsus unsegmented and bearing delicate spines below.

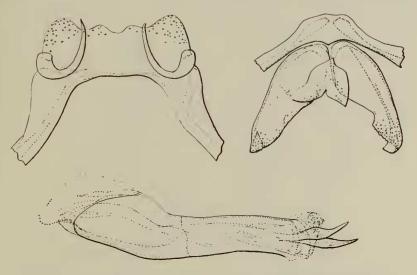


Fig. 114. Iolaus (Tanuetheira) timon orientius Hulstaert, of genitalia.

Wing shape. 3 fore wing inner margin feebly lobed basally: hind wing hind margin with a sharp tooth at end of vein 3, a long and delicate tail at vein 2, and a delicate and much longer tail at 1b; anal angle lobed. Male secondary sexual characters: midway on the inner margin of the fore wing below there is a tuft of yellowish hair directed towards vein 2; on the upperside of the hind wing, between vein 7 and the upper edge of the cell, a greyish oval patch covered with large oval erect scales and surrounded by a wide bronze shining area that fills the cell and is covered with smaller scales.

Wing venation (Text-fig. 294). The 3 fore wing with 12 veins, \$\phi\$ with 11, vein 8 being absent. Male genitalia (Text-fig. 115). Uncus composed of two long sharp, slightly curved points, separate almost to the base; subunci rudimentary, subtriangular and scarcely visible; tegumen reduced; vinculum broad, bearing on each side two large, triangular, weakly sclerotized expansions; in their natural position these expansions are joined to the lower fultura by a membrane; in silas the vinculum ends in a large triangular saccus, which is lacking in the other two species of the sub-genus. Fultura inferior composed of a ring born on a peduncle and surrounding the penis; valves oblong; penis short, massive, widely open above basally; vesica enclosing stout cunei; uncus pilose, upper margin of valves only slightly so.

Among the many species placed in *Argiolaus* by Druce and other authors, only two, *lalos* and *crawshayi* have male genitalia of the *silas* type. In the others they are so different that it has been necessary to arrange them in new subgenera. The area of distribution of *Argiolaus* even in this restricted sense is very extensive, comprising the whole of East and South Africa from Abyssinia to the Cape, and the species break up into more or less sharply defined geographical races, each of which differs within the species not only in external appearance but also in the form of the valves, which show constant modifications.

The early stages of *silas* and *crawshayi* have been described by Murray (1935: 75), Jackson (1937, *Trans. R. ent. Soc. Lond.* 86: 214) and Pinhey (1949: 104). The caterpillars live on the flowers of various kinds of *Loranthus* which are parasitic on trees inhabited by ants of the genus *Crematogaster*.

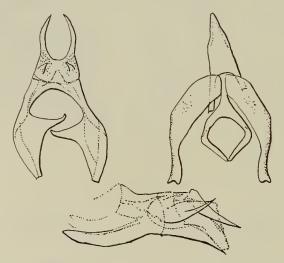


Fig. 115. Iolaus (Argiolaus) silas silas Westwood, & genitalia.

## LIST OF SPECIES OF Iolaus (Argiolaus)

- \*Iolaus (Argiolaus) crawshayi crawshayi Butler, 1900. Fig. genitalia, Stempffer & Bennett, 1958.
- \*Iolaus (Argiolaus) crawshayi elgonae Stempffer & Bennett, 1958: 1291, fig.
- \*Iolaus (Argiolaus) crawshayi littoralis Stempffer & Bennett, 1958: 1295, fig.
- \*Iolaus (Argiolaus) crawshayi maureli Dufrane, 1954, Bull. Annls Soc. R. ent. Belg. 90: 282. Fig. Stempffer & Bennett, 1958: 49.
- \*Iolaus (Argiolaus) crawshayi nyanzae Stempffer & Bennett, 1958: 1291, fig.
- \*Iolaus (Argiolaus) crawshayi niloticus Stempffer & Bennett, 1958: 1293, figs.
- \*Iolaus (Argiolaus) lalos lalos H. H. Druce, 1896. Fig. H. H. Druce, 1902; fig. genitalia, Stempffer & Bennett, 1958.
- \*Iolaus (Argiolaus) lalos kigezi Stempffer & Bennett, 1958: 1287, figs.
- \*Iolaus (Argiolaus) silas silas Westwood, 1852.
- \*Iolaus (Argiolaus) silas ab. lasius Suffert, see silas silarus.
- \*Iolaus (Argiolaus) silas silarus H. H. Druce, 1885. Fig. genitalia, Stempffer & Bennett, 1958.

silas ab. lasius Suffert, 1904.

## IOLAUS (IOLAPHILUS) Stempffer & Bennett

Iolaphilus Stempffer & Bennett, 1958: 1298. Type-species: Iolaus menas H. H. Druce, by original designation.

Iolaus Hübner (partim); Aurivillius, 1898: 323; 1923: 393; Murray, 1935: 75; Swanepoel, 1953: 172.

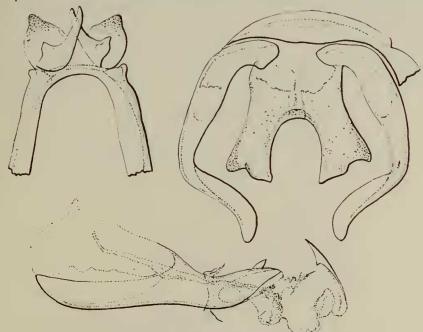


Fig. 116. Iolaus (Iolaphilus) menas Druce, & genitalia.

Eyes, palpi, antennae and legs as in typical Argiolaus.

Wing shape. Fore wing inner margin more or less strongly lobed near the base, weakly in menas, trimeni, maritimus, vansomereni, poecilaon, iturensis, aequatorialis, laonides and ismenias, more sharply in julus, aelianas, calisto and cottrelli. Hind wing with a tooth at vein 3, a delicate tail at the end of vein 2, and a longer one at 1a which, in caesareus, is extremely long and spatulate at the tip; anal angle lobed. Male secondary sexual characters: as in Argiolaus, namely, a hair tuft midway along the inner margin on the underside of the fore wing, and an androconical patch, small to large, on the upperside of the hind wings near the base.

Wing venation. Exactly as in Argiolaus, fore wing with 12 veins in the 3, 11 in the 9.

Male genitalia (Text-fig. 116). Uncus composed of two subtriangular lobes separated by a slight depression of the margin of the tegumen; subunci short, stout, gently curved. Tegumen subrectangular, hood-shaped; vinculum broad; lower fultura formed of two large laminae, broader at their extremities and with their basal halves fused together; valves long, narrow, curving, with rounded apices; penis very stout, more elongate than in Argiolaus, the inner part widely open dorsally, the outer part rather short; vesica enclosing two stout unequal spines: uncus and lower fultura pilose, valves almost naked.

The male genitalia of *menas* are evidently of the ordinary Thecline type, profoundedly different from typical *Argiolaus*. The armatures of the other *Iolaphilus* are certainly not all identical with those of *menas* either, but the general plan is the same. The following differences can be indicated:—

- a. Subunci weakly curved: menas, henryi, trimeni, schultzei, julus, aelianus, iturensis, maritimus, alcibiades, paneperata, lukabas, ndolae, ismenias, vansomereni and aequatorialis.
  - b. Subunci very bent: gabunica, caesareus, calisto, laonides and poecilaon.
- c. Lower fultura composed of large laminae: menas, henryi, trimeni, schultzei, julus, aelianus and gabunica.

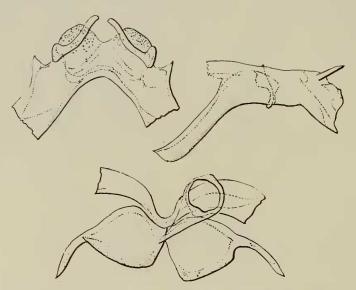


Fig. 117. Iolaus (Iolaphilus) alcibiades Kirby, of genitalia.

- d. Lower fultura in the form of a ring enclosing the penis and arising from a peduncle: iturensis, maritimus, alcibiades, paneperata, lukabas, ndolae, caesareus, ismenias, vansomereni and aequatorialis.
- e. Lower fultura in the form of a furca with delicate divergent branches: laonides and poecilaon.
- f. Uncus bearing horn-like expansions: in *bergeri* the uncus bears four sharp-pointed processes, the outer pair triangular, the inner pair digitate; subunci slender, gently curved; tegumen reduced; vinculum narrow; lower fultura large and crescentic; valves oblong; penis long and slender, strongly curved; uncus, lower margin and apex of valves lightly pilose. In *cottrelli* the uncal expansions are much shorter and the penis is of the *Iolaphilus* type. In *kayonza* the uncus has two expansions only.

The male genitalia of all the known species of *Iolaphilus* having already been figured (1958, Stempffer & Bennett) only those of *alcibiades* (Text-fig. 117), *calisto* (Text-fig. 118), *laonides* (Text-fig. 119) and *bergeri* (Text-fig. 120) are reproduced here.

The early stages of *julus*, *alcibiades* and *paneperata* have been described by Farquharson (1922, *Trans. ent. Soc. Lond.* 1922: 361) and Eltringham (l.c.: 479). Like those of *Argiolaus*, the larvae live on *Loranthus*.

## LIST OF SPECIES OF Iolans (Iolaphilus)

- \*Iolaus (Iolaphilus) aelianus aelianus Staudinger, 1891 (July) (\$\varphi\$). Fig. genitalia, Stempffer & Bennett, 1958.

  jamesoni H. H. Druce, 1891, August (\$\varphi\$).
- \*Iolaus (Iolaphilus) aelianus entebbae Riley, 1928, Novit. zool. 34: 376, fig.
- \*Iolaus (Iolaphilus) aequatorialis Stempffer & Bennett, 1958: 1322, figs.
- \*Iolaus (Iolaphilus) alcibiades, Kirby, 1871. Fig. Aurivillius in Seitz, 1923. julianus Staudinger, 1891.

Iolaus (Iolaphilus) anesius Hulstaert, see trimeni.

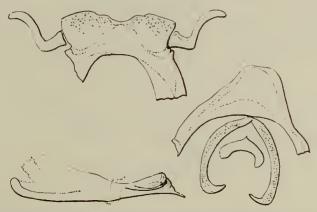


Fig. 118. Iolaus (Iolaphilus) calisto (Doubleday & Hewitson), & genitalia.

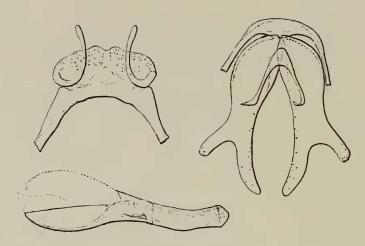


Fig. 119. Iolaus (Iolaphilus) laonides Aurivillius, 3 genitalia of type.

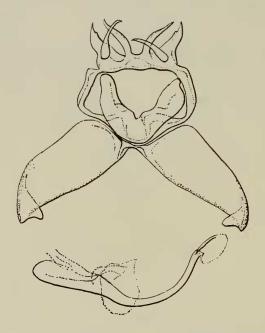


Fig. 120. Iolaus (Iolaphilus) bergeri Stempffer, of genitalia.

- \*Iolaus (Iolaphilus) bergeri Stempffer, 1953, Annls Mus. R. Congo belge 27: 27, fig.
- \*Iolaus (Iolaphilus) caesareus Aurivillius, 1895. Fig. Aurivillius in Seitz, 1923; fig. genitalia, Stempffer & Bennett, 1958.
- \*Iolaus (Iolaphilus) calisto (Doubleday & Hewitson), 1852.
- \*Iolaus (Iolaphilus) cottrelli Stempffer & Bennett, 1958: 1330, figs.

Iolaus (Iolaphilus) elisa Suffert, see paneperata.

\*Iolaus (Iolaphilus) gabunica Riley, 1928, Novit. zool. 34: 375 (3), fig. Fig. genitalia, Stempffer & Bennett, 1958. Stempffer, 1961: 93 fig. (2).

\*Iolaus (Iolaphilus) henryi Stempffer, 1961, l.c.: 88, figs.

- \*Iolaus (Iolaphilus) ismenias ismenias (Klug), 1834. Fig. genitalia, Stempffer & Bennett, 1958.
- \*Iolaus (Iolaphilus) ismenias piaggiae Oberthur, 1883. Fig. genitalia, Stempffer & Bennett, 1958.
- \*Iolaus (Iolaphilus) iturensis Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1:91 (2). Figs Stempffer & Bennett, 1958 (2 and 3 genitalia).

Iolaus (Iolaphilus) jamesoni H. H. Druce, see aelianus.

Iolaus (Iolaphilus) julianus Staudinger, see alcibiades.

Iolaus (Iolaphilus) julius Staudinger, see lukabas.

\*Iolaus (Iolaphilus) julus Hewitson, 1869. Fig. genitalia, Stempffer & Bennett, 1958

matilda Suffert, 1904.

- \*Iolaus (Iolaphilus) kayonza Stempffer & Bennett, 1958: 1333, figs.
- \*Iolaus (Iolaphilus) laonides Aurivillius, 1897. Fig. Aurivillius in Seitz, 1923. Iolaus (Iolaphilus) lekanion H. H. Druce, see lukabas.
- \*Iolaus (Iolaphilus) lukabas H. H. Druce, 1890. Fig. H. H. Druce, 1902; fig. genitalia, Stempffer & Bennett, 1958.

lekanion H. H. Druce, 1891; julius Staudinger, 1891.

- \*Iolaus (Iolaphilus) maritimus maritimus Stempffer & Bennett, 1958: 1308, figs. (3). Stempffer, 1961: 91, fig. (2).
- \*Iolaus (Iolaphilus) maritimus usambara Stempffer, 1961: 91, figs.

Iolaus (Iolaphilus) matilda Suffert, see julus.

- \*Iolaus (Iolaphilus) menas H. H. Druce, 1890. Fig. H. H. Druce, 1902.
- \*Iolaus (Iolaphilus) ndolae Stempffer & Bennett, 1958: 1313, figs.
- Iolaus (Iolaphilus) paneperata H. H. Druce, 1890. Fig. H. H. Druce, 1902. Fig. genitalia, Stempffer & Bennett, 1958. elisa Suffert, 1904.
- \*Iolaus (Iolaphilus) poecilaon Riley, 1928, Novit. zool. 34: 379, fig. Fig. genitalia, Stempffer & Bennett, 1958, fig.
- \*Iolaus (Iolaphilus) schultzei Aurivillius, 1905. Fig. genitalia, Stempffer & Bennett, 1958.
- \*Iolaus (Iolaphilus) trimeni Wallengren, 1875. Fig. Trimen, 1887; fig. genitalia, Stempffer & Bennett, 1958.

  anesius Hulstaert, 1924.
- \*Iolaus (Iolaphilus) vansomereni Stempffer & Bennett, 1958: 1319, figs.

## IOLAUS (PHILIOLAUS) Stempffer & Bennett

Philiolaus Stempffer & Bennett, 1958: 1336. Type-species: Iolaus parasilanus Rebel, 1914, by original designation.

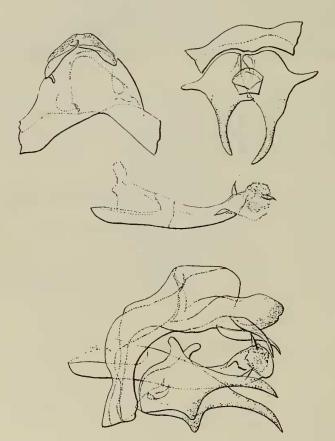
Iolaus Hübner (partim); Aurivillius, 1923: 395.

Eyes, palpi, antennae and legs as in Argiolaus and Iolaphilus.

Wing shape. Fore wing in  $\delta$  with inner margin weakly lobed; hind wing as in Argiolaus. Male secondary sexual characters as in Argiolaus and Iolaphilus.

Venation identical with the same two subgenera.

Male genitalia (Text-fig. 121, lateral view, Text-fig. 122). Of such a specialized type as to warrant the erection of a subgenus; uncus and tegumen almost completely separated from the pseudotergum, which forms above them a kind of hood, a disposition more easily seen in a lateral view, since the parts are superimposed when seen in postero-ventral aspect; uncus reduced to two small ribbons separated by a weak median depression; subunci almost straight, tapering to a sharp point; tegumen very ample; vinculum broad with a large pseudotergum; lower fultura very reduced; valves roughly lozenge-shaped with a sharp digitate apex; penis elongate, slightly curved; vesica enclosing three short spines; uncus and lower margin of valves pilose.



FIGS 121-122. Iolaus (Philiolaus) parasilanus divaricatus Riley, 121, 3 genitalia; 122, lateral view.

## LIST OF SPECIES OF Iolaus (Philiolaus)

Iolaus (Philiolaus) parasilanus parasilanus Rebel, 1914. Fig. Riley, 1928, Novit, 2001, 34.

- \*Iolaus (Philiolaus) parasilanus divaricatus Riley, 1928, I.c.: 377, figs. Fig. genitalia, Stempffer & Bennett, 1958.
- \*Iolaus (Philiolaus) parasilanus mabillei Riley, 1928, l.c.: 378, figs. Fig. genitalia, Stempffer & Bennett, 1958.
- \*Iolaus (Philiolaus) parasilanus maesseni Stempffer & Bennett, 1958, fig. and fig. genitalia.

### IOLAUS (APHNIOLAUS) H. H. Druce

Aphniolaus H. H. Druce, 1902, Proc. zool. Soc. Lond. 1902: 117. Type-species: Myrina pallene Wallengren, by original designation.

Iolaus Hübner (partim); Aurivillius, 1898; 326; 1924; 405; Murray, 1935; 81; Swanepoel, 1953; 178.

Eyes smooth; from clothed with yellowish white hairs; palpi long, ascending, parallel, second segment long, clothed below with yellow adpressed scales, third segment long, cylindrical, acuminate, clothed with black scales; antennae short, club cylindrical, not well differentiated; thorax robust, clothed below with long silky hairs; abdomen short;  $\mathcal J$  fore leg with tibia slightly shorter than femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing inner margin straight without lobe; hind wing produced at the anal angle, outer margin angular at the end of vein 3, a delicate tail at the end of vein 2 and a much longer one at the end of 1b, a lobe at the anal angle. No secondary 3 sexual characters.

Wing venation (Text-fig. 295). Fore wing with 11 veins in both sexes; hind wing, cell very short.

Male genitalia (Text-fig. 123). Uncus composed of two small triangular lobes separated by a slight depression of the margin of the tegumen; subunci short, very robust, almost straight, shaped like two blades with rounded apices; tegumen large, quadrangular, in their normal position uncus and tegumen together are hood-shaped; vinculum broad with no saccus; lower fultura strongly developed, Y-shaped, the two apices spatulate with a slightly serrated

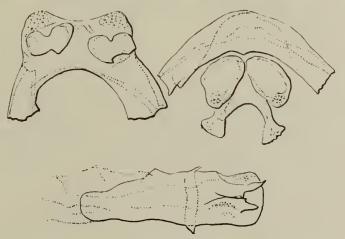


Fig. 123. Iolaus (Aphniolaus) pallene (Wallengren), & genitalia.

distal edge; valves oval, very small in comparison with the dorsal structures; penis large, very robust; vesica enclosing two large cunei; uncus, and the apices of the lower fultura and of the valvae, pilose.

Aphniolaus pallene is very easily recognized by the pale sulphur-yellow colour of its wings, a colour not found in any other species of *Iolaus*.

LIST OF SPECIES OF Iolaus (Aphniolaus)

\*Iolaus (Aphniolaus) pallene (Wallengren), 1857. Fig. Hewitson, 1878.

### IOLAUS (IOLAUS) Hübner

Iolaus Hübner, 1823, Verz. bek. Schmett.: 81. Type-species: Papilio eurisus Cramer, designated by Hewitson, 1863, Ill. diurn. Lep. Lycaenidae, 1:40. Iolaus Hübner (partim); Aurivillius, 1898: 325; 1924:403.

Eyes smooth; palpi long, slightly ascending, parallel, second segment long, slender, extending beyond the frons, third segment more slender, acuminate; antennae about half the length of the costa, slender, with a poorly differentiated club; thorax fairly robust, clothed below with silky hairs; fore leg of the 3 with tibia shorter than the femur and the tarsus unsegmented, finely spinose below, tibiae of mid and hind legs shorter than the femora, metatarsi long.

Wing shape. Fore wing in 3 with inner margin slightly lobed basally; hind wing, both sexes, outer margin angled at the end of vein 4, three delicate tails increasing in length at the ends of veins 3, 2 and 1b; a lobe at the anal angle. Male secondary sexual characters: on the underside of the fore wing a shining, scaly patch below the lower edge of the cell; in the middle of the inner margin a tuft of long, adpressed black hairs; on the upperside of the hind wing a large black scaly area below the base of vein 8 and covering the upper half of the cell.

Wing venation (Text-fig. 296). Fore wing with 11 veins in both sexes; hind wing, cell short, truncated.

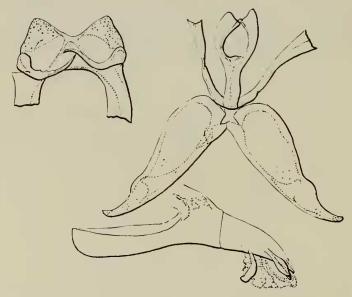


Fig. 124. Iolaus (Iolaus) eurisus (Cramer), of genitalia.

Male genitalia (Text-fig. 124). Uncus composed of two triangular lobes with rounded apices; subunci long, obtuse angled, robust at the base and tapering gradually to the apex; tegumen large; in their normal position uncus and tegumen together are hood-shaped; vinculum wide above, narrow below, no saccus; lower fultura consists of a ring, which closely encircles the penis, and is borne on a long peduncle fused to the point of origin of the valves; valves oblong, slightly falcate at the apices, the upper process bearing distally a lobe folded back on the valve; penis long, robust, internally widely open dorsally, external portion tapering gradually to the apex; vesica enclosing two large cunei, (exserted in the figure); uncus densely hairy, only a few hairs on the apices of the valves.

The genitalia of bolissus and carina only differ from those of eurisus in certain details of the shape of the valves.

# LIST OF SPECIES OF Iolaus (Iolaus)

\*Iolaus (Iolaus) bolissus bolissus Hewitson, 1873 (3). Fig. Hewitson, 1878. Fig. (2) and fig. genitalia, Stempffer & Bennett, 1958.

Iolaus (Iolaus) bolissus aurora Clench, 1964, Jl N.Y. ent. Soc. 72: 243.

Iolaus (Iolaus) bolissus azureus Clench, 1964, l.c.: 243.

\*Iolaus (Iolaus) carina carina Hewitson, 1873. Fig. Hewitson, 1878. Fig. genitalia, Stempffer & Bennett, 1958.

\*Iolaus (Iolaus) carina gabonensis Stempffer, 1951, Bull. Soc. ent. Fr. 56: 125; fig. op.c., 1952.

\*Iolaus (Iolaus) eurisus (Cramer), 1779.

helius (Fabricius), 1781; thuraui Suffert, 1904.

Iolaus (Iolaus) helius (Fabricius), see eurisus.

Iolaus (Iolaus) thuraui Suffert, see eurisus.

Iolaus (Iolaus) vexillarius Clench, 1964, l.c.: 241, fig. genitalia.

## IOLAUS (EPAMERA) H. H. Druce

Epamera H. H. Druce, 1891, Ann. Mag. nat. Hist. (6) 8:139. Type-species: Iolaus sidus Trimen, by original designation.

Iolaus Hübner (partim); Aurivillius, 1898: 325, 326; 1923: 394; 1924: 397; Murray, 1935: 76; Swanepoel, 1953: 173.

In his description H. H. Druce gave the following generic characters: "Allied to *Iolaus*, smaller. Venation the same. Fore-wing below without the thick patch of scales above the tuft of hairs on the inner margin. Head broader, antennae shorter, stouter and less distinctly clavate". However, with the exception of the venation, it is necessary to state that none of the other generic characters given is to be found in every one of the species generally placed in *Epamera*. In size the species of the *laon* group are at least as large as those of typical *Iolaus*; but it is true that most of the species of *Epamera* are small. The length and the shape of the antennae seem to vary from species to species, though only slightly. In no case do they show any striking peculiarities. As in all species of the large genus *Iolaus* the fore tarsi of the 3 are unsegmented.

Wing shape. This varies from species to species. In the of fore wing the inner margin is hardly lobed at all in sidus, mimosae, tajoraca, aphnaeoides, flavilinea, alienus, aemulus and penningtoni; the lobe is more pronounced in the other species, especially in those of the iasis and hemicyanus groups. In most of the species the fore wings are roughly of the subtriangular shape, but in the hemicyanus group the inner margin is much shorter than the costal margin so that the wing becomes almond-shaped. The hind margin of the hind wings is slightly concave in alienus, violacea and bakeri; on the other hand in the other species it is angled or even tailed at the end of vein 3, this tail being moderately well developed in the laon group. There is a delicate tail at the end of vein 2, and another longer one at vein 1b which in longicauda is very long. Male secondary sexual characters: on the underside of the fore-wings there is a glossy area, between the lower edge of the cell and the inner margin, which is small or even absent in some species, e.g. mimosae, aphnaeoides, alienus, aemulus and bellina, but large and conspicuous in sidus, iasis, cytaeis, laon, silanus, australis, creta etc. A hair tuft on the inner margin is present in all species except glaucus and those of the hemicyanus group. On the upperside of the hind wing, in all the species of the subgenus, there is a more or less well developed glossy androconial patch at the base of the wing.

Wing venation (Text-fig. 297). Fore wing with 11 veins in both sexes.

Male genitalia (Text-fig. 125, sidus). Uncus composed of two small triangular lobes narrowly fused to the margin of the tegumen on either side of the median depression; subunci long, robust, massive basally, obtuse angled, blunt ended; tegumen strap-like, its posterior margin with a rounded depression; as in all Iolaus, the tegumen in its normal position is hood-shaped; vinculum rather broad, prolonged into a squat rounded saccus; lower fultura blade-shaped, its deeply indented apex encircling the penis; valves oblong-oval, the apex slightly serrate and folded towards the lower margin; penis large, inner portion widely open dorsally, apex obliquely truncate; uncus pilose, a few hairs on the distal half of the valves along the upper margin.

The male genitalia in *Epamera* are all of the "Thecline" type, that is to say, with a hood-like tegumen and well developed subunci, but they differ amongst themselves in structural details, for example: the lobes of the uncus more or less separated from one another, subunci more or less curved, outline of the valves more or less incised,

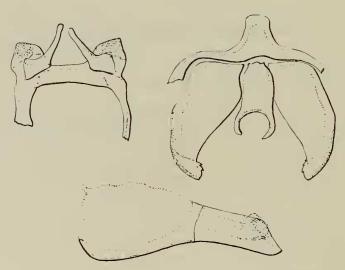


Fig. 125. Iolaus (Epamera) sidus Trimen, & genitalia.

penis more or less massive. In accordance with these characters one can form "groups" of species, but they are not homogeneous, they are ill-defined and connected by transitions, so that one cannot accord them serious systematic value. In illustration of each of these groups, reviewed below, illustration of the male genitalia is restricted to a single species, since all the species are figured in the Revision of the genus *Epamera* (Stempffer, 1959, *Bull. Inst. fr. Afr. noire* 21: 227–319).

Group A:—Tegumen with median notch; uncal lobes separated; subuncicurved; valves rather large, not much incised; penis robust and usually enclosing

large spines :—

I. (E.) sidus, silanus, scintillans, australis, mimosae (Text-fig. 126), tajoraca, jacksoni, aphnaeoides, umbrosa, nursei, laon, farquharsoni, moyambina, stenogrammica, arborifera, dubiosa, penningtoni.

Group B:—Tegumen without median notch, lobes of uncus confluent, subunci short, weakly curved, valves not much incised, penis short and massive:—

I. (E.) pollux (Text-fig. 127), coelestis, neavei, flavilinea, longicanda.

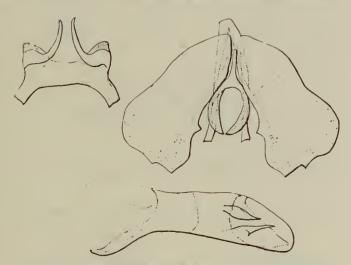


Fig. 126. Iolaus (Epamera) mimosae Trimen, & genitalia.

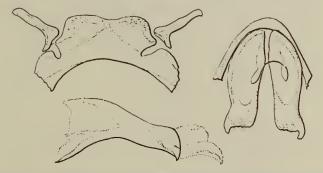


Fig. 127. Iolaus (Epamera) pollux pollux Aurivillius, of genitalia.

Group C:—Tegumen without median notch, lobes of the uncus confluent, subuncivery weakly bent, valves small, penis elongate and strongly bent (except in sapphirinus):—

I. (E.) gemmarius (Text-fig. 128), sapphirus, bellina, sciophilus, cytaeis, nolaensis,

sapphirinus, fontainei.

Group D:—Tegumen without median notch, lobes of the uncus confluent, subunci curved, valves elongate and usually deeply divided at the apex, which is at times deeply incised, penis elongate, usually sharp-pointed:—

I. (E.) frater (Text-fig. 129), hemicyanus, aethria, bansana, violacea, mermis, iasis, sibella, agnes, bakeri, pseudofrater.

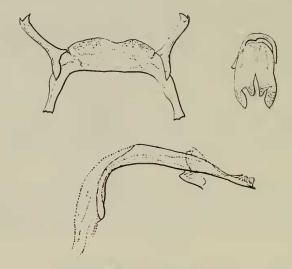


FIG. 128. Iolaus (Epamera) gemmarius Druce, & genitalia.

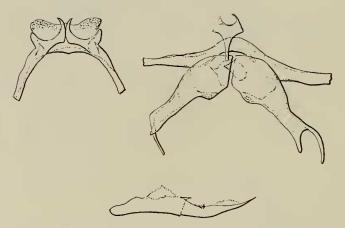


Fig. 129. Iolaus (Epamera) frater frater Joicey & Talbot, & genitalia.

Group E:—Tegumen with only a hint of a median notch, lobes of the uncus separate, subunci short, scarcely bent, valves not incised, penis very elongate, curved, ending in a tapering point:—

I. (E.) glaucus (Text-fig. 130).

Group F:—Tegumen with more or less of a median notch, uncus consisting of two ribbons fused to the lateral margins of the tegumen, subunci curved, valves elongate, penis short, vesica enclosing spines:—

I. (E.) alienus (Text-fig. 131), aemulus, obscurus.

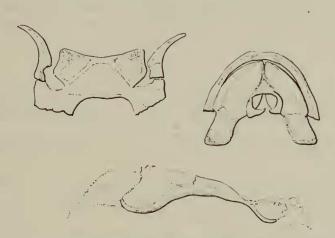


Fig. 130. Iolaus (Epamera) glaucus glaucus Butler, & genitalia.

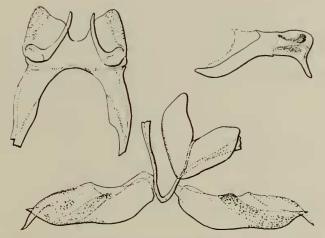


Fig. 131. Iolaus (Epamera) alienus alienus Trimen, & genitalia.

Group G:—Tegumen with a weak median notch, subunci robust, curved, valves strongly asymmetrical, penis robust:—

I. (E.) maesa (Text-fig. 132), creta.

The early stages of some of the species of *Epamera* have been described in the following papers:—

E. bansana, E. sidus, E. tajoraca, Jackson (1937, Trans. R. ent. Soc. Lond. 86: 214-6).

E. iasis, E. farquharsoni, E. laon, E. maesa, E. aethria (= mirabilis) Farquharson and Eltringham, (1922, Trans. ent. Soc. Lond. 1921: 362-3, 481-2).

E. mimosae, E. aemulus Murray (1935: 78, 80).

All the known caterpillars of *Epamera* live on leaves or flowers of species of *Loranthus* which are parasitic on various trees.

## LIST OF SPECIES OF Iolaus (Epamera)

Iolaus (Epamera) adamsi Lathy, see laon.

\*Iolaus (Epamera) aemulus Trimen, 1895. Fig. genitalia, Stempffer & Bennett, 1959.

*Iolaus (Epamera) aemulus* ♀-f. *apatosa* Talbot, 1935, *Entomologist's mon. Mag.* 71: 117, fig.

\*Iolaus (Epamera) aethria Karsch, 1893 (3). Fig. H. H. Druce, 1907 (as mirabilis). Fig. genitalia, Stempffer & Bennett, 1959.

Iolaus (Epamera) aethria Audeoud, 1936, see bellina.

\*Iolaus (Epamera) agnes Aurivillius, 1897 (3). Fig. and fig. genitalia, Stempffer & Bennett, 1959; fig. \( \begin{aligned} \partial \text{stempffer}, 1961: 98. \end{aligned} \)

Iolaus (Epamera) alberici Dufrane, 1945, see creta.

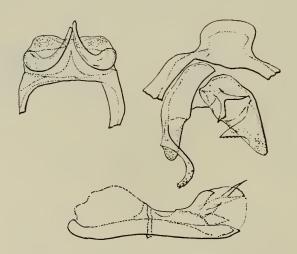


Fig. 132. Iolaus (Epamera) maesa (Hewitson), & genitalia.

\*Iolaus (Epamera) alienus alienus Trimen, 1898.

\*Iolaus (Epamera) alienus bicaudatus Aurivillius, 1905 (3). Fig. 2 and fig. genitalia, Stempffer & Bennett, 1959.

\*Iolaus (Epamera) alienae ugandae Stempffer, 1953, Annls Mus. R. Congo

Belge 29. Fig. Stempffer & Bennett, 1959.

\*Iolaus (Epamera) aphnaeoides aphnaeoides Trimen, 1873 (May). Fig. Hewitson, 1878. Fig. genitalia, Stempffer & Bennett, 1959. canissus Hewitson, 1873 (November).

\*Iolaus (Epamera) aphnaeoides diametra Karsch, 1895. Fig. Aurivillius in

Seitz, 1923. Fig. genitalia, Stempsfer & Bennett, 1959.

\*Iolaus (Epamera) aphnaeoides mafugae Stempffer & Bennett, 1959: 249.

\*Iolaus (Epamera) aphnaeoides nasissii Riley, 1928, Novit. 2001. 34: 391, fig. (2). Fig. 3, Talbot, 1935, Entomologist's mon. Mag. 71: 118.

\*Iolaus (Epamera) arborifera Butler, 1900 (♂). Fig. ♀ and genitalia, Stempffer & Bennett, 1959.

Iolaus (Epamera) aurivilli Röber, see sapphirinus.

\*Iolaus (Epamera) australis Stevenson, 1937, Occ. Papers natn. Mus. Sth. Rhod. 6:24. Fig. and fig. genitalia, Stempffer & Bennett, 1959.

\*Iolaus (Epamera) bakeri Riley, 1928, Novit. 2001. 34: 388, fig. (2). Fig. 3, Pennington, 1953, J. ent. Soc. sth. Afr. 16: 103. Fig. genitalia, Stempffer & Bennett, 1959.

\*Iolaus (Epamera) banco Stempffer, 1966, Bull. Inst. fond. Afr. noire 28: 1573, fig. Iolaus (Epamera) bansana bansana Bethune Baker, 1926, Ann. Mag. nat. Hist.

(9) 17:395.

\*Iolaus (Epamera) bansana yalae Riley, 1928, Novit. 2001. 34: 385, fig. Fig. genitalia, Stempffer & Bennett, 1959.

Iolaus (Epamera) belli Hewitson, 1869 (い); Aurivillius in Seitz, 1923 (る).

Iolaus (Epamera) belli Aurivillius, nec Hewitson, see pollux.

\*Iolaus (Epamera) bellina bellina (Plötz), 1880. Fig. Aurivillius in Seitz, 1923. Fig. genitalia, Stempffer & Bennett, 1959.

iaspis H. H. Druce, 1890; aethria Audeoud, 1936 (2).

\*Iolaus (Epamera) bellina exquisita Riley, 1928, Novit. zool. 34: 388, fig. (3). Fig. Q, Stempffer, 1961, Bull. Inst. fr. Afr. noire 23: 96.

Iolaus (Epamera) bellina maris Riley, 1928, Novit. 2001. 34: 388.

Iolaus (Epamera) bertha Suffert, see iasis.

Iolaus (Epamera) bryki Aurivillius, see hemicyanus.

Iolaus (Epamera) canissus Hewitson, see aphnacoides.

\*Iolaus (Epamera) coelestis Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 394 (♂). Fig. Stempffer, 1957, Lycaen. Afrique noire franç. Fig. ♀ and fig. genitalia, Stempffer & Bennett, 1959.

\*Iolaus (Epamera) creta (Hewitson), 1878 (3). Aurivillius, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910–11, 1: 1227 (2). Fig. genitalia, Stempffer & Bennett,

1959.

fuscomarginata Joicey & Talbot, 1921; alberici Dufrane, 1945.

\*Iolaus (Epamera) cytaeis cytaeis Hewitson, 1875. Fig. Hewitson, 1878. Fig genitalia, Stempffer & Bennett, 1959.

\*Iolaus (Epamera) cytaeis caerulea Riley, 1928, Novit. 2001. 34: 390, fig. Fig. genitalia, Stempffer & Bennett, 1959.

\*Iolaus (Epamera) cytaeis leonis Riley, 1928, Novit. zool. 34: 390, fig. (3); \$\,\text{Stempffer & Bennett, 1959: 276.}

\*Iolaus (Epamera) dubiosa Stempffer & Bennett, 1959: 310, fig. (2). Fig. 3 and genitalia, Stempffer, 1961, Bull. Inst. fr. Afr. noire 23: 99.

Iolaus (Epamera) emma Suffert, see laon.

\*Iolaus (Epamera) farquharsoni Bethune Baker, 1922, Trans. ent. Soc. Lond. 1921: 462. Fig. genitalia, Stempffer & Bennett, 1959.

\*Iolaus (Epamera) flavilinea Riley, 1928, Novit. zool. 34: 389, fig. (3). Fig. genitalia, Stempffer & Bennett, 1959,  $\mathcal{P}$ , Stempffer, 1962: 1163.

\*Iolaus (Epamera) fontainei Stempffer, 1956: 33, fig. and fig. genitalia (3); Q, Stempffer, 1962: 1163.

\*Iolaus (Epamera) frater frater Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1: 92, figs.

Iolaus (Epamera) frater kamerunica Riley, 1928, Novit. zool. 34: 385, fig.

Iolaus (Epamera) frater kumboae Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 397.

Iolaus (Epamera) fuscomarginata Joicey & Talbot, see creta.

Iolaus (Epamera) gazei H. H. Druce, see scintillans.

\*Iolaus (Epamera) gemmarius H. H. Druce, 1910 (3). Fig. genitalia, Stempffer & Bennett, 1959; Stempffer, 1961, Bull. Inst. fr. Afr. noire 23: 97, fig. (2).

\*Iolaus (Epamera) glaucus glaucus Butler, 1886.

\*Iolaus (Epamera) glaucus jordanus Staudinger, 1897.

Iolaus (Epamera) handmani Gifford, 1965, Butt. Malawi: 52, figs.

\*Iolaus (Epamera) hemicyanus hemicyanus E. Sharpe, 1904, Entomologist 37: 203. Fig. genitalia, Stempffer & Bennett, 1959.

bryki Aurivillius, 1925; barbara toroensis Riley, 1929.

Iolaus (Epamera) hemicyanus barbara Suffert, 1904. Fig. Aurivillius in Seitz, 1923.

Iolaus (Epamera) hemicyanus barnsi Joicey & Talbot, Bull. Hill Mus. Witley  $\mathbf{1}: 92$ , fig. (3); Stempffer & Bennett, 1959, fig. ( $\mathcal{P}$ ).

Iolaus (Epamera) hemicyanus mildbraedi Schultze, 1910. Fig. Aurivillius in Seitz, 1923.

yokoana Bethune Baker, 1926.

\*Iolaus (Epamera) iasis iasis Hewitson, 1865. Fig. genitalia, Stempffer & Bennett, 1959.

bertha Suffert, 1904.

\*Iolaus (Epamera) iasis albomaculatus E. Sharpe, 1904, (3); Talbot, 1937, Trans. ent. Soc. Lond.: 65, fig. (9).

Iolaus (Epamera) iaspis H. H. Druce, see bellina.

\*Iolaus (Epamera) jacksoni Stempffer, 1950, Revue fr. Ent. 17: 139, fig. Fig.

and fig. genitalia, Stempffer & Bennett, 1959.

Iolaus (Epamera) katanganus Romieux, see violacea.

\*Iolaus (Epamera) laon Hewitson, 1878 (\$\varphi\$); Aurivillius, 1898 (\$\varphi\$). Fig. genitalia, Stempffer & Bennett, 1959.

adamsi Lathy, 1903; emma Suffert, 1904.

- \*Iolaus (Epamera) longicauda Stempffer & Bennett, 1959: 264, fig. and fig. genitalia.
- \*Iolaus (Epamera) maesa (Hewitson), 1863 ( $\mathcal{P}$ ); Riley, 1928, Novit. zool. 34: 382, fig. (3). Fig. genitalia, Stempffer & Bennett, 1959.
- \*Iolaus (Epamera) mermis H. H. Druce, 1869 (3). Fig. H. H. Druce, 1902, 1907 (2); fig. genitalia, Stempffer & Bennett, 1959.

\*Iolaus (Epamera) mimosae mimosae Trimen, 1874.

\*Iolaus (Epamera) mimosae berbera Bethune Baker, 1924, Ann. Mag. nat. Hist. (9) 14: 131.

septentrionalis Stempffer, 1948.

- \*Iolaus (Epamera) mimosae haemus Talbot, 1935, Entomologist's mon. Mag. 71: 118, fig. 3; : 206, fig. Q. Fig. genitalia, Stempffer & Bennett, 1959.
- \*Iolaus (Epamera) mimosae rhodosense Stempffer & Bennett, 1959: 237.
  Iolaus (Epamera) mirabilis H. H. Druce, see aethria.
- \*Iolaus (Epamera) moyambina Stempffer & Bennett, 1959: 254, figs. and fig. genitalia.
- \*Iolaus (Epamera) neavei neavei H. H. Druce, 1910 (3). Fig. genitalia, Stempffer & Bennett, 1959. Fig. Q, Stempffer, 1961: 58.
- Iolaus (Epamera) neavei katera Talbot, 1937, Trans. ent. Soc. Lond. 86:65, fig. (♂). Stempffer, 1961, Bull. Inst. fr. Afr. noire 23:99, fig. (♀).
- \*Iolaus (Epamera) nolaensis nolaensis Stempffer, 1951, Bull. Soc. ent. Fr. 56: 125, fig.; l.c. 1952, 57: figs.
- \*Iolaus (Epamera) nolaensis amanica Stempffer, 1951, l.c.: 126; fig. 1952, l.c.
- \*Iolaus (Epamera) nursei Bulter, 1896. Fig. Aurivillius in Seitz, 1923. Fig. genitalia, Stempffer & Bennett, 1959.
- \*Iolaus (Epamera) obscurus Aurivillius, 1923. Fig. genitalia, Stempffer & Bennett, 1959.

Iolaus (Epamera) parva Bethune Baker, see pollux.

- \*Iolaus (Epamera) penningtoni Stempffer & Bennett, 1959: 312, fig. and fig. genitalia.
- \*Iolaus (Epamera) pollux pollux Aurivillius, 1895. Fig. genitalia, Stempffer & Bennett, 1959.

belli Aurivillius nec Hewitson, 1898 ; parva Bethune Baker, 1926.

- \*Iolaus (Epamera) pollux albocaerulea Riley, 1929, Trans. ent. Soc. Lond. 77: 497, fig.
- \*Iolaus (Epamera) pollux oberthueri Riley, 1929, l.c. : 496.
- \*Iolaus (Epamera) pseudofrater Stempffer, 1962: 1167, fig. and fig. genitalia.
- \*Iolaus (Epamera) pseudopollux Stempffer, 1962: 1164, fig.

\*Iolaus (Epamera) sapphirinus Aurivillius, 1897 (3). Fig. Aurivillius, 1898. Aurivillius, 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910–11, 1:558 (\$\varphi\$). Fig. genitalia Stempffer & Bennett, 1959, l.c. aurivillii Röber, 1900.

\*Iolaus (Epamera) sappirus H. H. Druce, 1902. Fig. genitalia, Stempffer &

Bennett, 1959.

\*Iolaus (Epamera) scintillans Aurivillius, 1905. Fig. genitalia, Stempffer & Bennett, 1959.

gazei H. H. Druce, 1912.

\*Iolaus (Epamera) sciophilus Schultze, 1916. Fig. Aurivillius in Seitz, 1923. Fig. genitalia, Stempffer & Bennett, 1959.

Iolaus (Epamera) septentrionalis Stempffer, see mimosae berbera.

- \*Iolaus (Epamera) sibella H. H. Druce, 1910. Fig. genitalia, Stempffer & Bennett, 1959.
- \*Iolaus (Epamera) sidus Trimen, 1864. Fig. Hewitson, 1865.

\*Iolaus (Epamera) silanus silanus Smith, 1889. Fig. Seitz, 1923. Fig. genitalia, Stempffer & Bennett, 1959.

\*Iolaus (Epamera) silanus alticola Stempffer, 1961, Bull. Inst. fr. Afr. noire 23: 94, fig. and fig. genitalia.

Iolaus (Epamera) silanus silenus Hawker Smith, 1928, Rev. Zool. Bot. afr. 16: 214.

\*Iolaus (Epamera) stenogrammica Riley, 1928, Novit. zool. 34: 384, fig. Q. Fig. 3 and genitalia, Stempffer & Bennett, 1959.

\*Iolaus (Epamera) tajoraca tajoraca Walker, 1870, (3). Fig. genitalia, Stempffer & Bennett, 1959; fig. \$\varphi\$, Stempffer, 1961, Bull. Inst. fr. Afr. noire 23: 06.

\*Iolaus (Épamera) tajoraca ertli Aurivillius, 1926. Fig. genitalia, Stempffer & Bennett, 1959.

Iolaus (Epamera) toroensis Riley, see hemicyanus.

\*Iolaus (Epamera) umbrosa umbrosa (Butler), 1885. Fig. genitalia, Stempffer & Bennett, 1959.

\*Iolaus (Epamera) umbrosa sudanica Aurivillius, 1905. Fig. genitalia, Stempffer & Bennett, 1959.

\*Iolaus (Epamera) violacea Riley, 1928, Novit. zool. 34: 386, fig. Fig. genitalia, Stempffer & Bennett, 1959.

katanganus Romieux, 1934.

Iolaus (Epamera) yokoana Bethune Baker, see hemicyanus mildbraedi.

# IOLAUS (ETESIOLAUS) Stempffer & Bennett

Etesiolaus Stempffer & Bennett, 1959, Bull. Inst. fr. Afr. noire 21: 319. Type-species: Iolaus catori Bethune Baker, 1904, by original designation.

Iolaus Hübner (partim); Aurivillius, 1923; 394.

Eyes, palpi, antennae and legs as in the preceding subgenera.

Wing shape. If fore wing inner margin lobed; hind wing hind margin angled at vein 3, a delicate tail at vein 2, another much longer at vein 1b, anal angle lobed. Male secondary sexual characters: a hair tuft on the inner margin of the fore wing below, an androconial patch surrounded by a wide glossy area, at the base of the hind wing above.

Wing venation. Fore wing with 11 veins in both sexes.

Male genitalia (Text-fig. 133). Very different from all the other *Iolaus*, justifying separation as a distinct subgenus. Uncus composed of two small lobes roughly subtriangular in shape and distally strongly serrate; tegumen very big, but with its distal margin deeply indented, centrally reduced to a narrow band; vinculum broad, with a short round saccus; lower fultura carried on a short stalk; valves small in relation to the dorsal structures, deeply incised the two processes separate almost to the base, upper process with serrate apical edge; the distal part of the lower process cut to form sharp teeth of which the longest is bent back like a sickle; penis small, almost straight, the outer part tapering evenly; uncus and upper process of valves lightly pilose.

## LIST OF SPECIES OF Iolaus (Etesiolaus)

\*Iolaus (Etesiolaus) catori catori Bethune Baker, 1904, Ann. Mag. nat. Hist (7) 14:233 (3). Riley, 1928, Novit. zool. 34:380, fig. (4).

Iolaus (Etesiolaus) catori cottoni Bethune Baker, 1908, Proc. zool. Soc. Lond.: 113, fig. (3). Fig. Riley, 1928, Novit. zool. 34:380. (4).

# IOLAUS (SUKIDION) H. H. Druce

Sukidion H. H. Druce, 1891, Ann. Mag. nat. Hist. (6) 8:142. Type-species: Iolaus inores Hewitson, 1872, by original designation.

Iolaus Hübner (partim); Aurivillius, 1898: 324; 1923: 397.

This species is very seldom represented in collections. It was described on the basis of the unique male now in the British Museum (Natural History) the provenance of which could be Gaboon or even the East Indies<sup>1</sup>. I am indebted to the late

<sup>1</sup> While the present work was already in page proof, I was informed by Lt. Col. C. F. Cowan that I. (S.) incres had recently been collected in N. E. Borneo, thus confirming that its distribution is Indo-Malayan, not African.

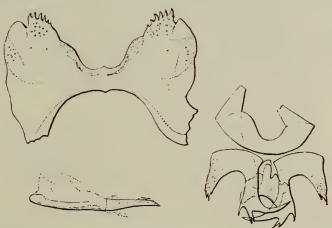


Fig. 133. Iolaus (Etesiolaus) catori catori Bethune Baker, & genitalia.

Dr. A. S. Corbet, of the British Museum, for the loan of the slide of the male genitalia of the holotype and the gift of a photograph of its venation. Other characters are derived from the descriptions of H. H. Druce and Aurivillius.

Head broad; eyes large; antennae long, with a well differentiated, elongate club.

Wing shape. Fore wing with costa very convex at its base, then almost straight, outer margin straight, slightly scalloped at the ends of the veins, inner margin straight, with a fringe of long black hairs below from near the base to the inner angle; hind wing almost circular, not produced at the anal angle, two short, fine tails at the ends of veins 2 and 1b respectively. No secondary 3 sexual characters.

Wing venation (Text-fig. 298). Fore wing with 11 veins.

Male genitalia (Text-fig. 134). Uncus composed of two lobes with rounded apices which are slightly folded over inwards, one on either side of the deep indentation of the posterior margin of the tegumen; subunci with massive bases, bent in an acute angle, the free portion tapering regularly to the apex, the lower edge bearing at the bend a short, pointed apophysis; tegumen large; vinculum narrow, with a poorly marked saccus; lower fultura with strong valves oblong, deeply notched at the apices; penis elongate, with a vesica bearing many cornuti which give it a shagreened appearance; uncus bears many long hairs, a few short hairs on the apices of the valves.

LIST OF SPECIES OF Iolaus (Sukidion)

\*Iolaus (Sukidion) inores Hewitson, 1872. Fig. Hewitson, 1878.

### IOLAUS SPECIES INCERTAE SEDIS

Iolaus bilineata Bethune Baker, 1908, Proc. zool. Soc. Lond. 1908: 113.

Described from a unique female in a private collection, which I have been unable to examine. The description does not enable one to decide to which subgenus it belongs.

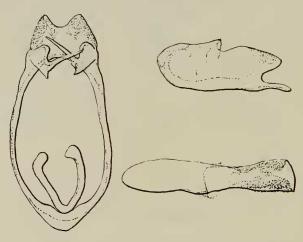


Fig. 134. Iolaus (Sukidion) inores Hewitson, & genitalia of type.

### Genus APHNAEUS Hübner

Aphnaeus Hübner, 1826, Verz. bek. Schmett.: 81; Aurivillius, 1898: 327; 1924: 407; Pinhey, 1949: 104. Type-species: Papilio orcas Drury, 1782, designated by Scudder (1875, Proc. Am. Acad. Arts Sci. 10: 116).

Eyes clothed in dense short hair; palpi fairly long, parallel, second segment long, slightly ascending, clothed with dense scales, third segment very short, slender, horizontal, with blunt apex; antennae three-fifths the length of the costa, club progressively swollen, not well differentiated;  $\eth$  fore leg with tibia as long as femur, tarsus unsegmented, femur bearing long black hairs, tibia and tarsus bearing strong spines; mid and hind legs, femora bearing long black hairs, tibiae as long as femora, metatarsi very long.

Wing shape. Fore wing subtriangular with pointed apex; hind wing much produced at the anal angle, a delicate tail at the end of vein 2, a longer tail at the end of vein 1b, an anal

lobe, the abdominal margin excised between the lobe and the end of 1a.

Wing venation (Text-fig. 299). Fore wing with 12 veins.

Aphnaeus are similar to those of orcas.

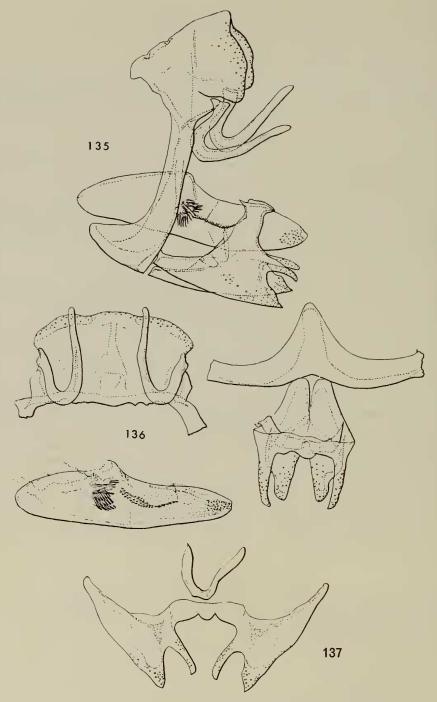
Male genitalia (Text-fig. 135: side view; Text-fig. 136: dissected, with the parts separated and spread out; Text-fig. 137: view of the valves detached from the vinculum and rotated through 180° in order to give a better view of the median band). Uncus reduced to a narrow strip bordering the edge of the tegumen; subunci long, bent in an acute angle, apices blunt; tegumen subrectangular, hood-shaped; vinculum fairly broad with a short rounded saccus; lower fultura composed of two flattened arms attached to the valves about midway, not to their base near the vinculum; valves elongate, the upper and lower processes distinctly separate in their distal half and both having rounded apices, the upper process being more slender and a little longer than the lower one; a weakly sclerotized band unites the two upper processes, passing above the external portion of the penis, which thus lies between the body of the valves and this band; penis large, with a protuberance on its upper surface situated a little before the wall of the genital cavity, the external portion tapering gradually to a blunt apex; the vesica enclosing in its base a tuft of long, strong spines, and midway and at the apex a group of cornut; uncus and distal portions of the valves pilose. The genitalia of the other species of

This peculiar arrangement of valves sheathing the penis is not confined to the genus Aphnaeus, similar structures being found in the allied genera Cigaritis (palaearctic), Apharitis, Spindasis, Chloroselas, Zeritis, Desmolycaena, Axiocerses, Phasis and Erikssonia, and in nearly all species of these genera the markings on the underside of the wings show some likeness to those of species of Aphnaeus, e.g. the presence of metallic or nacreous bands, spots or dots. The median band uniting the valves may easily escape observation because it is very little sclerotized, translucid, and so fragile that it is difficult not to break it when dissecting out the penis; when broken it is easy to mistake it for some membranous fragment.

# LIST OF SPECIES OF Aphnaeus

- \*Aphnaeus adamsi Stempffer, 1954: 513, fig. and fig. genitalia.
- \*Aphnaeus affinis affinis Riley, 1921, Trans. ent. Soc. Lond. 54: 249, figs. Fig. genitalia, Stempffer, 1954.
- \*Aphnaeus affinis seydeli Berger, 1952, Lambillionea 52:68. Fig. Stempffer, 1954.
- \*Aphnaeus argyrocyclus Holland, 1890. Fig. Holland, 1893. Fig. genitalia, Stempffer, 1954.

  propinguus Holland, 1893.



Figs 135-137. Aphnaeus orcas orcas (Drury), & genitalia.

\*Aphnaeus asterius Plötz, 1880. Fig. E. Sharpe, 1890. Fig. genitalia, Stempffer, 1954.

chalybeatus E. Sharpe, 1890; ilogo Holland, 1890.

\*Aphnaeus asterius f. argenteola Holland, 1890.

\*Aphnaeus brahami Lathy, 1903. Fig. genitalia, Stempffer, 1954. Aphnaeus bruneeli Berger, see erikssoni.

Aphnaeus chalybeatus E. Sharpe, see asterius.

Aphnaeus chapini chapini (Holland), 1920, Bull. Am. Mus. nat. Hist. 43: 225, fig.

Aphnaeus chapini occidentalis Clench, 1963, Ent. News 74: 46.

\*Aphnaeus chapini ugandae Stempffer, 1961: 59, fig.

\*Aphnaeus coronae coronae Talbot, 1935, Entomologist's mon. Mag. 71: 118, figs. ∂. Fig. genitalia and fig. Q, Stempffer, 1954.

\*Aphnaeus coronae f. vansomereni Stempffer, 1954: 512, fig.

Aphnaeus coronae littoralis Carcasson, 1964, Il E. Africa nat. Hist. Soc. 24:71,

Aphnaeus erikssoni erikssoni Trimen, 1891, Q. Fig. of genitalia, Stempffer, 1954. bruneeli Berger, 1951.

\*Aphnaeus erikssoni barnesi Stempffer, 1954: 507, fig.

\*Aphnaeus erikssoni mashunae Stempffer, 1954: 507. erikssoni (3) Trimen, 1898.

\*Aphnaeus erikssoni rex Aurivillius, 1909, \$\overline{\gamma}\$; \$\delta\$, Stempffer, 1954: 509.

\*Aphnaeus flavescens flavescens Stempffer, 1954: 514, fig. and fig. genitalia. Aphnaeus flavescens williamsi Carcasson, 1964, Il E. Africa nat. Hist. Soc. 24: 70, fig.

Aphnaeus gilloni Stempffer, 1966, Bull. Inst. fond. Afr. noire, 28: 1575, fig. Aphnaeus heliodorus Schultze, see orcas hollandi.

\*Aphnaeus jacksoni Stempffer, 1954: 512, fig. and fig. genitalia.

\*Aphnaeus jefferyi Hawker-Smith, 1928, Bull. Hill Mus. Witley 2: 30 (2); 3, and fig. genitalia, Stempffer, 1954.

Aphnaeus ilogo Holland see asterius.

\*Aphnaeus marshalli Neave, 1910. Fig. genitalia, Stempffer, 1954.

\*Aphnaeus neavei Bethune Baker, 1926. Fig. and fig. genitalia, Stempffer, 1954.

\*Aphnaeus nyanzae Stempffer, 1954: 499, fig. and fig. genitalia.

\*Aphnaeus orcas orcas (Drury), 1782. Fig. genitalia, Stempffer, 1954. \*Aphnaeus orcas f. fontainei Berger, 1952, Lambillionea 52: 70.

\*Aphnaeus orcas f. overlaeti Berger, 1952, l.c.: 70.

\*Aphnaeus orcas f. paupera Stempffer, 1954: 498, fig.

\*Aphnaeus orcas hollandi Butler, 1902.

rattrayi E. Sharpe, 1904; heliodorus Schultze, 1916.

Aphnaeus propinquus Holland, see argyrocyclus.

\*Aphnaeus questiauxi Aurivillius, 1903 (3). Fig. genitalia, Stempffer, 1954; ♀, Neave, 1910.

Aphnaeus rattrayi E. Sharpe, see orcas hollandi.

## Genus PARAPHNAEUS Thierry-Mieg

Paraphnaeus Thierry-Mieg, 1904, Naturaliste: 140. Type-species: Aphnaeus hutchinsoni Trimen, by monotypy.

Aphnaeus Hübner (partim); Aurivillius, 1898: 328; 1924: 409; Murray, 1935: 82; Swanepoel, 1953: 163.

Eyes, palpi and antennae similar to those of Aphnaeus; thorax robust, clothed below with long silky hairs;  $\beta$  fore leg, femur clothed with long fulvous hairs, tibia shorter than the femur, tarsus short, unsegmented, bearing many spines below; mid and hind legs with tibiae shorter than femora, metatarsi long.

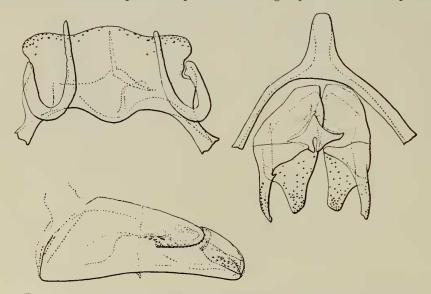
Wing shape. Similar to that of Aphnaeus except that the outer margin of the fore wing is a little more convex between the apex and the ending of vein 4.

Wing venation (Text-fig. 300). Same as that of Aphnaeus.

Male genitalia (Text-fig. 138). Uncus, subunci and tegumen as in Aphnaeus, except that the posterior edge of the dorsum has a rounded median prominence; vinculum fairly wide; lower fultura consists of a small shield-shaped plate with a deep notch in the upper edge; valves like those of Aphnaeus, the upper and lower processes of each valve separated in their distal halves, the middle portions of the upper processes folded over the inner face and connected by a membranous strip which passes above the penis; penis very bulky, internal portion swollen, the external portion bearing on the dorsal surface, at the base, a sort of lobe bearing short regular teeth on its inferior edge, the apex of penis split into two longitudinally; vesica with cornuti; uncus and distal half of valves clothed with fine hairs.

The early stages of *P. hutchinsoni drucei* Neave have been described by Jackson (1937, *Trans. R. ent. Soc. Lond.* **86**: 216). The caterpillar lives on *Acacia stenocarpa* Hochst., *Entada abyssinica* Stend., and on certain species of *Loranthus* in symbiosis with ants.

The genus *Paraphnaeus*, which has only one species, is very closely allied to the genus *Aphnaeus*, except that the lower fultura reminds one of that of *Apharitis* and *Spindasis*, and that the shape of the penis differs slightly from that of *Aphnaeus*.



Figs. 138. Paraphnaeus hutchinsoni hutchinsoni (Trimen), 3 genitalia.

## LIST OF SPECIES OF Paraphnaeus

- \*Paraphnaeus hutchinsoni hutchinsoni (Trimen), 1887. Fig. Butler, 1898. zanzibarensis (Smith), 1889.
- \*Paraphnaeus hutchinsoni drucei (Neave), 1904.
  Paraphnaeus zanzibarensis (Smith), see hutchinsoni.

## Genus APHARITIS Riley

Apharitis Riley, 1925, Novit. 2001. 32: 78. Type-species: Polyommatus epargyros Eversman (a palaearctic species), by original designation.

Spindasis Wallengren (partim); Aurivillius, 1898: 332; 1924, 414.

Eyes glabrons; palpi fairly long, second segment long, laterally compressed, clothed below with dense white adpressed scales, third segment rather short, slightly inflected, acuminate; antennae slightly longer than half the length of the costa, white-annulated, club elongate, flattened, rust-coloured below; thorax robust, clothed below with long silky hairs; of fore leg with tibia as long as femur, tarsus unsegmented, finely spinose below; mid and hind legs with tibiae shorter than femora, metatarsi very long.

Wing shape. Fore wing with apex pointed, outer margin convex between the apex and vein 4, slightly concave between vein 4 and the inner angle; hind wing with costa much arched, apex rounded, outer margin slightly concave between veins 4 and 2, angular at the end of vein 2, a delicate tail at the end of 1b, a small lobe at the anal angle.

Wing venation (Text-fig. 301). Fore wing with 11 veins.

Male genitalia (Text-fig. 139; for lateral view see Riley, 1925, fig. 5). Uncus composed of two subtriangular lobes with rounded apices and separated by a deep groove in the hind margin of the tegumen; subunci long and robust, strongly bent at about two-fifths of their length and tapering gradually to a blunt apex; tegumen rather small, in situ uncus and tegumen together are hood-shaped; vinculum fairly wide, with a short, broad saccus; lower fultura consists of a small shield-shaped plate which is deeply indented on its upper edge; valves subtriangular with blunt apices, their upper processes folded inwards and connected midway by a narrow membranous band, which passes over the penis in the same way as in species of Aphnaeus, Paraphnaeus, Spindasis, etc.; penis robust, with a cylindrical basal portion, the dorsal surface distinctly at an angle to the general axis, as in species of Spindasis (Riley, 1925, calls it "funnel-shaped"); the ventral surface of the external portion is gutter-

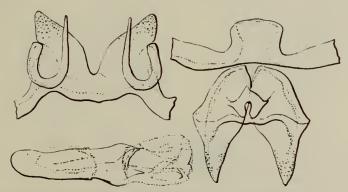


Fig. 139. Apharitis epargyros (Eversman), & genitalia.

shaped and longer than the dorsal surface; vesica unarmed; uncus and apices of valves finely pilose. (*Note*. In the mount from which this figure was drawn the penis has unfortunately undergone a partial rotation, so that the characters given in the description are not clearly shown in the figure.)

The genitalia of the Ethiopian species of Apharitis examined are very similar to those of A. epargyros.

The early stages of the *Apharitis* from tropical Africa are unknown. Those of *A. myrmecophila* Dumont, from Tunisia, and of *A. acamas chitralensis* Riley, from Chitral, have been observed. The caterpillar of *A. myrmecophila* is nocturnal in its habits, and lives and pupates at the foot of tufts of *Calligonum comosum* L'Herit (Polygonaceae), in the large tunnels made by ants.

## LIST OF ETHIOPIAN SPECIES OF Apharitis

\*Apharitis acamas bellatrix (Butler), 1886. Fig. Klug, 1834.

Apharitis buchanani (Rothschild), 1921, Novit. zool. 28: 155 ( $\mathcal{P}$ ). Talbot, 1942, Entomologist 75: 249 ( $\mathcal{F}$ ).

Apharitis gilletti Riley, 1925, Novit. zool. 32:85, fig. genitalia.

\*Apharitis nilus (Hewitson), 1865. Fig. Aurivillius in Seitz, 1924. Fig. genitalia, Riley, 1925, l.c.

subaureus (Smith), 1898.

Apharitis nilus f. kaduglii (Bethune Baker), 1916.

Apharitis nilus f. sabulosa (Hawker Smith), 1929, Bull. Hill Mus. Witley 3: 231.

Apharitis subaureus (Smith), see nilus.

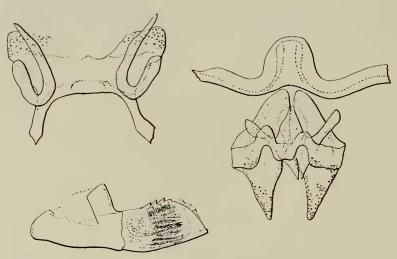


Fig. 140. Spindasis natalensis (Doubleday & Hewitson), & genitalia.

### Genus SPINDASIS Wallengren

Spindasis Wallengren, 1857, K. svenska VetenskAkad. Handl., (n.f.) 2 (1) 4:45; Murray, 1935:83; Pinhey, 1949:104; Swanepoel, 1953:165. Type-species: Spindasis masilikazi Wallengren, 1857 (Aphnaeus natalensis Westwood, 1851), by monotypy. Spindasis Wallengren (partim); Aurivillius, 1898:328; 1923:410.

Eyes glabrous; palpi fairly long, slightly divergent, second segment long, ascending and clothed with dense scales, third segment horizontal, short; antennae slightly longer than half the length of the costa, club fusiform, not well differentiated; thorax robust, clothed with long silky hair; abdomen black and white-annulated; 3 fore leg, femur clothed with long, white, silky hair, tibia slightly shorter than the femur and armed below with long spines, tarsus unsegmented and also armed below with numerous spines; mid and hind legs strong, femora clothed with long white hair, tibiae shorter than the femora and bearing apical spurs, tarsi long, segmented and bearing numerous spines.

Wing shape. Fore wing triangular, the apex pointed; hind wing produced, a delicate tail at the end of vein 2 and a longer one at the end of 1b, a small lobe at the anal angle.

Wing venation (Text-fig. 502). Fore wing with 11 veins.

Male genitalia (Text-fig. 140). Uncus composed of two lobes with straight posterior edges and rounded apices, widely separated by a shallow rounded depression with a small median prominence; subunci long and robust, bent at an acute angle at about two-fifths of their length; tegumen large and trapezoidal, tegumen and uncus together in situ are hood-shaped; vinculum rather broad with a rounded saccus; lower fultura shield-shaped, with a deep notch in its upper edge; valves elongate, subtriangular, with an almost straight inferior edge and blunt apices, the two upper processes connected by a weakly sclerotized median band, which passes above the penis as in species of Aphnaeus and allied genera; penis short, massive, with cylindrical basal portion widely open dorsally, the dorsal surface then becoming bent at an acute angle to the general axis of the penis, the large prominence thus formed being held, in the natural position, in a notch of the upper edge of the median band, which unites the valves, an arrangement which must circumscribe considerably the possible movement of the penis during copulation; the external portion of penis swollen, with an obliquely cut apex, its dorsal edge bristling with short spines; vesica with numerous fine cornuti; the lobes of the uncus, apices of the valves and lower edge of the median band, all pilose.

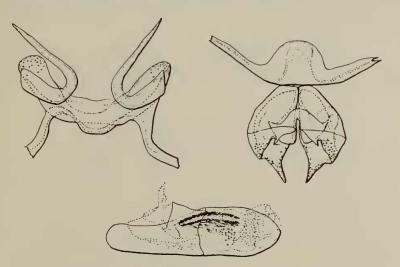


Fig. 141. Spindasis phanes (Trimen), & genitalia.

The genitalia of *Spindasis nyassae*, victoriae, mozambica, apelles, tavetensis and banyoana are similar to those of natalensis. Those of S. phanes (Text-fig. 141) are also of the same type, except that the lobes of the uncus are more lanceolate, the tegumen is narrower in the middle and its anterior margin forms a rounded prominence. The genitalia of S. homeyeri, trimeni, namaqua, somalina, ella, iza and cynica are more or less close to those of phanes. The armature of S. waggae (Text-fig. 142) is different from the preceding species, the tegumen being more ample and quadrangular, and the subunci less robust.

The larvae of S. natalensis, mozambica and ella have been described by Murray (1935: 86, 88, 89) and Pinhey (1949: 105). They live on Vigna angustifolia and Mundulea subrosa. Those of S. nyassae have been found on the bark of Acacia stenocarpa and Entada abyssinica, look like larvae of Lymantriidae, and live in company with ants; in captivity they soon die if separated from the ants (see Jackson, 1937, Trans. R. ent. Soc. Lond. 1937: 217).

## LIST OF SPECIES OF Spindasis

\*Spindasis apelles apelles (Oberthur), 1878.

Spindasis apelles nairobiensis E. Sharpe, 1904.

Spindasis apuleia Hulstaert, 1924, Revue zool. afr. 12: 127.

Spindasis avriko (Karsch), 1893.

Spindasis baghirmi Stempffer, 1946, Revue fr. Ent. 13: 14, fig.

\*Spindasis banyoana Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 398.

Spindasis brunnea Jackson, 1965, Ann. Mag. nat. Hist. (13) 8: 529, fig.

Spindasis caffer (Trimen), see mozambica.

Spindasis chaka (Wallengren), see ella.

Spindasis crustaria crustaria (Holland), 1890. Fig. Holland, 1893.

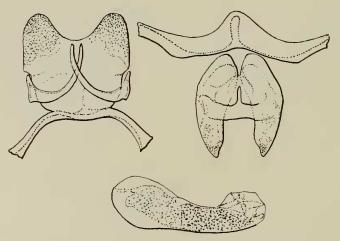


Fig. 142. Spindasis waggae Sharpe, of genitalia.

- Spindasis crustaria mysteriosa Clench, 1965, Butt. Liberia: 360, figs.
- \*Spindasis cynica Riley, 1921, Trans. ent. Soc. Lond. 54: 247, figs.
- \*Spindasis ella (Hewitson), 1865. chaka (Wallengren), 1875.
- \*Spindasis ella f. barnesi Stempffer, 1953, Annls Mus. R. Congo belge 27:30. Spindasis erna (Staudinger), see phanes.
- \*Spindasis homeyeri homeyeri (Dewitz), 1886.
- \*Spindasis homeyeri f. fracta Stempffer, 1948, Revue fr. Ent. 15: 189.

Spindasis homeyeri kallimon H. H. Druce, 1905.

- \*Spindasis iza (Hewitson), 1865.
- Spindasis lutosa (Plötz), 1880.
- Spindasis masilikazi Wallengren, see natalensis.
- Spindasis menelas H. H. Druce, 1907.
- Spindasis modestus (Trimen), 1891.
- Spindasis montana Joicey & Talbot, 1924, Bull. Hill Mus. Witley 1: 545.
- \*Spindasis mozambica (Bertoloni), 1851. Fig. Hewitson, 1865, (as natalensis). caffer (Trimen), 1868.
- \*Spindasis namaqua (Trimen), 1874.
- \*Spindasis natalensis (Westwood), 1851.

masilikazi Wallengren, 1857.

- Spindasis natalensis f. obscura Aurivillius, 1923.
- \*Spindasis nyassae (Butler), 1884. Fig. Butler, 1894.
- \*Spindasis phanes (Trimen), 1873.

erna (Staudinger), 1888.

- Spindasis scotti Gabriel, 1954, Exped. S.W. Arabia 1937/38: 378, fig.
- \*Spindasis somalina Butler, 1886.
- \*Spindasis tavetensis Lathy, 1906.
- \*Spindasis trimeni trimeni Neave, 1910.
- Spindasis trimeni congolanus Dufrane, 1954, Bull. Annls Soc. R. ent. Belg. 90: 283.
- \*Spindasis victoriae (Butler), 1884. Fig. Aurivillius in Seitz, 1923.
- \*Spindasis waggae E. Sharpe, 1898. Fig. Aurivillius in Seitz, 1923.

### Genus LIPAPHNAEUS Aurivillius

Lipaphnaeus Aurivillius, 1916, Ark. Zool. 10 (14): 2. Type-species: Aphnaeus spindasoides Aurivillius, 1916 (a subspecies of Aphnaeus aderna Plötz, 1880), by original designation. Spindasis Aurivillius (partim); Aurivillius, 1898: 332; 1923: 415.

Eyes, palpi, antennae and legs as in Spindasis.

Wing shape like that of Spindasis, except that in aderna and loxura the tail at the end of vein 1b on the hind wing is wider than that of Spindasis, which is thread-like.

Venation is not constant in Lipaphnaeus. Vein 8 of the fore wing is very short and often absent. When Aurivillius described spindasoides in 1916 he attributed to it 12 veins in the fore wing and placed it in the genus Aphnaeus, then, at the end of the text, he designated it as type of a "subgenus Lipaphnaeus", intermediate between Aphnaeus and Spindasis. In 1923

(in Seitz, Grossschmett. Erde: 413) he went back on his original opinion in these words: "By a regrettable error of observation, I was induced to describe this form as an Aphnaeus; on a closer examination I have found that the fore wing has only 11 veins, vein 8 being absent and vein 7 terminating at the apex of the wing as in all the species of Spindasis. The form is in fact so closely allied to S. aderna that I can now only consider it to be the eastern race of it". This statement, however, only confuses the issue because when I was able to examine closely the holotype of spindasoides, preserved in the Natural History Museum at Stockholm, I found that in fact it had 12 veins in the fore wing.

From an examination of a fairly large number of specimens of *Lipaphnaeus*, the following facts emerge:

- L. aderna aderna: 1 ♀ Ghana, 1 ♀ Katanga: 11 veins.
- L. aderna spindasoides: 3 & S. Rhodesia, 12 veins; 1 \( \rightarrow \) S. Rhodesia, 1 \( \rightarrow \) Mozambique: 11 veins.
- L. aderna pan: 3 ♂, 2 ♀ Uganda: 11 veins.
- L. leonina bitje: 2 ♂, 1 ♀ Middle Congo, 1 ♀ Katanga: 11 veins.
- L. loxura: 2 & Katanga, 1 & Uganda: 12 veins; 3 ♀ Uganda, 11 veins.
- L. eustorgia: 1 ? Katanga: 11 veins.

I give these figures with some reserve because, when present, vein 8 is very difficult to detect, even when the wing is soaked in alcohol. To be absolutely certain, it is necessary to remove the scales completely by immersing the wing in Eau-de-Javelle, which results in permanent damage to the specimen. However, I do not think I made a mistake in every case, and can only conclude that Lipaphnaeus sometimes has 12, sometimes 11 veins in the fore wing. One interesting fact emerges from the figures, which naturally, are too small to be the basis of safe conclusions: whilst the males are almost equally divided (6:5) between 12 and 11 veins, none of the females has more than 11. Lipaphnaeus is much better characterized by its 3 genitalia, in which the dorsal structures sharply differ from Spindasis.

Male genitalia (aderna aderna, Text-fig. 143). Uncus subtriangular, deeply divided to form two tapering points; subunci long, very slender, evenly curved and folded (when in situ) inwardly below the uncus; tegumen very small, well differentiated from the vinculum; vinculum narrow dorsally, wider ventrally, with a rounded saccus; inferior fultura lozenge-shaped, deeply divided distally; valves oblong, tapering to a point, the upper processes united midway by a narrow band as in Spindasis; penis elongate, much less massive than in Spindasis, devoid of dorsal protuberance but with a rounded depression midway on dorsal margin, the outer portion tapering to a blunt end; margins of uncus and apex of valves pilose.

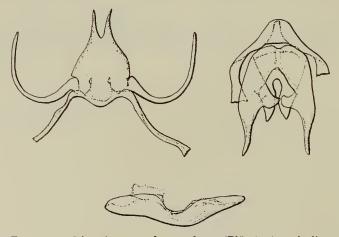


Fig. 143. Lipaphnaeus aderna aderna (Plötz), & genitalia.

The male genitalia of aderna spindasoides are identical to those of aderna aderna. Those of loxura and leonina (Text-fig. 144) are slightly different, but of the same type. I have not been able to examine eustorgia, of which only the female holotype is known.

The armatures of the three species of *Lipaphnaeus* examined are remote from those of *Spindasis*, but, on the other hand very close to those of *Chloroselas* and *Desmolycaena*. Their general appearance is similarly different from *Spindasis*. I conclude that *Lipaphnaeus* is a distinct, valid genus, not a subgenus.

## LIST OF SPECIES OF Lipaphnaeus

\*Lipaphnaeus aderna aderna (Plötz), 1880. Fig. Crowley, 1890 (as fallax and latifimbriata).

fallax (E. Sharpe), 1890; latifimbriata (E. Sharpe), 1890.

\*Lipaphnaeus aderna pan (Talbot), 1935, Entomologist's mon. Mag. 71: 120, fig.

\*Lipaphnaeus aderna spindasoides (Aurivillius), 1916.

Lipaphnaeus bicolor (E. Sharpe), see leonina.

Lipaphnaeus eustorgia (Hulstaert), 1924, Revue zool. afr. 12: 128.

Lipaphnaeus fallax (E. Sharpe), see aderna.

Lipaphnaeus latifimbriata (E. Sharpe), see aderna.

Lipaphnaeus leonina leonina (E. Sharpe), 1890. Fig. Crowley, 1890. bicolor (E. Sharpe), 1891.

\*Lipaphnaeus leonina bitje (H. H. Druce), 1910.

Lipaphnaeus leonina ivoirensis Stempffer, 1966, Bull. Inst. fond. Afr. noire 28: 1577, fig.

Lipaphnaeus leonina paradoxa (Schultze), 1908.

\*Lipaphnaeus loxura (Rebel), 1914.

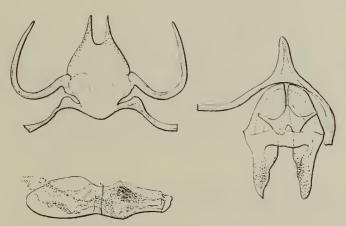


Fig. 144. Lipaphnaeus leonina leonina (Sharpe), & genitalia.

#### Genus CHLOROSELAS Butler

Chloroselas Butler, 1886, Proc. zool. Soc. Lond. 1885: 765; Aurivillius, 1898: 323; 1923: 416; Murray, 1935: 90; Swanepoel, 1953: 168. Type-species: Chloroselas esmeralda Butler, 1886, by original designation.

Eyes glabrous; palpi long, reaching beyond the frons, second segment ascending, wider and more curved than in Spindasis, clothed below with large erect scales, third segment horizontal, shorter than in Spindasis, clothed with adpressed scales; antennae more slender than those of Spindasis, with a better differentiated, fusiform club; thorax clothed below with white hair; abdomen white-annulated;  $\eth$  fore leg more slender than in Spindasis, tibia shorter than the femur, tarsus long, unsegmented, spinose below.

Wing shape. Hind wing produced at the anal angle, a delicate tail at the end of 1b (in pseudozeritis, overlaeti, azurea and taposana there is also a short delicate tail at the extremity of vein 2), an indistinct lobe at the anal angle.

Wing venation (Text-fig. 303). Fore wing with 10 or 11 veins; vein 11 is sometimes absent entirely; when present, it is reduced to a short stalk joining the upper margin of the cell to the costal vein; veins 6 and 7 are also sometimes slightly stalked. Individual variations of this kind are frequent.

Male genitalia (Text-fig. 145). Dorsally similar to Lipaphnaeus, except that the apex of the uncus is less deeply divided and the lateral margins are serrate; subunci long, evenly curved and tapering; tegumen reduced, intimately fused to the uncus but not with the vinculum; vinculum narrow with a subtriangular saccus; lower fultura shield-shaped, with deeply notched apex; valves oblong with blunt apices, their basal thirds fused together, the upper processes connected midway by a membranous band which surrounds the penis as in the preceding genera; internal portion of penis ovoid, the external portion cylindrical with an obliquely truncate apex; vesica with large cornuti; uncus and distal extremity of valves pilose.

The male genitalia of *pseudozeritis*, overlaeti, argentea and azurea closely resemble those of *C. esmeralda*, and in the first three the vesica encloses a sheaf of long, fine spines; in azurea the external portion of the penis is slightly shorter and the vesica and devoid of cornuti.

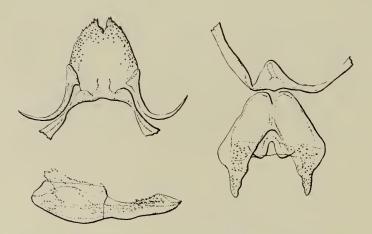


Fig. 145. Chloroselas tamaniba esmeralda Butler, o genitalia.

The caterpillar of *C. pseudozeritis tytleri* Riley has been described by Jackson (1937, *Trans. R. ent. Soc. Lond.* 86: 218). It lives on *Acacia stenocarpa* Hochst. (ex. A. Rich) (Leguminosae), in symbiosis with ants of the genus *Crematogaster*.

### LIST OF SPECIES OF Chloroselas

- \*Chloroselas argentea Riley, 1932, Ann. Mag. nat. Hist. (10) 10: 145, fig.
- \*Chloroselas azurea Butler, 1899.

Chloroselas minima Jackson, 1965, Ann. Mag. nat. Hist. (13) 8:524, fig.

Chloroselas ogadenensis Jackson, 1965, l.c.: 526, fig.

\*Chloroselas overlaeti Stempffer, 1956: 36, fig.

\*Chloroselas pseudozeritis pseudozeritis (Trimen), 1873.

\*Chloroselas pseudozeritis tytleri Riley, 1932, Ann. Mag. nat. Hist. (10) 10: 147, fig.

Chloroselas pseudozeritis tytleri f. umbrosa Talbot, 1935, Entomologist's mon. Mag. 71: 207, fig.

Chloroselas tamaniba tamaniba (Walker), 1870.

\*Chloroselas tamaniba esmeralda Butler, 1886.

Chloroselas taposana Riley, 1932, Ann. Mag. nat. Hist. (10) 10: 146, fig.

Chloroselas vansomereni Jackson, 1965, l.c.: 525, fig.

### Genus ZERITIS Boisduval

Zeritis Boisduval, 1836, Spec. Gen. Lep. 1, pl. 22, fig. 6; Aurivillius (as a "gen. nov."), 1898: 333; 1924: 417. Type-species: Zeritis neriene Boisduval, by monotypy. Aurivillius, believing Boisduval's Zeritis invalid, re-introduced the name as new.

Eyes glabrous; palpi rather short but extending beyond the frons, second segment clothed with long white scales, third segment cylindrical with rounded apex; antennae about half the length of the costa, club elongated, not well differentiated;  $\eth$  fore leg with tibia shorter than the femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing apex angular, outer margin very convex; hind wing with two short, delicate tails, at the ends of vein 2 and 1b, a small lobe at the anal angle.

Wing venation (Text-fig. 304). Fore wing with only 10 veins.

Male genitalia (Text-fig. 146). Uncus composed of two subtriangular rounded lobes; subunci long, curved, with robust bases, tapering gradually to the apices with, on the inner margin of the basal half, a long apophysis; tegumen with convex anterior edge; in situ uncus and tegumen together are hood-shaped; vinculum rather narrow, prolonged to form a short triangular saccus; lower fultura fused to the base of the valves, asymmetrical, one branch having a spatulate apex bearing short spines, the other distinctly longer branch having a slightly serrate edge and tapering gradually to the pointed apex; valves oblong, their upper processes connected by a membrane as in the preceding genera; penis long, robust, weakly curved, with an obliquely truncated apex; vesica containing numerous small cornuti; uncus and distal portions of the valves pilose.

The male genitalia of Z. pulcherima resembles so closely those of Z. neriene that one wonders whether these are two species or just two forms of one species, differing only by the markings on the underside of the wings. The male genitalia of Z. sorhageni and fontainei, whilst being of the same type as those of Z. neriene, differ

from them in the following particulars:—in Z. sorhageni the lobes of the uncus are quadrangular, the subunci bear two small apophyses instead of one large one, and the two branches of the lower fultura are much stouter, though still asymmetrical. In fontainei the lobes of the uncus are rounded, there is only one small apophysis on the lower margin of the subunci, the protuberance on the margin of the tegumen is rounded and very large, and the branches of the inferior fultura are as in sorhageni. In the shape of the uncus, tegumen and subunci, the species of Zeritis are more closely allied to species of Axiocerses than to those of Chloroselas and Desmolycaena.

### LIST OF SPECIES OF Zeritis

Zeritis aurivillii Schultze, 1908. Fig. Aurivillius in Seitz, 1924.

\*Zeritis fontainei Stempffer, 1956: 35, fig.

\*Zeritis neriene neriene Boisduval, 1836.

Zeritis neriene f. muzizii Bethune Baker, 1924, Ann. Mag. nat. Hist. (9) 14: 134.

Zeritis neriene amine (Butler), 1874.

\*Zeritis pulcherrima Aurivillius, 1916. Fig. Aurivillius in Seitz, 1924.

\*Zeritis sorhageni (Dewitz), 1879.

### Genus DESMOLYCAENA Trimen

Desmolycaena Trimen, 1898, Trans. ent. Soc. Lond. 1898: 7; Aurivillius, 1898: 334; 1924: 419; Swanepoel, 1953: 169. Type-species: Desmolycaena mazoensis Trimen, by original designation.

Eyes glabrous; palpi long, ascending beyond the frons, second segment long, flattened laterally, clothed below with long, erect scales, third segment very short, slender, cylindrical, with obtuse apex, clothed with adpressed scales; antennae three-fifths of length of costa, rather slender with a well differentiated, fusiform club; thorax clothed below with long white

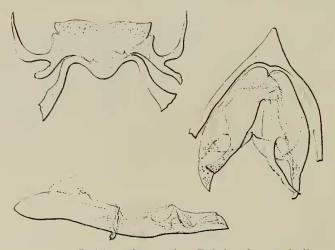


Fig. 146. Zeritis neriene neriene Boisduval, & genitalia.

hair; legs short, rather robust; 3 fore leg with femur clothed with white hairs, tibia shorter than femur, tarsus rather long, unsegmented, bearing long spines below.

Wing shape. Fore wing with apex angular; hind wing with a small filiform tail at the end of vein 1b.

Wing venation (Text-fig. 305). Fore wing with 11 veins; 11 free, but very close to the costal vein and sometimes in contact with it. As in *Chloroselas*, variations in the venation are not rare. Riley cited examples in his original description of rogersi.

Male genitalia (Text-fig. 147). The & genitalia of D. mazoensis are very similar to those of Chloroselas esmeralda, except that the lateral margins of the uncus are not serrate and the apical depression is rounded instead of triangular; the subunci are a little shorter and the vesica has no cornuti.

The male genitalia of *D. arabica* are almost identical with those of *D. mazoensis*. On the other hand, those of *D. rogersi* (Text-fig. 148) differ considerably; the dorsum is rectangular instead of oval and the subunci are reduced to two rudiments; the inferior fultura, valves and penis are as in *D. mazoensis*. By the structure of the genitalia as well as by the markings of the wings, the species of the genus *Desmolycaena* are closely allied to those of *Chloroselas*.

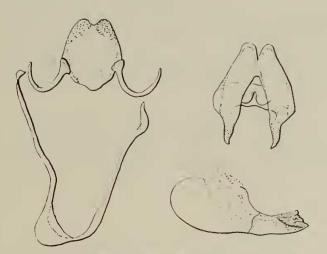


Fig. 147. Desmolycaena mazoensis Trimen, of genitalia.

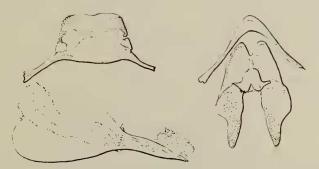


Fig. 148. Desmolycaena rogersi Riley, & genitalia.

## LIST OF SPECIES OF Desmolycaena

- \*Desmolycaena arabica Riley, 1932, Ann. Mag. nat. Hist. (10) 10: 148, fig.
- \*Desmolycaena mazoensis Trimen, 1898.
- \*Desmolycaena rogersi Riley, 1932 Ann. Mag. nat. Hist. (10) 10: 149, fig.

### Genus AXIOCERSES Hübner

Axiocerses Hübner, 1826, Verz. bek. Schmett.: 71; Aurivillius, 1898: 334; 1924: 419; Murray, 1935: 91; Pinhey, 1949: 105; Swanepoel, 1953; 162. Type-species: Papilio perion Stoll, 1781 (i.e. Papilio harpax Fabricius, 1775), by monotypy. Chrysorichia Wallengren, 1857, Rhop. Caffr.: 44.

In 1857 Wallengren (*Rhop. Caffr.*: 44) erected the genus *Chrysorichia* for what he regarded as two species, which he called *thyra* Linn and *tjoane* sp. nov. These, according to Aurivillius, who had access to the type specimens, were in fact the male and female respectively of the species now known as *Axiocerses harpax* Fabricius. However, in 1875 Scudder selected "thyra Linn" as the type-species. According to the present International Rules of Zoological Nomenclature, Article 70, a case such as this, in which the type-species of a genus has obviously been misidentified, is to be submitted to the Commission for a ruling. I propose therefore to invite the Commission to designate *Papilio harpax* Fabricius as the type-species of *Chrysorichia*, which will make that name an objective synonym of *Axiocerses*, and in the meantime to regard *Chrysorichia* as such.

Eyes glabrous; frons clothed with erect hairs; palpi rather short, parallel, hardly protruding beyond the frons, second segment clothed with erect scales and hair, third segment short, acuminate, slightly ascending; antennae more than half the length of the costa, white-annulated, club cylindrical, well differentiated; thorax robust, clothed above with long silky hair and below with close woolly hair;  $\eth$  fore leg very robust, tibia as long as femur, both densely

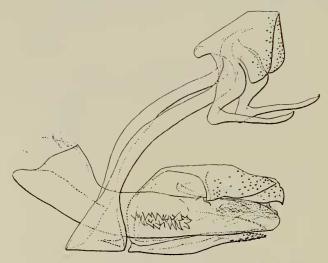


Fig. 149. Axiocerses harpax harpax (Fabricius), of genitalia.

clothed with hair, tibia with a dorsal outer spur and two inner apical spines, tarsus stout, unsegmented, clothed with short adpressed hair and bearing below long spines, the apical claw slightly curved; mid and hind legs very robust, with tibiae shorter than femora, tarsi long and stout, strongly spinose below, the metatarsus very long.

Wing shape. Fore wing with apex pointed, outer margin angled at the end of vein 4; hind wing oval, produced at the anal angle, outer margin slightly scalloped at the ends of the veins,

a delicate tail at the end of vein 1b, a small, well marked lobe at the anal angle.

Wing venation (Text-fig. 306). Fore wing with only 10 veins.

Male genitalia (Text-fig. 149, side view of genitalia, right valve removed and other parts in situ; Text-fig. 150, postero-ventral view of genitalia, parts separated and flattened out); uncus composed of two oval lobes; subunci long, much swollen basally, bent in an acute angle, the free branch slender and ending in a widely open claw, the lower edge bearing a short apophysis at the angle; tegumen lozenge-shaped; in situ uncus and tegumen together are hood-shaped; vinculum fairly wide, prolonged to form a short, robust saccus; lower fultura composed of two long conical processes bristling with strong spines at the tip; valves oblong, their upper processes rolled back on to the inner surface and connected in the middle by a thinly sclerotized band which passes above the penis; penis long and robust, slightly curved, its internal portion swollen; vesica (exserted in Text-fig. 150) encloses a number of large cornuti; uncus and upper processes of the valves pilose.

In *bambana* the inferior fultura is formed of two suboval lobes with rounded ends, a character which permits its easy separation from *harpax*, a species with which it is easily confused if reliance is placed only on the very variable external appearance (cf. Stempffer, 1957, *Bull. Inst. fr. Afr. noire* 19: 217).

On the other hand, in *amanga* the two branches of the inferior fultura are very long and tapering and extend beyond the tips of the valves.

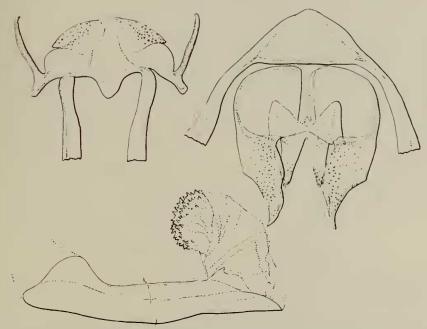


FIG. 150. Axiocerses harpax harpax (Fabricius) 3 tegumen, penis; harpax f. hadugli Talbot, valves.

In *punicea* and *jacksoni* the inferior fultura is composed of lobes which are clearly longer than in *bambana*, and in *jacksoni* the subunci are devoid of apophyses.

The early stages of A. amanga have been described by Jackson (1937,  $Trans.\ R$ . ent. Soc. Lond. 86: 219) and Pinhey, 1949: 106. The caterpillar lives on the leaves of Ximenia americana L., in symbiosis with ants (Camponotus niveosetosus Mayr). The caterpillar of the widely distributed A. harpax is so far unknown.

### LIST OF SPECIES OF Axiocerses

\*Axiocerses amanga amanga (Westwood), 1881. Fig. Trimen, 1887.

\*Axiocerses amanga mendeche (Smith), 1889. Fig. Aurivillius in Seitz, 1924. mendeche bistrigata Aurivillius, 1924 (2).

Axiocerses amanga mendeche borealis Aurivillius, 1915.

Axiocerses argenteomaculata Pagenstecher, 1902.

\*Axiocerses bambana Smith, 1900.

mendeche bistrigata Aurivillius, 1924 (3).

Axiocerses baumi Weymer, 1901.

Axiocerses croesus (Trimen), see harpax.

Axiocerses cruenta (Trimen), see punicea.

\*Axiocerses harpax harpax (Fabricius), 1775. Fig. Cramer, 1781 (as perion). tjoane (Wallengren), 1857; croesus (Trimen), 1862; harpax piscatoris Clench, 1943.

\*Axiocerses harpax f. perion (Cramer), 1781.

\*Axiocerses harpax f. kadugli Talbot, 1935, Entomologist's mon. Mag. 71: 120, pl. 2, fig. 9.

Axiocerses harpax efulena Clench, 1963, Jl New York ent. Soc. 71: 183.

Axiocerses harpax styx Rebel, 1908.

Axiocerses harpax piscatoris Clench, see harpax.

Axiocerses harpax ugandana Clench, 1863, l.c.: 184.

- \*Axiocerses jacksoni Stempffer, 1948, Revue fr. Ent. 15: 19, fig. genitalia. Axiocerses maureli Dufrane, 1954, Bull. Annls Soc. R. ent. Belg. 90: 284.
- \*Axiocerses mendeche bistrigata Aurivillius; & see bambana; \( \rightarrow \) see amanga mendeche.
- \*Axiocerses punicea (Smith), 1889. Fig. Trimen, 1894 (as cruenta). cruenta (Trimen), 1894.

Axiocerses tjoane (Wallengren), see harpax.

## Genus LEPTOMYRINA (LEPTOMYRINA) Butler

Leptomyrina Butler, 1898, Proc. zool. Soc. Lond. 1898: 405; Aurivillius, 1898: 335; 1924: 421; Murray, 1935: 93; Pinhey, 1949: 106; Swanepoel, 1953: 171. Type-species: Hesperia phidias Fabricius, 1793, by original designation.

Eyes smooth; vertex and frons clothed with long erect hair; palpi fairly long, horizontal, widely divergent, second segment clothed below with erect bristles, third segment long, acuminate; antennae slender, a little longer than half the length of the costa, white-annulated, club

short, ovoid, abruptly swollen; thorax clothed above with long silky hair and below with close-set white hair; & fore leg with tibia shorter than the femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing apex angular, outer margin slightly convex, inner margin straight; hind wing oval, produced at the anal angle, outer margin slightly scalloped, a filiform tail at the end of vein 1b, a lobe at the anal angle, abdominal margin concave between the lobe and

Wing venation (Text-fig. 307). Fore wing with only 10 veins.

Male genitalia (Text-fig. 151). Uncus composed of two lobes with rounded apices, separated by the depression in the posterior margin of the tegumen; subunci long, robust, bent at an acute angle, tapering gradually to the apex, a short apophysis on the lower edge at the bend; tegumen large, hood-shaped; vinculum very wide with a short saccus; no lower fultura; valves small in relation to the dorsal structures, simple, apical margin serrate, upper margin with a protuberance at the second point of attachment; penis elongate, swollen at the base and slightly expanded at the apex; vesica covered with fine cornuti which give it a shagreened appearance; uncus and distal halves of valves finely pilose.

The male genitalia of Leptomyrina are quite similar to those of certain Hypolycaena. It seems to me therefore that these two genera ought to be placed close together and not far apart as in Aurivillius' classification, in which Leptomyrina is inserted between the group containing Aphnaeus, Spindasis, Axiocerses etc., and the Phasis group, with which they have no true affinity, at least no more than a resemblance in external facies.

The male genitalia of L. hirundo, L. boschi and L. sudanica closely resemble those of L. phidias.

# LIST OF Species of Leptomyrina (Leptomyrina)

- \*Leptomyrina boschi Strand, 1911. Fig. Aurivillius in Seitz, 1924. Leptomyrina handmani Gifford, 1965, Butt. Malawi: 53, figs.
- \*Leptomyrina hirundo (Wallengren), 1857. Fig. Trimen, 1866. Leptomyrina makala Bethune Baker, 1908.

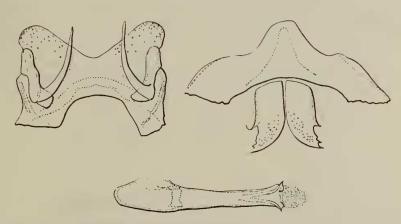


Fig. 151. Leptomyrina (Leptomyrina) phidias (Fabricius), & genitalia.

\*Leptomyrina phidias (Fabricius), 1793. Fig. Mabille, 1885. rabe (Boisduval), 1833.

Leptomyrina rabe (Boisduval), see phidias.

\*Leptomyrina sudanica Stempffer, 1964: 1285, fig.

## LEPTOMYRINA (GONATOMYRINA) Aurivillius

Gonatomyrina Aurivillius, 1924, in Seitz, Macrolep. World 13: 422. Type-species: Papilio lara Linnaeus, 1764, by monotypy.

Only distinguished from other species of *Leptomyrina* by the shape of the hind wing in which the filiform tail at vein 1b is lacking but, on the other hand, the actual anal angle is produced as a triangular prolongation which forms a kind of broad, blunt tail. The anal lobe is absent. These characters are accentuated in *L. gorgias* which Aurivillius regarded as a variety of *lara*.

Male genitalia (Text-fig. 152). Similar to those of other species of Leptomyrina; those of gorgias almost identical with lara.

The early stages of Gonatomyrina have been dealt with in the following publications:—Murray, 1935; Clark and Dickson, 1944, J. ent. Soc. sth. Afr. 7:97 and 1947, 10:128; Pinhey 1949:106. The larvae of lara and gorgias live within the leaves of fleshy plants of the genera Cotyledon, Kalanchoe, Echeveria, Crassula and Mesembryanthemum, feeding on their tissues. It was principally on the characters of larval chaetotaxy that Clark and Dickson specifically separated gorgias and lara (1957, J. ent. Soc. sth. Afr. 20:333).

## LIST OF SPECIES OF Leptomyrina (Gonatomyrina)

\*Leptomyrina (Gonatomyrina) gorgias (Stoll), 1790.

\*Leptomyrina (Gonatomyrina) lara lara (Linne), 1764.

Leptomyrina (Gonatomyrina) lara cana Talbot, 1935, Entomologist's mon. Mag. 71: 121.

Leptomyrina (Gonatomyrina) lara sobrina Talbot, 1935: 121.

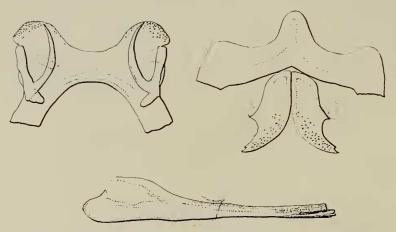


Fig. 152. Leptomyrina (Gonatomyrina) lara lara (Linnaeus), & genitalia.

#### Genus CAPYS Hewitson

Capys Hewitson, 1865, Ill. Diurn. Lep.: 59; Aurivillius, 1898: 337, 1924: 423; Murray, 1935: 36; Swanepoel, 1953: 156. Type-species: Papilio alphaeus Cramer, 1777, by original designation.

Scoptes Hübner, [1819], Verz. bek. Schmett.: 111, an unused senior synonym of Capys is the subject of an application to the Commission, under Article 23 (b), for rejection.

Head broad; eyes large, densely pilose; from clothed with erect hair; palpi horizontal, second segment laterally compressed, third segment short and slender in the  $\beta$ , slender but much longer in the  $\beta$ ; antennae about two-thirds the length of the costa, club fusiform, well differentiated; thorax robust, clothed with long hair;  $\beta$  fore leg with tibia much shorter than femur, tarsus unsegmented, finely spinose below; mid and hind legs with tibiae shorter than femora, metatarsi very long.

Wing shape. Fore wing, apex pointed, outer margin very oblique; hind wing oval, produced at the anal angle, outer margin slightly scalloped, a short obtuse projection at the end of vein 1b. Male secondary sexual characters: on underside of fore wing a tuft of hairs in the middle of the inner margin; on upperside of hind wing a small scaly spot at the origin of vein 7. These characters are present in the type-species, alphaeus, and in disjunctus and penningtoni; they are absent in brunneus and catharus.

Wing venation (Text-fig. 308). Fore wing with 11 veins.

Male genitalia (Text-fig. 153). Uncus composed of two lobes separated by the rounded depression of the distal margin of the tegumen; subunci long, bent at an acute angle, tapering gradually in the apical third; tegumen large, in situ hood-shaped; vinculum narrow with an indistinct saccus; no lower fultura; valves very elongate, narrow, fused together on the lower edge for about half their length, apex slightly hook-shaped; penis elongate, widely open dorsally for almost the whole length of its internal portion, the external portion narrowing gradually to a slightly wider, obliquely truncate apex; vesica with a series of large cornuti and many smaller ones; uncus and upper edges of the distal part of valves clothed with long, fine hair.

The male genitalia of *C. catharus* are almost identical with those of *C. alphaeus*, except for some minor differences in the valves. The male genitalia of these two species of *Capys* resemble those of species of *Deudorix* and indicate a generic affinity to which Hewitson called attention in his original description of *Capys*.

The early stages of *C. alphaeus* and *C. disjunctus* Trimen have been described by Dr. J. Lunt and J. F. Leigh (see Murray, 1935: 97-8) and by C. G. C. Dickson (1947, *J. ent. Soc. sth. Afr.* 10: 128). The larva of *penningtoni* has been described by Pennington (1946, *J. ent. Soc. sth. Afr.* 9: 22).

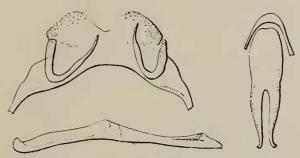


Fig. 153. Capys alphaeus (Cramer), & genitalia.

## LIST OF SPECIES OF Capys

\*Capys alphaeus (Cramer), 1777.

brunneus Aurivillius, 1916.

\*Capys catharus catharus Riley, 1932, Ann. Mag. nat. Hist. (10) 10: 144, fig. (3); Stoneham, 1938, Bull. Stoneham Mus. No. 36: 36 (2).

Capys catharus rileyi Stoneham, 1938, l.c.: 36.

Capys disjunctus disjunctus Trimen, 1895.

Capys disjunctus bamendanus Schultze, 1909.

Capys disjunctus connexivus Butler, 1897.

Capys penningtoni Riley, 1932, Ann. Mag. nat. Hist. (10) 10: 142, fig.

### Genus PHASIS Hübner

Phasis Hübner, 1826, Verz. bekannt. Schmett: 73. Type-species: Papilio salmoneus Cramer, 1781 (Papilio thero Linnaeus, 1764) designated by Scudder, 1875.

Phasis Hübner (partim), Aurivillius, 1898: 337; 1924: 424; Swanepoel, 1953: 121.

Pseudocapys Murray, 1935: 106. Type-species, by original designation, Papilio thero Linnaeus.

Murray (1935: 103) erroneously attempted to designate, as type-species of *Phasis*, *Papilio pierus* Cramer, which was not one of the originally included species (see *Aloeides*). *Pseudocapys* Murray is an objective junior synonym of *Phasis* Hubner.

 $\it Head$  rather broad; eyes glabrous; frons clothed with long, erect hair; palpi long, second segment robust, ascending, clothed with scales, third segment rather long, slender; antennae about half the length of the costa, robust, gradually increasing in thickness to the clavate end; thorax robust, clothed below with woolly hair;  $\it 3$  fore leg with tarsus unsegmented.

Wing shape. Fore wing with costa arched at the base then straight, apex truncate, outer margin straight from the apex to vein 5, deeply concave between veins 5 and 3 and convex

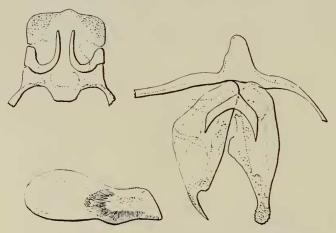


Fig. 154. Phasis thero thero (Linnaeus), & genitalia.

from 3 to the inner angle; hind wing subtriangular, costa convex at the base, outer margin scalloped at the ends of vein 5 to 3, a short, rounded tail at the end of vein 2 and another slightly longer at the end of vein 1b, a small, weak lobe at the anal angle.

Wing venation (Text-fig. 309). Fore wing with 12 veins.

Male genitalia (Text-fig. 154). Uncus subrectangular, the distal margin rounded at the sides and with a slight median, obtuse protuberance; subunci long and robust, strongly curved, tapering evenly to the apices; tegumen oval, its proximal edge jutting out towards segment 8; in situ uncus and tegumen together are hood-shaped; vinculum narrow, with a robust triangular saccus; lower fultura crescentic; valves oblong, subtriangular, with rounded, serrated apices, the edges of the upper processes folded over the inner surface and connected by a membrane which sheathes the penis, as in species of Aphnaeus, Spindasis, Axiocerses, etc.; penis short and massive, swollen in its internal portion and with obliquely truncate apex; vesica enclosing numerous long delicate spines; uncus and distal portions of the valves clothed in fine, delicate hair.

The male genitalia of sardonyx and argyraspis are of the same type as those of P. there although the saccus is less developed. The lower fultura is shield-shaped in argyraspis; but composed of two long digitate processes in sardonyx.

The early stages of *P. thero* have been well studied by Gowan C. Clark, 1942, *J. ent. Soc. sth. Afr.* 5: 111-115, pl. 3; the caterpillar feeds on *Rhus longisperma*, in association with the ants called "cocktailed ants".

### LIST OF SPECIES OF Phasis

\*Phasis argyraspis (Trimen), 1873.

Phasis argyraspis f. labuschagnei van Son, 1959, Koedoe 2:56.

Phasis erosine (Fabricius), see thero.

Phasis pulsius (Herbst), see thero.

Phasis rumina (Drury), see thero.

Phasis salmoneus (Cramer), see thero.

\*Phasis sardonyx (Trimen), 1868.

Phasis sardonyx f. peringueyi Aurivillius, 1924: 430.

Phasis sardonyx f. knobeli van Son, 1959, Koedoe 2:56.

\*Phasis thero thero (Linne), 1764. Fig. Drury, 1773 (as rumina).

rumina (Drury), 1773; salmoneus (Cramer), 1781; erosine (Fabricius), 1787; pulsius (Herbst), 1793.

Phasis thero clavum Murray, 1935: 104, fig. (regarded by Swanepoel, 1953, as a distinct species).

### Genus ALOEIDES Hübner

Aloeides Hübner, 1826, Verz. bekannt. Schmett: 73. Type-species: Papilio pierus Cramer, 1779, selected by Scudder, 1875.

Phasis Hübner (partim); Aurivillius, 1898: 337; 1924: 424; Murray, 1935: 113; Swanepoel, 1953: 125.

Head rather broad; eyes glabrous; from clothed with erect hair; palpi long, second segment ascending, laterally compressed, clothed below with close-set long scales, third segment short and conical; thorax robust, clothed below with close-set hair; 3 fore leg with tibia

shorter than femur, tarsus unsegmented and finely spinose below; mid and hind legs with tibiae shorter than femora.

Wing shape. Fore wing with costa arched at the base, then straight, apex pointed, outer margin slightly convex; hind wing oval, apex rounded, outer margin slightly scalloped at the ends of veins 2 and 3, a small rounded projection at the end of vein 1b. Wing shape is not uniform in the genus Aloeides. In wallengreni the anal angle is much more produced. On the other hand, in barklyi the hind margin of the hind wings is almost evenly rounded.

Wing venation (Text-fig. 310). Veins 6 and 7 of the fore wings arise from a short common stalk.

Male genitalia (Text-fig. 155). Uncus trapezoidal, distal margin feebly convex and forming the long base of the trapezoid; subunci robust, bent near the base and tapering gradually to their apices; the proximal edge of the tegumen forms a large rounded projection directed towards the eighth abdominal segment, in situ uncus and tegumen together are hood-shaped;

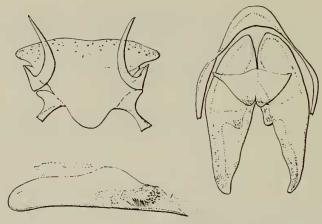


Fig. 155. Aloeides pierus (Cramer), & genitalia.

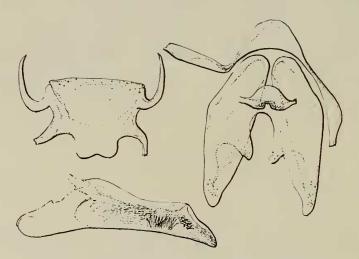


Fig. 156. Aloeides thyra thyra (Linnaeus), of genitalia.

vinculum rather narrow, with a rounded saccus; lower fultura a triangular plate with a shallow notch at the apex; valves oblong with rounded apices, their upper processes folded over the inner surface, and connected by a membrane which passes over the penis; penis robust, widely open dorsally in its internal portion, swollen in the middle and with obliquely truncate apex; vesica enclosing small cornuti and delicate spines; distal margins of uncus and valves sparsely pilose.

As in the type-species, A. pierus, the distal margin of the uncus is straight or feebly convex in aranda, damarensis, molomo, orthrus and thyra (Text-fig. 156). In barklyi it is very convex. On the other hand it is slightly concave in taikosama and almeida. In wallengreni (Text-fig. 157) the uncus is distinctly different in form, being subrectangular with a shallow depression on the distal margin, lateral angles rounded and sides straight, the subunci angled further from the base, equally thick throughout, instead of tapered and ending in a blunt tip. A. malagrida approaches wallengreni in the form of its uncus.

The larva of A. pierus has been described by Dickson (1945, J. ent. Soc. sth. Afr. 8: 161). It lives on a species of Aspalathus (Leguminosae).

### LIST OF SPECIES OF Aloeides

Aloeides aglaspis (Trimen), see malagrida.

\*Aloeides almeida (Felder), 1862. Fig. Felder, 1865, Reise Novara.

\*Aloeides aranda aranda (Wallengren), 1857. Fig. Aurivillius in Seitz, 1924. mars (Trimen), 1862; pierus var. A. Trimen, 1866.

Aloeides aranda f. rougemonti (Oberthür), 1910.

Aloeides aranda zilka Smith, 1900.

\*Aloeides barklyi (Trimen), 1874.

\*Aloeides conradsi (Aurivillius), 1907. Fig. Aurivillius in Seitz, 1924.

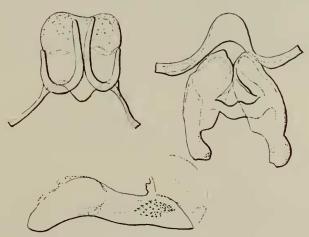


Fig. 157. Aloeides wallengreni (Trimen), & genitalia.

Aloeides conradsi f. ochraceus (Joicey & Talbot), 1924, Bull. Hill Mus. Witley 1:546.

\*Aloeides damarensis (Trimen), 1891.

Aloeides damarensis f. punctata (Aurivillius), 1924.

Aloeides euadrus (Fabricius), 3, see pierus; 9, see thyra.

Aloeides griseus Riley, 1921, Trans. ent. Soc. Lond. 1921: 251, fig.

\*Aloeides malagrida (Wallengren), 1854. Fig. Murray, 1935. aglaspis (Trimen), 1862.

Aloeides mars (Trimen), see aranda.

Aloeides marshalli (Aurivillius), 1924.

\*Aloeides molomo molomo (Trimen), 1870.

Aloeides molomo mumbuensis Riley, 1921, Trans. ent. Soc. Lond. 1921: 250, fig.

Aloeides molomo kiellandi Carcasson, 1961, Occ. Pap. Coryndon meml Mus. No. 7: 19, fig.

Aloeides nycetus (Cramer), see thyra.

\*Aloeides orthrus (Trimen), 1874.

\*Aloeides pierus (Cramer), 1779.

euadrus (Fabricius), 1787 (3); suetonius (Fabricius), 1793.

Aloeides pierus var. A Trimen, see aranda.

Aloeides pierus var. B Trimen, see taikosama.

Aloeides simplex (Trimen), 1893.

Aloeides suetonius (Fabricius), see pierus.

\*Aloeides taikosama (Wallengren), 1857. Fig. Aurivillius in Seitz, 1924. pierus var. B Trimen, 1866.

\*Aloeides thyra thyra (Linnaeus), 1764. Fig. Hübner, Samml. Exot. Schmett. 1816–24.

nycetus (Cramer), 1781; enadrus (Fabricius), 1787, Q.

Aloeides thyra f. egerides (Riley), 1938, Trans. R. ent. Soc. Lond. 87: 238.

\*Aloeides thyra f. pallida (Riley), 1938: 238.

\*Aloeides thyra dentatis (Swiestra), 1909.

Aloeides thyra maseruna (Riley), 1938, Trans. R. ent. Soc. Lond. 87: 239, fig.

\*Aloeides wallengreni (Trimen), 1887. Fig. Trimen, 1866 (as malagrida).

### Genus POECILMITIS Butler

Poecilmitis Butler, 1899, Entomologist 32:78. Type-species: Zeritis lycegenes Trimen, by original designation.

Phasis Hübner (partim); Aurivillius, 1898; 340; 1924; 430; Murray, 1935; 107; Swanepoel, 1953; 135.

Eyes glabrous, palpi long, ascending, second segment very long, laterally compressed, clothed below with erect hair, third segment short, laterally compressed, apex blunt; antennae about half the length of the costa, becoming gradually stouter right up to the poorly differentiated club; of fore leg with tibia shorter than femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing, costa arched at the base, then straight, apex slightly rounded, outer margin convex; hind wing oval, apex rounded, outer margin slightly scalloped at the endings of the veins.

The shape of the hind wing is not constant in the genus *Poecilmitis*. Whilst in *aethon, dicksoni*, and *pyroeis* it is approximately the same as in the type-species, there is a short broad tail at vein 1b in *chrysaor*, *palmus* and *felthami* which is more or less pronounced also in the *thysbe* group.

Wing venation (Text-fig. 311). Fore wing with 12 veins; 6 and 7 from the upper angle of the cell.

Male genitalia (Text-fig. 158). Uncus roughly rectangular with broadly rounded angles and a shallow median depression in the distal margin; subunci long, curved; robust, with a short apophysis on the lower edge of the curve, apex blunt; tegumen oval; vinculum rather narrow with a short rounded saccus; lower fultura small, shield-shaped, notched apically; valves oblong, with apex digitate, the upper processes connected on their inner surfaces by a thin membrane which surrounds the penis; penis long; curving, swollen at its base, apex obliquely truncate; uncus and distal portion of valves sparsely pilose.

In P. thysbe, P. palmus, P. felthami, P. chrysaor and P. aethon, the subunci bear an apophysis as in lycegenes. This is lacking in dicksoni.

In aethon and felthami the saccus is rounded and not prominent, but in thysbe, palmus and chrysaor it is quadrangular and very prominent.

The early stages of thysbe, palmus, chrysaor and pyroeis have been described by Dickson 1943 (J. ent. Soc. sth. Afr. 6:37; 1944, 7:97; 1945, 8:99; 1947, 9:178, 10:128; 1948, 11:50).

### LIST OF SPECIES OF Poecilmitis

Poecilmitis adonis Pennington, 1962, J. ent. Soc. sth. Afr. 25: 277, fig.

\*Poecilmitis aethon (Trimen), 1887.

Poecilmitis aridus Pennington, 1953, J. ent. Soc. sth. Afr. 16: 104, fig.

Poecilmitis atlantica Dickson, 1966, Entomologist's Rec. J. Var. 78: 181, fig.

Poecilmitis beaufortia Dickson, 1966, l.c., 78: 109, fig.

\*Poecilmitis chrysaor (Trimen), 1864. Fig. Trimen, 1887.

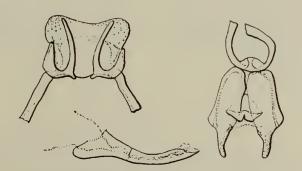


Fig. 158. Poecilmitis lycegenes (Trimen), & genitalia.

\*Poecilmitis dicksoni (Gabriel), 1947, Entomologist 80: 60.
Poecilmitis endymion Pennington, 1962, J. ent. Soc. sth. Afr. 25: 274, fig.

\*Poecilmitis felthami (Trimen), 1904.

\*Poecilmitis lycegenes (Trimen), 1874.

Poecilmitis lycia Riley, 1938, Trans. R. ent. Soc. Lond. 87: 242, fig.

Poecilmitis lyncurium (Trimen), 1868. Fig. Trimen, 1887.

Poecilmitis lysander Pennington, 1962, J. ent. Soc. sth. Afr. 25: 275, fig.

Poecilmitis midas Pennington, 1962: 272, fig.

Poecilmitis nais (Cramer), see thysbe.

\*Poecilmitis nigricans (Aurivillius), 1924. Fig. Murray, 1935.

\*Poecilmitis palmus (Cramer), 1781.

Poecilmitis pan Pennington, 1962, J. ent. Soc. sth. Afr. 25: 273, fig.

Poecilmitis pelion Pennington, 1953, J. ent. Soc. sth. Afr. 16: 106, fig.

Poecilmitis penningtoni Riley, 1938, Trans. R. ent. Soc. Lond. 87: 239, fig.

Poecilmitis phosphor (Trimen), 1864. Fig. Trimen, 1866.

Poecilmitis pyramus Pennington, 1953, J. ent. Soc. sth. Afr. 16: 105, fig.

Poecilmitis pyroeis (Trimen), 1864. Fig. Trimen, 1866.

Poecilmitis splendens (Swainson), see thysbe.

Poecilmitis swanepoeli Dickson, 1965, J. ent. Soc. sth. Afr. 27: 160, figs.

\*Poecilmitis thysbe thysbe (Linn.), 1764. Fig. Butler, 1868. nais (Cramer), 1775; splendens (Swainson), 1833.

\*Poecilmitis thysbe osbecki (Aurivillius), 1882. Fig. genitalia, Stempffer, 1945, Ann. Soc. ent. Fr.: 82.

Poecilmitis thysbe brooksi Riley, 1938, Trans. R. ent. Soc., 87: 241, fig.

Poecilmitis thysbe trimeni Riley, 1938: 240, fig.

Poecilmitis turneri Riley, 1938: 241, fig.

Poecilmitis uranus Pennington, 1962, J. ent. Soc. sth. Afr. 25: 277, fig.

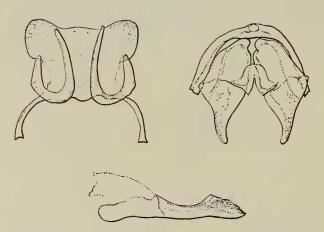


Fig. 159. Chrysoritis oreas (Trimen), & genitalia.

### Genus CHRYSORITIS Butler

Chrysoritis Butler, 1898, Proc. zool. Soc. Lond. 1897: 848. Type-species: Zeritis oreas Trimen, by original designation.

Phasis Hübner (partim); Aurivillius, 1898: 340; 1924: 431; Murray, 1935: 107, 113, 116; Swanepoel, 1953: 127, 144, 146.

Eyes glabrous; vertex clothed with long brown erect hair; palpi long, second segment extending well beyond the frons, clothed above with adpressed scales and below with long white hair, third segment very short with blunt tip; antennae about half the length of the costa, becoming gradually stouter right up to the poorly differentiated club, which is elongate and bright orange beneath; thorax robust, clothed below with white hair; 3 fore leg, femur flattened, tibia shorter than femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing costa almost straight, apex somewhat rounded, outer margin strongly convex, hind wing with a small rounded projection at the end of vein 1b.

Wing venation (Text-fig. 312). Fore wing with 11 or 12 veins.

In his description of *Poecilmitis*, Butler stated that *oreas* has only 11 veins in the fore wing. This assertion is repeated by Aurivillius (1924: 429); it is, however, incorrect. A close examination reveals that vein 8 is present, though short and not very apparent; on the other hand vein 8 is lacking in *zeuxo* and *chrysantas*, which in consequence have only 11 veins in the fore wing. It is quite possible that vein 8 was absent in the specimen examined by Butler, the character being a variable one.

Male genitalia (Text-fig. 159). Uncus subrectangular, the distal edge slightly depressed in the middle, lateral angles broadly rounded; subunci long, robust, bent at about two-fifths of their length from the base, devoid of apophyses, apex blunt, tegumen subrectangular, in situ uncus and tegumen together hood-shaped; vinculum narrow, no saccus, lower fultura shield-shaped with a deep apical notch; valves oblong with rounded apices, their upper processes connected by a membrane which surrounds the penis, as is the case throughout the *Phasis* group; penis elongate, swollen in the middle, the external portion dilated just before the obliquely truncated apex; uncus and distal parts of the valves pilose.

The male genitalia of *zeuxo* and *chrysantas* are of the same type as those of *oreas*, but the distal margin of the uncus, instead of being concave, bears a weak median prominence, the subunci have an apophysis and the saccus is quadrangular and very prominent.

The early stages of *C. zeuxo* have been described by Dickson (1952, *Trans. R. Soc. S. Afr.* **23**: 447, fig.).

The two genera, *Poecilmitis* and *Chrysoritis*, are not sharply defined. The typespecies of *Chrysoritis* (oreas) has 12 veins in the fore wing, like *Poecilmitis*. The two genera could well be united.

# LIST OF SPECIES OF Chrysoritis

- \*Chrysoritis chrysantas (Trimen), 1868.
- \*Chrysoritis oreas (Trimen), 1891. Fig. Trimen, 1906.
- \*Chrysoritis zeuxo zeuxo (Linnaeus), 1764. Fig. Trimen, 1866.
  - Chrysoritis zeuxo zonarius (Riley), 1938, Trans. R. ent. Soc. Lond. 87: 239, fig.

## Genus CRUDARIA Wallengren

Crudaria Wallengren, 1875, Öfvers K. svenska Vetensk Akad. Förh. 32: 86; Aurivillius, 1924: 431; Swanepoel, 1953: 149. Type-species: Arhopala leroma Wallengren, by monotypy. Phasis Hübner (partim); Aurivillius, 1898: 343; Murray, 1935: 106.

Head rather broad; eyes glabrous, palpi long, ascending, second segment laterally compressed, clothed with long scales and hair, third segment long, slender, acuminate; antennae slightly more than half the length of the costa, becoming gradually stouter up to the poorly differentiated fusiform club; thorax rather robust, thickly clothed below with white hair; of fore leg with tibia shorter than femur, tarsus rather slender, unsegmented, finely spinose below; mid and hind legs with tibiae shorter than femora.

Wing shape. Fore wing with costa arched at its base then straight, apex angular, outer margin convex between apex and vein 4, then straight; hind wing oval, outer margin rounded, a short delicate tail at the end of vein 1b a small lobe at the anal angle.

Wing venation (Text-fig. 313). Fore wing with 11 veins; hind wing cell short.

Male genitalia (Text-fig. 160). Uncus composed of two large semicircular lobes separated by the rounded depression on the distal edge of the tegumen; subunci much reduced, curved, apex blunt, with a short rounded apophysis on the lower margin, tegumen large; vinculum fairly broad with a rounded saccus; lower fultura consists of a triangular plate with deeply notched base; valves oblong with digitate apices, their upper margins connected in the middle by a membrane; penis elongate, robust, swollen at the base, apex obliquely truncate; uncus and apices of valves clothed with long, fine hair.

The early stages of Crudaria leroma have been described by Gowan Clarke (1958, J. ent. Soc. sth. Afr. 24: 127).

#### LIST OF SPECIES OF Crudaria

Crudaria capensis van Son, 1956, Ann. Transv. Mus. 22: 505. Crudaria delagoensis (E. Sharpe), see leroma. \*Crudaria leroma leroma (Wallengren), 1857. Fig. Trimen, 1870. zorites (Hewitson), 1874; delagoensis (E. Sharpe), 1891. Crudaria leroma albomaculate Aurivillius, 1924: 432. Crudaria zorites (Hewitson), see leroma.

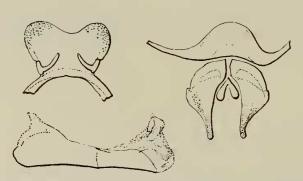


Fig. 160. Crudaria leroma leroma (Wallengren), & genitalia.

#### Genus ERIKSSONIA Trimen

Erikssonia Trimen, 1891, Proc. zool. Soc. Lond. 1891: 91; Aurivillius, 1898: 343; 1924: 432. Type-species: Erikssonia acraeina Trimen, by monotypy.

Head small, downy; eyes glabrous; palpi long, parallel, horizontal, second segment laterally compressed, clothed below with long white scales, third segment long, acuminate; antennae rather short, thick, with a blunt ill-defined club; thorax short and slender; 3 fore leg, tibia with a strong apical spur, tarsus longer than the tibia, unsegmented, ending in a long sharp pointed claw; mid and hind legs, tibiae with a short spur, metatarsi longer than the tibiae, terminal claws large and strong.

Wing shape. Fore wing with costa weakly arched at the base then straight, outer margin convex; hind wing oval, apex rounded, outer margin slightly scalloped at the ends of veins 3 and 2, an obtuse projection at the end of vein 1b.

Wing venation (Text-fig. 314). Fore wing with 12 veins.

Male genitalia (Text-fig. 161). Uncus crescent-shaped; subunci long, curved, robust at the base then slender to the blunt apex; tegumen with convex proximal edge; uncus and tegumen together hood-shaped; vinculum rather narrow with a weak rounded saccus; lower fultura shield-shaped with a deep notch in the upper edge; valves oblong with rounded apices, their upper processes connected together by a membrane which surrounds the penis; penis swollen at its base, apex obliquely truncate; vesica armed with large cornuti; uncus and apices of valves sparsely pilose.

The male genitalia of E. cooksoni are of the same type as those of E. acraeina.

In his generic description Trimen wrote "Erikssonia exhibits considerable divergence from the typical groups of the family and is probably best placed between Zeritis and Mimacraea, but nearer to the former than the latter". It is true that Erikssonia belongs to the same subfamily as Zeritis, Axiocerses, Phasis etc. but on the other hand it has no affinity whatever with Mimacraea. Trimen mentions that E. acraeina bears a superficial resemblance to certain species of Acraea, e.g. to Acraea buxtoni on the upperside and to A. axina and A. atergatis on the underside. He attributes this resemblance to mimicry.

### LIST OF SPECIES OF Erikssonia

- \*Erikssonia acraeina Trimen, 1891.
- \*Erikssonia cooksoni H. H. Druce, 1905.

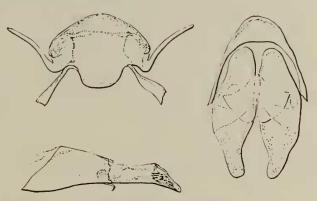


Fig. 161. Erikssonia acraeina Trimen, & genitalia.

#### Genus THESTOR Hübner

Thestor Hübner, 1823, Verz. beh. Schmett. (5): 73; Swanepoel, 1953: 149. Type-species: Papilio petalus Cramer, 1779 (Papilio protumnus Linnaeus, 1764), designated by Scudder (1875, Proc. Am. Acad. Arts Sci. 10: 281).

Arrugia Wallengren, 1872, Öfvers. K. svenska VetenskAkad. Förh. 29:47; Aurivillius, 1898: 343; 1924:433; Murray, 1933:99.

Head small; eyes glabrous; palpi long, extending far beyond the frons, second segment clothed below with white adpressed scales, third segment with a blunt apex, palpi longer in  $\mathcal Q$  than in  $\mathcal J$ ; antennae very short, thick shaft gradually increasing in thickness from the base to the undifferentiated club, which has a rounded apex; thorax robust; abdomen long and thick, especially in the  $\mathcal Q$ ; legs short, scaly, tibiae much shorter than femora and without apical spurs;  $\mathcal J$  fore tarsus distinctly five-segmented like that of the female, and with two curved terminal claws.

Wing-shape. Fore wing costa straight, apex slightly rounded, outer margin slightly convex, inner margin straight and much shorter than the costa; hind wing oval, outer margin convex, anal angle very rounded. In their massive build and dull brown and black colours, the species of *Thestor* have more the appearance of Hesperiidae than Lycaenidae.

Wing venation (Text-fig. 315). Fore wing with 11 veins.

Male genitalia (Text-fig. 162). Uncus a narrow band whose distal edge bears in the middle two very long recurved processes, each with a sharp pointed apex; subunci long, curved, rather slender; tegumen subrectangular; vinculum narrow with a saccus; above the penis are two subtriangular processes which have a blunt and slightly serrate apex, and are connected by a membrane both to the dorsal section of the vinculum and to the middle of the dorsal edge of the valves [Van Son, in his paper on some species of Thestor (1957, Ann. Transv. Mus. 21:442, fig.) gave these processes the name of "Labiles". I believe that they correspond to the upper fultura]; lower fultura crescentic, fused to the base of valves, valves oblong, distal portion widened, with an almost straight terminal edge; penis elongate, robust at the base, curved, tapering gradually in its external portion, vesica clothed with numerous cornuti which give it a shagreened appearance; uncus almost glabrous, distal portion of valves pilose.

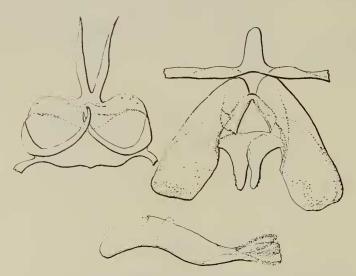


Fig. 162. Thestor protumnus protumnus (Linnaeus), & genitalia.

I have examined the male genitalia of Thester brachycora, T. obscurus, T. holmesi and T. strutti. Bethune Baker (1914, Trans. ent. Soc. Lond. 1914: 333) described and figured those of T. basuta. van Son (1949, Ann. Transv. Mus. 21: 215) dealt with T. penningtoni and the same author (1951, Ann. Transv. Mus. 21: 440) described those of dukei, obscurus, holmesi, penningtoni and strutti. Pennington (1962, J. ent. Soc. sth. Afr. 25: 281) figured the male genitalia of vansoni, petra, tempe, rileyi and obscurus. A comparison of these works shows clearly that all the species examined belong to the same type as T. protumnus, the type-species of the genus. The genus Thestor seems to be at the same time both very specialized and very homogeneous.

Bethune Baker (1914, Trans. ent. Soc. Lond. 1914: 333) found some resemblance between the dorsal elements of T. basuto and of Mimacraea. I do not believe, however, that there is any affinity between these two genera. The uncus in Mimacraea is asymmetrical; in Thestor it is not. In Mimacraea the tegumen is extremely reduced and subunci are absent; they are well developed in Thestor. The only resemblance lies in the two long curved and pointed processes of the uncus, which are a quite spectacular character, but not one to which any considerable taxonomic importance should, in my opinion, be attributed, since it recurs in various degrees of development in several other widely separated groups, for example in Myrina ficedula and certain species of palaearctic Theclinae such as icana, butleri and attilia.

The early stages of E. protumnus, E. basuta, E. brachycera and E. obscurus have been described by Murray (1935: 99), Dickson (1945, J. ent. Soc. sth. Afr. 8: 151) and, in the same journal, by Clark (1960, 23:279,282).

### LIST OF SPECIES OF Thestor

Thestor basuta (Wallengren), 1857. Fig. Trimen, 1887.

zaraces (Hewitson), 1874.

\*Thestor brachycera (Trimen), 1883. Fig. Trimen, 1887.

Thestor braunsi van Son, 1941, J. ent. Soc. sth. Afr. 4: 186.

Thestor dicksoni dicksoni Riley, 1954, Entomologist 87: 98, fig.

Thestor dicksoni calviniae Riley, 1954: 101, fig.

Thestor dukei van Son, 1951, Ann. Transv. Mus. 21: 439, fig.

\*Thestor holmesi van Son, 1951: 441, fig.

Thestor montanus montanus van Son, 1941, J. ent. Soc. sth. Afr. 4: 191.

Thestor montanus pictus van Son, 1941: 192.

Thestor murrayi Swanepoel, 1953, J. ent. Soc. sth. Afr. 16: 191, fig.

\*Thestor obscurus van Son, 1941, J. ent. Soc. sth. Afr. 4: 186.

Thestor penningtoni van Son, 1949, Ann. Transv. Mus. 21: 214, fig.

Thestor petalus (Cramer), see protumnus.

Thestor petra Pennington, 1962, J. ent. Soc. sth. Afr. 25: 280, fig.

\*Thestor protumnus protumnus (Linnaeus), 1764. Fig. Cramer, 1779 (as petalus).

petalus (Cramer), 1779; silvius (Fabricius), 1787.

Thestor protumnus aridus van Son, 1941, J. ent. Soc. sth. Afr. 4: 188.

Thestor rileyi Pennington, 1956, J. ent. Soc. sth. Afr. 19: 33, fig.

Thestor silvius (Fabricius), see protumnus.

\*Thestor strutti van Son, 1951, Ann. Transv. Mus. 21:444, fig.

Thestor tempe Pennington, 1962, J. ent. Soc. sth. Afr. 25: 282, fig.

Thestor vansoni Pennington, 1962: 278, fig.

Thestor zaraces (Hewitson), see basuta.

#### Genus SPALGIS Moore

Spalgis Moore, 1879, Proc. zool. Soc. Lond. 1879: 137; Aurivillius, 1898: 344; 1924: 433. Type-species: Lucia epeus Westwood 1851 (an Indo-Malayan species) by monotypy.

Eyes glabrous; palpi long, second segment laterally much compressed, extending well beyond the frons, third segment short, also laterally compressed, with pointed apex; antennae less than half the length of the costa, thickening gradually to an ovoid club which is not clearly differentiated from the shaft; thorax moderately robust; abdomen long, reaching slightly beyond the anal angle of the wing;  $\Im$  fore leg with tibia shorter than femur, tarsus unsegmented, finely spinose below; mid and hind tibiae slightly shorter than the femora.

Wing shape. Fore wing with costa arched at its base, then straight, apex angular, outer margin slightly convex; hind wing subtriangular, costa arched, apex rounded, outer margin slightly convex, anal angle well marked.

Wing venation (Text-fig. 376). Fore wing with 11 veins.

Male genitalia (Text-fig. 163). Uncus pentagonal with an obtuse median projection on the terminal edge, the angles rounded; subunci short with very robust bases, curving, the free portion tapering gradually to a pointed apex; tegumen subrectangular, in situ uncus and tegumen are hood-shaped; vinculum moderately wide, without saccus; lower fultura composed of two long digitate processes, swollen at their bases, their blunt apices level with the apices of the valves; valves oblong with a small tooth at the apex, distally connected by a membrane; penis very elongate, slender, slightly curved, uncus clothed with short hair, longer hair on the apices of the valves.

The armature of S. lemolea resembles that of S. epeus; in S. tintinga the subunci are less massive basally, evenly curved, and taper progressively to a sharp-pointed tip.

The caterpillar of S. lemolea has been described by W. A. Lamborn (1911, Proc. ent. Soc. Lond. 1911: 105, and 1913, Trans. ent. Soc. Lond. 1913: 475) and also by

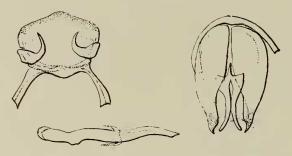


Fig. 163. Spalgis epius (Westwood), & genitalia.

T. H. E. Jackson (1937, Trans. R. ent. Soc. Lond. 86: 220). It feeds on Coccidae of the genus Dactylopius.

# LIST OF SPECIES OF Spalgis

Spalgis docus (Druce), see tintinga.

Spalgis latimarginata E. Sharpe, see lemolea.

\*Spalgis lemolea H. H. Druce, 1890. Fig. Aurivillius in Seitz, 1924. latimarginata E. Sharpe, Oct. 1890; s-signata Holland, Nov. 1890.

Spalgis pilos H. H. Druce, 1890.

Spalgis s-signata Holland, see lemolea.

\*Spalgis tintinga (Boisduval), 1833. Fig. Mabille, 1887. docus (Druce), 1875.

#### Genus CUPIDESTHES Aurivillius

Cupidesthes Aurivillius, 1895, Ent. Tidskr. 16: 215; 1898: 345; 1924: 435; Bethune Baker, 1910: 7. Type-species: Cupidesthes robusta Aurivillius, by monotypy.

Frons black with two white lateral lines; eyes glabrous or sparsely pilose; palpi long, ascending, second segment laterally compressed, clothed below with white adpressed scales, third segment slender, acuminate; antennae slender, a little longer than half the length of the costa, club ovoid, well differentiated; thorax more robust than in Anthene, clothed below with long white hair; of fore leg, femur clothed with white hair, tibia shorter than the femur, tarsus long, unsegmented.

Wing shape. Fore wing triangular, costa almost straight, outer margin slightly convex; hind wing abdominal margin slightly concave before the anal angle.

Wing venation (Text-fig. 317). Fore wing with 11 veins; 10 and 11 free from the upper edge of the cell.

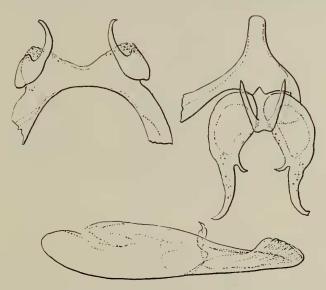


Fig. 164. Cupidesthes robusta Aurivillius, & genitalia.

Male genitalia (Text-fig. 164). Uncus composed of two small lobes fused to either side of the tegumen; subunci short and robust, bent close to their massive bases; tegumen a rather wide ribbon; vinculum continuing as a long robust saccus; lower fultura shaped like a furca, with straight divergent arms; valves elongate, oval in their basal halves, tapered and slightly recurved in their distal halves; penis very robust, the internal portion almost ovoid, the external portion rather short, tapering, dilated at the apex; uncus and distal portions of the valves sparsely pilose.

The male genitalia of the other species of *Cupidesthes* differ in varying degree from those of *C. robusta*, not in general plan, but in the relative dimensions and shapes of the parts.

In Cupidesthes arescopa (Text-fig. 165) the subunci are extremely long and slender, the tegumen is reduced to a narrow ribbon in the median area, the valves are digitate and little swollen basally, and the penis is very long, tapering and dilated apically.

The genitalia of C. voltae, C. thyrsis, C. lithas and C. paralithas are rather like those of C. arescopa, although the subunci are shorter in thyrsis and the valves are not so tapered in the other species.

Bethune Baker did not examine the genitalia of *robusta*, but only those of *voltae*, *thyrsis*, *lithas* and *arescopa*, which explains his statement (l.c.: 5) that "the genitalia are also very different from those of the following genera, the clasps being of a totally different structure as will be seen from the descriptions of these organs".

Other species of *Cupidesthes* present intermediate characters. In *C. irumu* the valves are digitate, very narrow, the penis slender, but the subunci are only moderately long and the posterior margin of the tegumen is deeply excavate. In *C. cuprifascia* the subunci are very long, the penis slender, but the valves are rather short and moderately wide. In *C. leonina* the subunci are only of moderate length, the penis less long than in the foregoing species, the valves rather broad with a deeply cleft apex and the upper process ends in a long sharp point.

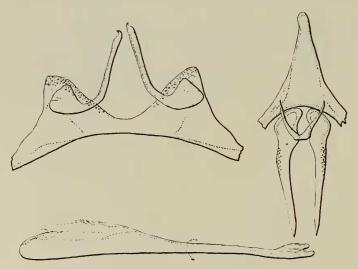


Fig. 165. Cupidesthes arescopa arescopa Bethune Baker, & genitalia.

C. wilsoni (Text-fig. 166) differs again: posterior margin of tegumen deeply excavate on the median line, subunci fairly long, robust, bent, basally massive, vinculum rather broad, with a tapering saccus, lower fultura with small divergent branches; valves oval with rounded apex, the upper process with very convex dorsal margin, the lower process almost straight and bearing a fine sharp tooth at three-fifths its length from base; penis long and rather slender, slightly curved; its faintly expanded tip obliquely truncate.

It is evident that in *Cupidesthes* there are patterns of male genitalia intermediate between those that Bethune Baker considered typical of the genus (arescopa, thyrsis, etc.) and those of the large genus Anthene. Again, it is to be noted that, if on the one hand robusta, voltae, arescopa and wilsoni are large robust insects, on the other hand other species, such as thyrsis, leonina, and irumu have the stature and structure of most species of Anthene. The "genus" Cupidesthes seems but feebly differentiated and might well be reduced to the rank of a subgenus.

The larva of *C. wilsoni* has been described by Jackson (1937, *Trans. R. ent. Soc. Lond.* **86**: 220). It lives in ants' nests in hollow trees of *Acacia abyssinica* and *Entada abyssinica* (Mimosaceae).

## LIST OF SPECIES OF Cupidesthes

Cupidesthes albida (Aurivillius), 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910-11, 1:1232.

\*Cupidesthes arescopa arescopa Bethune Baker, 1910, fig. and fig. genitalia.

\*Cupidesthes arescopa orientalis Stempffer, 1962: 1169, fig. Cupidesthes brunneus (Smith & Kirby), see paludicola.

\*Cupidesthes caerulea Jackson, 1965, Ann. Mag. nat. Hist. (13) 8:531, figs.

\*Cupidesthes cuprifascia Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1:95 (3). Fig. genitalia, Stempffer, 1950 Revue fr. Ent. 17:141. Fig. Q, Stempffer 24:1170.

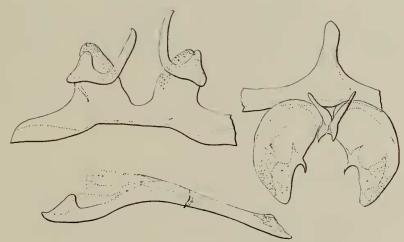


Fig. 166. Cupidesthes wilsoni Talbot, & genitalia.

Cupidesthes hilarion Hulstaert, 1924, Revue zool. afr. 12: 129.

\*Cupidesthes irumu Stempffer, 1948, Revue fr. Ent. 15: 192, fig. genitalia.

\*Cupidesthes leonina (Bethune Baker), 1903.

\*Cupidesthes lithas (H. H. Druce), 1890. Fig. genitalia, Bethune Baker, 1910. Cupidesthes mimetica (H. H. Druce), 1910.

Cupidesthes minor Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1: 95, fig.

Cupidesthes paludicola (Holland), 1891.

brunneus (Smith & Kirby), 1893, fig.

- \*Cupidesthes paralithas Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 399.
- \*Cupidesthes robusta Aurivillius, 1895. Fig. Bethune Baker, 1910. Fig. genitalia, Stempffer, 1945, Annls Soc. ent. Fr. 94:83.
- \*Cupidesthes thyrsis (Hewitson), 1878. Fig. genitalia, Bethune Baker, 1910.

Cupidesthes thyrsis Q.-f. unicolor (Aurivillius), 1924.

Cupidesthes vidua Talbot, 1929, Bull. Hill Mus. Witley 3: 140, fig.

\*Cupidesthes voltae voltae (E. Sharpe), 1890. Fig. and fig. genitalia, Bethune Baker, 1910.

Cupidesthes voltae gabunica (Aurivillius), 1899.

\*Cupidesthes wilsoni Talbot, 1935, Entomologist's mon. Mag. 71: 121, fig. (2); 1937, Trans. R. ent. Soc. Lond. 86: 67, fig. (3).

Cupidesthes ysobelae Jackson, 1965, l.c.: 530, fig.

# Genus ANTHENE Doubleday

Anthene Doubleday, 1847, List Lep. Ins. B.M. 2:27; Pinhey, 1949: 108; Swanepoel, 1953:116. Type-species: Papilio larydas Cramer, designated by Hemming (1935, Trans. R. ent. Soc. Lond. 1935: 435).

Lycaenesthes Moore, 1866, Proc. zool. Soc. Lond. 1865: 773 (partim); Aurivillius, 1898: 345, 353; 1924: 435–456; Bethune Baker, 1910: 14, 64; Murray, 1935: 131, 138. Pseudoliptena (partim); Stempffer, 1946 (see Liptena synonymy).

Eyes clothed in short but dense hair; palpi long, ascending, extending far beyond the frons, second segment laterally compressed, third segment long, slender, acuminate; antennae slender, more than half the length of the costa, with a well differentiated fusiform club; thorax and abdomen robust;  $\eth$  fore leg, femur velvety, tibia shorter than the femur, tarsus unsegmented, finely spinose below; mid and hind legs, femora velvety, tibiae shorter than the femora, tibiae with two spurs.

Wing shape. Fore wing with costa evenly curved, apex pointed, outer margin slightly convex; hind wing oval, apex very slightly angled, outer margin evenly rounded from vein 6 to 1b, three tufts of short hairs at the ends of vein 3, 2 and 1b respectively.

Wing venation (Text-fig. 318). Fore wing with 11 veins, 10 and 11 free from upper edge of cell.

Male genitalia (Text-fig. 167). Uncus composed of two small lobes closely fused to the posterior edge of the tegumen on either side of the median depression; subunci rather short, robust, bent almost at their base and tapering gradually to the apex, tegumen rather large, hood-shaped, vinculum broad with an inconspicuous saccus; lower fultura (furca) with two wide lanceolate arms; valves oblong with rounded apices, bearing, at about three-fifths of their length, a long digitate sclerite which ends level with the apex of the valve; penis robust, very wide in its internal portion, abruptly narrowed in its external portion and ending in a point, uncus clothed with long thick hair, valves almost bare except just at the apices.

The genus Anthene is very numerous in species, and it is one of those of which the male genitalia have been much studied. In his revision of the African Lycaenesthes, Bethune Baker (1910) figured and described those of nearly all the species known at that time. Unfortunately not all his figures show the same aspect, which renders comparison often rather difficult. Since 1936 I have myself published in a variety of papers both figures and descriptions of a number of species. There is no point in republishing them here, but references to them will be found in the list of species which follows. Comparison of these figures brings to light in nearly every case good specific characters, and also a perfect uniformity of type. The median indentation of the posterior margin of the tegumen varies in abruptness and depth, the curvature of the subunci and their thickness vary, and the outline of the valves is more or less incised, but the structural plan remains constant in all the species. Anthene is, in fact, a very homogeneous genus.

The early stages of several species have been described by various authors:—
A. liodes, A. lacheres and A. sylvanus by Lamborn, 1913, Trans. ent. Soc. Lond.

1913: 476.

- A. rubricinctus by Farquharson, 1921: 381.
- A. amarah and A. definita by Murray, 1935: 132 and by Pinhey, 1949: 109.
- A. ligures, A. definita, A. otacilia kikuyu, A. pitmani, A. lunulata, A. princeps ugandae, A. livida, A. levis grisea, A. amarah, A. larydas and A. crawshayi by Jackson, 1937, Trans. R. ent. Soc. Lond. 86: 221, and Pinhey, 1949: 110.
  - A. definita by Dickson, 1944, J. ent. Soc. sthn Afr. 7:97.

The larvae of several species live in company with ants of the genera Camponotus, Crematogaster and Pheidole.

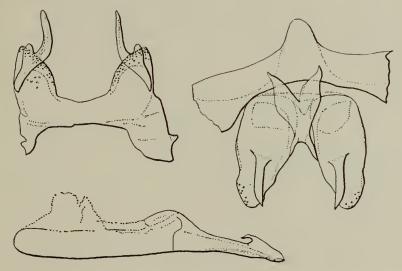


Fig. 167. Anthene larydas larydas (Cramer), & genitalia.

#### LIST OF SPECIES OF Anthene

As the genus *Anthene* has not been the subject of any comprehensive revision since 1910, it is probable the following list, which has been prepared with the help of suggestions by Mr. G. E. Tite, contains a certain amount of hidden synonymy.

Anthene abruptus (Gaede), 1915. Possibly a mere aberration of larydas Cramer. Anthene adherbal (Mabille), see liodes.

\*Anthene afra (Bethune Baker), 1910.

lysias (Hulstaert), 1924.

- \*Anthene alberta (Bethune Baker), 1910. Fig. genitalia, Stempffer, 1944, Revue fr. Ent. 10: 50.
- \*Anthene amarah amarah (Guerin), 1847. Fig. genitalia, Bethune Baker, 1910. olympusa (Walker), 1870.

Anthene amarah orphna Clench, 1965, Butt. Liberia: 369, figs.

Anthene arnoldi Jones, 1919, Proc. Trans. Rhod. scient. Ass. 16: 19.

Anthene aurea (Bethune Baker), 1910.

Anthene bakeri (H. H. Druce), 1910. ja (H. H. Druce MS).

\*Anthena barnesi Stevenson, 1940, J. ent. Soc. sth. Afr. 3: 106, fig. Anthene bihe (Bethune Baker), 1910, see liodes bihe.

\*Anthene bipuncta (Joicey & Talbot), 1921, Bull. Hill Mus. Witley 1:96, fig.

\*Anthene bitje (H. H. Druce), 1910. Fig. genitalia, Stempffer, 1944, Revue fr. Ent. 10:56.

Anthene buchholzi (Plötz), 1880.

\*Anthene butleri butleri (Oberthur), 1880.

\*Anthene butleri arabicus Gabriel, 1954, Exp. S.W. Arabia, 1937-38: 379.

\*Anthene butleri aureobrunnea (Ungemach), 1932, Mem. Soc. Sci. nat. phys. Maroc. 32: 85. Fig. genitalia, Stempffer, 1936, Revue fr. Ent. 3: 138.

\*Anthene butleri galla Stempffer, 1947, Bull. Soc. ent. Fr. 52: 38.

\*Anthene butleri livida (Trimen), 1881. Fig. Trimen, 1887. Fig. genitalia, Bethune Baker, 1910.

\*Anthene butleri stempfferi Storace, 1954, Annali Mus. civ. Stor. nat. Genova 66: 321.

Anthene chirinda (Bethune Baker), 1910.

\*Anthene contrastata contrastata (Ungemach), 1932, Mem. Soc. Sci. nat. phys. Maroc 32: 86. Fig. genitalia, Stempffer, 1936, Revue fr. Ent. 3: 137. otacilia mashuna (Stevenson), 1937.

\*Anthene contrastata turkana Stempffer, 1936, Revue fr. Ent. 3: 137.

\*Anthene crawshayi crawshayi (Butler), 1899. Fig. and fig. genitalia, Bethune Baker, 1910.

Anthene crawshayi Q-f. albilunulata (Ungemach), 1932, Mem. Soc. Sci. nat. phys. Maroc 3:88, fig.

\*Anthene crawshayi marginata (Hulstaert), 1924, Revue zool. afr. 12: 131.

Anthene crawshayi minuta (Bethune Baker), 1916, Trans. ent. Soc. Lond. 49: 277.

- \*Anthene crawshayi parallela (Aurivillius), 1924. Genitalia, Stempffer, 1945, Annls Soc. ent. Fr. 94:83.
- \*Anthene crawshayi sobrina (Talbot), 1935, Entomologist's mon. Mag. 71: 208, fig.

Anthene definita definita (Butler), 1899. Fig. Bethune Baker, 1910. nigrocaudata Pagenstecher, 1902.

\*Anthene definita liguroides (Strand), 1911. Fig. 1924, Aurivillius in Seitz. definita fasciata (Ungemach), 1932.

Anthene dewitzi (Staudinger), see juba.

\*Anthene discimacula (Joicey & Talbot,) 1921, Bull. Hill Mus. Witley 1:96 (3). Fig.  $\mathcal{Q}$  and genitalia, Stempffer, 1950, Revue fr. Ent. 17:142.

Anthene dulcis (Pagenstecher), see otacilia.

Anthene emolus (Trimen), see liodes.

Anthene erythropeocilus (Holland), 1893. Fig. Bethune Baker, 1910.

\*Anthene flavomaculatus (Smith & Kirby), 1893. Genitalia, Bethune Baker, 1910.

Anthene grosei (Aurivillius), see lunulata.

Anthene hewitsoni (Aurivillius), see lunulata.

- \*Anthene hobleyi hobleyi (Neave), 1904. Fig. genitalia, Bethune Baker, 1910.
- \*Anthene hobleyi elgonensis (Aurivillius), 1925, Ent. Tidsk. 46: 210. Genitalia, Stempffer, 1945, Annls Soc. ent. Fr. 94: 83.
- \*Anthene hobleyi kigezi Stempffer, 1961:63.
- \*Anthene hobleyi teita Stempffer, 1961:63.
- \*Anthene hodsoni hodsoni (Talbot), 1935, Entomologist's mon. Mag. 71: 125, 126, 207, fig.
- \*Anthene hodsoni usamba (Talbot), 1937, Trans. R. ent. Soc. Lond. 86: 68, fig.
- \*Anthene indefinita (Bethune Baker), 1910, fig. et genitalia. indefinita bigamica (Strand), 1911.
- \*Anthene indefinita f. oculata, Stempffer, 1946, Revue fr. Ent. 13: 15, fig.
- \*Anthene ituria (Bethune Baker), 1910 (3). \$\overline{\cappa}\$, Talbot, 1935, Entomologist's mon. Mag. 71: 207, fig.

Anthene ja (H. H. Druce) (MS), see bakeri.

\*Anthene juba (Fabricius), 1787. Fig. 1924, Aurivillius in Seitz. Genitalia, Bethune Baker, 1910.

dewitzi (Staudinger), 1891.

- \*Anthene kampala (Bethune Baker), 1910 (3).  $\bigcirc$ , Stempffer, 1947, Bull. Soc. ent. Fr. 52: 40. Genitalia, Stempffer, 1944, Revue fr. Ent. 11: 56.
- \*Anthene lachares (achares (Hewitson), 1874 (\$\varphi\$). \$\displays (as pulcher) Smith & Kirby, 1893. Genitalia, Bethune Baker, 1910.

liparis (Smith), 1898.

\*Anthene lachares obsolescens (Bethune Baker), 1910.

- \*Anthene lachares toroensis Stempffer, 1947, Bull. Soc. ent. Fr. 52: 40.
- \*Anthene larydas larydas (Cramer), 1780. Genitalia, Bethune Baker, 1910. pericles (Fabricius), 1793.

Anthene larydas kersteini (Gerstaecker), 1871.

Anthene larydas var. (Godart), see sylvanus.

- \*Anthene lasti (Smith & Kirby), 1894. Genitalia, Bethune Baker, 1910.
- \*Anthene lemnos lemnos (Hewitson), 1878. Genitalia, Bethune Baker, 1910. Anthene lemnos loa (Strand), 1911.

Anthene leptala (Strand), 1914.

- \*Anthene leptines (eptines (Hewitson), 1874. Fig. Hewitson, 1878. Genitalia Bethune Baker, 1910.
- \*Anthene leptines arnoldi (Aurivillius), 1923, Ergebn. 2te D. Zent. Afr. Exp. 1910–11, 1: 1236. Homonym of arnoldi Jones, 1919.
- \*Anthene levis levis (Hewitson), 1878. Genitalia, Bethune Baker, 1910.
- \*Anthene levis grisea (Talbot), 1935, Entomologist's mon. Mag. 71: 208, fig.
- \*Anthene ligures (Hewitson), 1874. Fig. Hewitson, 1878. Genitalia, Stempffer, 1953, Annls Mus. R. Congo belge 29: 32.

ligures amanica (Strand), 1909; ukerewensis pauperula Strand, 1909.

- \*Anthene liodes liodes (Hewitson), 1874. Fig. Bethune Baker, 1910.
  emolus (Trimen), 1866; sichela (Hewitson), 1878; adherbal (Mabille), 1877;
  monteironis (Kirby), 1878.
- \*Anthene liodes bihe (Bethune Baker), 1910 (3). Q Stevenson, 1937, Occ. Pap. natn. Mus. sth. Rhod. No. 6: 33.

Anthene liparis (Smith), see lachares.

Anthene lithas Q (Smith & Kirby), see lunulata.

Anthene lochias (Hewitson), see princeps smithi.

Anthene locra (Plötz), see sylanus.

\*Anthene locuples (Smith) 1898. Fig. Smith & Kirby, 1901; fig., genitalia, Stempffer, 1944, Revue fr. Ent. 11:56.

Anthene lukokesha (Karsch), see xanthopoecilus.

\*Anthene lunulata lunulata Trimen, 1894. Genitalia, Bethune Baker, 1910. otacilia (Hewitson), 1878; hewitsoni (Aurivillius), 1898. grosei (Aurivillius), 1898; lithas Q (Smith & Kirby), 1894.

Anthene lunulata f. magna (Hulstaert), 1924, Revue zool. afr. 12: 131.

Anthene lunulata aquilonis (Hulstaert), 1924, l.c.: 130.

\*Anthene lychnaptes (Holland), 1891. Fig. 1924, Aurivillius in Seitz. Genitalia, Stempffer, 1944, Revue fr. Ent. 11: 59.

lychnoptera (Smith & Kirby), 1893.

Anthene lychnides (Hewitson), 1878.

Anthene lychnoptera (Smith & Kirby), see lychnaptes.

Anthene lycotas (Smith), see xanthopoecilus.

Anthene lysias (Hulstaert), see afra.

\*Anthene lysicles (Hewitson), 1874. Fig. Hewitson, 1878. Genitalia, Bethune Baker, 1910.

Anthene madibirensis (Wichgraf), 1921, Int. ent. Z. 14: 179.

\*Anthene mahota (Smith), 1887. Fig. Smith & Kirby, 1893. Genitalia, Stempffer, 1944, Revue fr. Ent. 11:58.

\*Anthene makala (Bethune Baker), 1910. Genitalia, Bethune Baker, 1910.

Anthene melambrotus (Holland), 1893. Fig. Bethune Baker, 1910.

\*Anthene millari (Trimen), 1893. Genitalia, Bethune Baker, 1910.

\*Anthene minima (Trimen), 1893. Genitalia, Stempffer, 1936, Revue fr. Ent. 3:141.

Anthene moncus (Fabricius), see sylvanus.

Anthene monteironis (Kirby), see liodes.

Anthene musagetes musagetes (Holland), see rubricinctus.

Anthene musagetes elgonensis Stempffer, see rubricinctus jeanneli.

Anthene neglecta (Trimen), see princeps.

\*Anthene ngoko Stempffer, 1962: 1174; fig. and fig. genitalia.

Anthene nigrocaudata (Pagenstecher), see definita.

\*Anthene nigropunctata (Bethune Baker), 1910. Genitalia, Stempffer, 1944, Revue fr. Ent. 11:53.

Anthene ochreofascia (Talbot), 1935, Entomologist's mon. Mag. 71: 123, fig. Anthene olympusa (Walker), see amarah.

\*Anthene onias (Hulstaert), 1924, Revue zool. Afr. 12: 132. Genitalia, Stempffer, 1953, Annls Mus. R. Congo Belge 27: 37.

\*Anthene opalina opalina Stempffer, 1946, Revue fr. Ent. 13: 16, fig. and fig. genitalia.

\*Anthene opalina janna Gabriel, 1949, Proc. R. ent. Soc. Lond. (B) 18: 214, fig.

\*Anthene otacilia otacilia (Trimen) 1868. Fig. Trimen, 1887. Genitalia, Bethune Baker, 1910.

dulcis (Pagenstecher) 1902; tongidensis (Bethune Baker), 1926.

\*Anthene otacilia benadirensis Stempffer, 1947, Bull. Soc. ent. Fr. 52: 39.

\*Anthene otacilia kikuyu (Bethune Baker), 1910. Genitalia, Stempffer, 1944, Revue fr. Ent. 11:54.

Anthene otacilia (Hewitson) see lunulata.

Anthene otacilia mashuna (Stevenson), see contrastata.

Anthene pericles (Fabricius), see larydas.

\*Anthene pitmani pitmani Stempffer, 1936, Revue fr. Ent. 3: 139, genitalia. Fig. Stempffer, 1944, Revue fr. Ent. 11.

\*Anthene pitmani somalina Stempffer, 1936, Revue fr. Ent. 3: 140.

\*Anthene princeps princeps (Butler), 1876. Fig. and fig. genitalia, Bethune Baker, 1910.

neglecta (Trimen), 1891.

\*Anthene princeps smithi (Mabille), 1877. Fig. Mabille, 1887. Genitalia, Stempffer, 1936, Revue fr. Ent. 3: 142. lochias (Hewitson), 1878.

\*Anthene princeps smithi f. mabillei (Lathy), 1921, Ann. Mag. nat. Hist. (9) 8: 208. Fig. Lathy, 1926, Lepidoptera, 2. Genitalia, Stempffer, 1944, Revue fr. Ent. 11: 53.

\*Anthene princeps ugandae (Bethune Baker), 1910. Genitalia, Stempffer, 1944, Revue fr. Ent. 11: 52.

Anthene pulcher (Smith & Kirby), see lachares.

\*Anthene pyroptera (Aurivillius), 1895. Fig. Aurivillius, 1898. Genitalia, Stempffer, 1944, Revue fr. Ent. 11:58.

Anthene radiata (Bethune Baker), 1910. Genitalia, Bethune Baker, 1910.

\*Anthene rhodesiana Stempffer, 1962: 171, fig. and fig. genitalia.

Anthene rothschildi (Aurivillius), 1922, Voyage M. Rothschild en Ethiopie: 364.

- \*Anthene rubricinctus rubricinctus (Holland), 1891, (\$\varphi\$). Fig. Smith & Kirby, 1893. \$\delta\$ (as musagetes) Holland, 1893. Fig. Aurivillius, 1924, in Seitz. Genitalia, Bethune Baker, 1910.
- \*Anthene rubricinctus anadema (H. H. Druce), 1905. Genitalia, Stempffer, 1944, Revue fr. Ent. 11: 50.
- \*Anthene rubricinctus jeanneli Stempffer, 1961 : 64. musagetes elgonensis Stempffer, 1936 (invalid homonym).
- \*Anthene rubricinctus jeanneli f. latefasciata Stempffer, 1944, Revue fr. Ent. 11:50, fig.
- \*Anthene rubrimaculata (Strand), 1909. Genitalia, Stempffer, 1944, l.c.: 51.
- \*Anthene rufomarginata (Bethune Baker), 1910, genitalia.

Anthene ruwenzoricus (Grünberg), 1912.

Anthene saddacus (Talbot), 1935, Entomologist's mon. Mag. 71: 123, fig.

\*Anthene sanguinea (Bethune Baker), 1910, genitalia.

- \*Anthene scintillula (Holland), 1891. Fig. Smith & Kirby, 1893. Genitalia, Bethune Baker, 1910.
- \*Anthene sheppardi Stevenson, 1940, J. ent. Soc. sth. Afr. 3: 103, fig.
- \*Anthene schoutedeni (Hulstaert), 1924, Revue zool. afr. 12: 130. Genitalia, Stempffer, 1953, Annls Mus. R. Congo belge, 27: 33.

Anthene sichela (Hewitson), see liodes.

Anthene suquala (Pagenstecher), 1902.

Anthene syllidus (Hübner), see sylvanus.

- \*Anthene sylvanus sylvanus (Drury), 1773. Genitalia, Bethune Baker, 1910. moncus \( \partial \) (Fabricius), 1781; syllidus (Hübner), 1826; larydas var. (Godart), 1823; locra \( \partial \) (Plötz), 1890.
- \*Anthene sylvanus albicans (Grunberg), 1910. sylvanus bugalla Stempffer & Jackson, 1961.
- \*Anthene sylvanus niveus Stempffer, 1953, Annls Mus. R. Congo belge 27: 36.
- \*Anthene talboti Stempffer, 1936, Bull. Soc. ent. Fr. 41: 283 (3). Fig. Stempffer, 1944 Revue fr. Ent. 11; Genitalia, Stempffer, 1938, Mission Omo 4: 186. Q, Stempffer, 1951, Bull. Soc. ent. Fr. 56: 127.

Anthene tongidensis (Bethune Baker), see otacilia.

Anthene ukerewensis (Strand), 1909.

- \*Anthene versatilis (Bethune Baker), 1910.
- \*Anthene xanthopoecilus (Holland), 1893. Fig. Aurivillius in Seitz, 1924. lukokesha (Karsch), 1895; lycotas (Smith), 1898.

\*Anthene zenkeri (Karsch), 1895. Fig. Bethune Baker, 1910. Genitalia, Stempffer, 1944, Revue fr. Ent. 11: 60.

Anthene zenkeri ab. connexa (Aurivillius), 1923, Ergebn. 2te D. Zent. Afr. Exped., 1910–11, 1: 1237.

#### Genus NEURYPEXINA Bethune Baker

Neurypexina Bethune Baker, 1910, Trans. ent. Soc. Lond. 1910: 64. Type-species: Lycaenesthes lyzanius Hewitson, by original designation.

Lycaenesthes Moore, 1866, (partim); Aurivillius 1898: 351; 1924: 454.

Eyes densely but shortly pilose; palpi long, second segment laterally compressed, clothed below with long scales, third segment long, slender, acuminate; antennae as in Anthene; of fore leg, femur clothed below with long hair, tibia shorter than the femur, tarsus unsegmented, finely spinose below; mid and hind legs, femora clothed with long hair, tibiae shorter than the femora and bearing two strong spurs, metatarsi very long.

Wing shape. As in Anthene.

Wing venation (Text-fig. 319). Fore wing with 11 veins; 11 fused with 12 for some distance. Male genitalia (Text-fig. 168) (see also Bethune Baker, 1910, pl. 11, figs 33-34). Uncus composed of two small lobes closely fused to the tegumen on either side of the median indentation, subunci long, slender, bent at about one-third of the way from base, tegumen deeply notched on its posterior edge, vinculum fairly wide with spatulate saccus; lower fultura (furca) with slender arms; valves oblong, sub-oval, the upper process ending in a short point recurved close to the terminal edge, the lower process ending in a long sharp point; penis narrow at the base, dilated in the middle, then abruptly constricted to the apex; uncus densely pilose, valves almost bare.

The genitalia of *lyzanius* closely resemble those of *Anthene flavomaculatus* Smith & Kirby, although Bethune-Baker says "the genitalia are decidedly different" without mentioning in what way they differ. I can see no striking difference in his figures, (1910, pl. 9, fig. 25); however, the subunci of *lyzanius* seem to me to be more slender and longer than those of *A. flavomaculatus*.

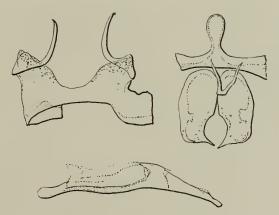


Fig. 168. Neurypexina lyzanius (Hewitson), & genitalia.

The male genitalia of *N. lamprocles* scarcely differ from those of *lyzanius* except in the shape of the tip of the valves. In *kalinzu* the subunci are short and robust and the saccus robust and triangular. In *quadricaudata* the subunci are long and slender as in *lyzanius*, but the saccus is disproportionately long, the valves digitate, almost filiform, and the penis extremely long and thin. *N. quadricaudata* was described as a *Triclema*, but the presence of vein 8 in the fore wing places it in *Neurypexina*, which is only to be distinguished from *Anthene* by the fore wing venation.

The larva of N. lyzanius has been described by Lamborn, 1913, Trans. ent. Soc. Lond. 1913: 484. It lives in company with the ant Pheidole rotundata.

# List of Species of Neurypexina

- \*Neurypexina kalinzu Stempffer, 1950, Revue fr. Ent. 16: 143, fig. genitalia.
- \*Neurypexina lamprocles (Hewitson), 1878. Fig. genitalia, Stempffer, 1944, Revue fr. Ent.: 60.
- \*Neurypexina lyzanius (Hewitson), 1874. Fig. Hewitson, 1878. Fig. genitalia, Bethune Baker, 1910.

regillus (Holland), 1891; turbatus (Smith & Kirby), 1893.

\*Neurypexina quadricaudata (Bethune Baker), 1926, Ann. Mag. nat. Hist. (9) 17: 400 (2). Stempffer, 1951, Bull. Soc. ent. Fr. 56: 69, fig. 3 genitalia.

Neurypexina regillus (Holland), see lyzanius.

Neurypexina turbatus (Smith & Kirby), see lyzanius.

#### Genus NEURELLIPES Bethune Baker

Neurellipes Bethune Baker, 1910, Trans. ent. Soc. Lond. 1910: 66. Type-species: Lycaenesthes lusones Hewitson, by original designation.

Lycaenesthes Moore, 1866 (partim); Aurivillius, 1898: 352; 1924: 455.

Monile Ungemach, 1932. Type-species: Monile pluricauda Ungemach (gemmifera Neave, 1910).

Eyes densely pilose, palpi extending well beyond the frons, second segment laterally compressed, clothed above with scales and below with stiff hair, third segment fairly long, acuminate; antennae like those of Anthene;  $\eth$  fore leg tibia shorter than femur, tarsus unsegmented, finely spinose below; mid and hind legs, femora clothed with long hair, tibiae shorter than the femora and bearing two robust spurs, metatarsi very long.

Wing shape. Like that of Anthene.

Wing venation (Text-fig. 320). Fore wing with 10 veins, veins 8 and 9 absent, 10 and 11 free. Male genitalia (Text-fig. 169) (see also Bethune Baker, l.c. pl. 13, figs 35–36). Uncus composed of two oval lobes narrowly fused to the tegumen on either side of the median indentation, subunci with very robust bases, tapered, and bent at about one-third, posterior margin of tegumen deeply excised, vinculum fairly wide, with a long saccus; lower fultura (furca) with a pyramidal base and long slender arms; valves wide with finely serrated distal edges, then deeply excised, the upper process ending in a sharp point, the lower process in a strong tooth, penis elongate, the upper surface of the internal portion widely open, the external portion evenly tapered and ending in a trumpet-shaped apex, uncus and apices of valve pilose.

The male genitalia of N. chryseostictus and N. fulvus are of the same type as those of N. lusones, that is to say their valves are quadrangular, and even expanded

distally. In N. likouala and N. aequatorialis the subunci are a little longer and more slender than in the preceding species, and the upper process of the valves is folded back towards the apex, against the lower process, both of them ending in a point. In N. staudingeri the valves take a very characteristic form, the two processes being separate almost from the base, the upper one being long, very curved and ending in a point, the lower one, equally long, being apically spatulate. The armature of N. gemmifera is remarkable in the size of the penis, which is enormous in relation to the other parts, shaped like a very elongate oval and encloses, towards the apex, a rather large cuneus.

### LIST OF SPECIES OF Neurellipes

- \*Neurellipes aequatorialis Stempffer, 1962: 1178, fig. and fig. genitalia.
- \*Neurellipes chryseostictus Bethune Baker, 1910, fig. genitalia.

Neurellipes fulvimacula (Mabille), see lusones.

- \*Neurellipes fulvus Stempffer, 1962: 1180, fig. and fig., genitalia.
- \*Neurellipes gemmifera (Neave), 1910. Fig. genitalia, Stempffer, 1944, Revue fr. Ent. 11:61.

pluricauda (Ungemach), 1932.

- \*Neurellipes likouala Stempffer, 1962: 1176, fig. and fig. genitalia.
- \*Neurellipes lusones (Hewitson), 1874. Fig. Hewitson, 1878. Fig. genitalia, Bethune Baker, 1910.

fulvimacula (Mabille), 1890.

Neurellipes meander (Plötz), 1880. Fig. Aurivillius in Seitz, 1924. Neurellipes pluricauda (Ungemach), see gemmifera.

- \*Neurellipes staudingeri staudingeri (Smith & Kirby), 1894. Fig. genitalia, Bethune Baker, 1910.
- \*Neurellipes staudingeri obsoleta Stempffer, 1947, Bull. Soc. ent. Fr. 52: 41.

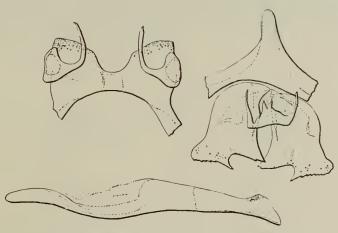


Fig. 169. Neurellipes lusones (Hewitson), & genitalia.

#### Genus TRICLEMA Karsch

Triclema Karsch, 1893, Berl. ent. Z. 38:227; Bethune Baker, 1910:70 Type-species: Lycaenesthes lucretilis Hewitson, by original designation.

Lycaenesthes Moore, 1866 (partim); Aurivillius, 1898: 345, 1924: 450.

Eyes finely pilose; palpi long, second segment laterally compressed, clothed below with long stiff black and white bristles, third segment fairly long, slender, acuminate; antennae slender, three-fifths the length of the costa, club fusiform; 3 fore leg, femur clothed with long white hair, tibia shorter than femur, tarsus unsegmented.

Wing shape. Like that of Anthene, the hind wing with a small pencil of hair at the ends of veins 1b, 2 and 3.

Wing venation (Text-fig. 321). Fore wing with 10 veins; 8 and 9 absent; 10 free from the

upper edge of the cell; 11 broadly confluent with 12.

Male genitalia (Text-fig. 170). Uncus composed of two small subtriangular lobes with rounded apices and fused to the lateral angles of the tegumen; subunci long with massive bases, bent in an obtuse angle near the base and with a terminal claw; tegumen subrectangular with a shallow rounded depression in the posterior margin; vinculum fairly wide, with a long tapering saccus; lower fultura two slender curved arms; valves elongate, the folded over upper edge forming a sharp point at about three-quarters of the total length and hooked at the apex, the lower edge also forming a sharp point in the distal half; penis elongate, slightly bent, swollen in its internal portion, then tapering in the external part and dilated just before the pointed apex; uncus and distal part of valves clothed with long fine hair.

The male genitalia of nigeriae, measseni, hades, phoenicis, kamilila ituriensis, lutzi and rufoplagata are very like those of lucretilus, the posterior margin of the tegumen being scarcely at all excavate on the median line, and the margins of both processes of the valves being provided with a robust tooth. In T. lamias the posterior margin of the tegumen is more deeply concave, and the valves are more produced with a recurved apex. The genitalia of obscura, inconspicua and lacidas are much the same, but the terminal margin of the tegumen is much more deeply concave, the apex of the valves more deeply incised and the two processes each end in a sharp stout tooth. I figure the genitalia of obscura (Text-fig. 171).

I have not been able to examine *Triclema fasciatus*, of which Aurivillius writes in his original description "Alis subtus albidis signaturis nigris fere ut in *L. staudingeri* formatis et ordinatis" and, again, "Die Art steht dem *L. staudingeri* ohne zweifel

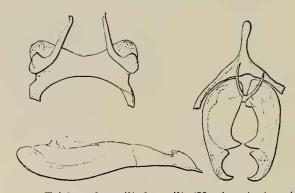


Fig. 170. Triclema lucretilis lucretilis (Hewitson), 3 genitalia.

ziemlich nahe". Bethune Baker, however, under the name fasciatus Aurivillius, figures firstly (1910, pl. 111, figs 15, 16) a male and female of a species of which the underside seems very different from that of staudingeri, and secondly (pl. x111, fig. 45) genitalia which, on the other hand, seem almost identical with those of staudingeri. It would appear that some confusion has crept into the explanations of these figures.

The caterpillar of *lucretilis* has been described by Lamborn (1913, *Trans. ent. Soc. Lond.* 1913: 485); that of *lamias* by Farquharson (1921, *Trans. ent. Soc. Lond.* 1921: 387); and that of *nigeriae* by Jackson, (1937, *Trans. R. ent. Soc. Lond.* 86: 227).

#### LIST OF SPECIES OF Triclema

Triclema africana Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 400.

Triclema caerulea (Aurivillius), 1895. Fig. Bethune Baker, 1910.

Triclema fasciatus (Aurivillius), 1895. Fig. et genitalia, Bethune Baker, 1910 (?).

subnitens (Bethune Baker), 1903.

- \*Triclema hades Bethune Baker, 1910. Fig. genitalia, Bethune Baker, 1910.
- \*Triclema inconspecua H. H. Druce, 1910.
- Triclema inferna Bethune Baker, 1926, Ann. Mag. nat. Hist. (9) 17: 399.
- \*Triclema ituriensis Joicey & Talbot, 1921, Bull. Hill Mus. Witley 1: 98, fig.
- \*Triclema kamilila Bethune Baker, 1910. Fig. genitalia, Bethune Baker, 1910.
- \*Triclema lacides (Hewitson), 1874. Fig. ♂, Hewitson, 1878; ♀ Bethune Baker, 1910.
- \*Triclema lamias (Hewitson), 1878, (3). Q and genitalia, Bethune Baker, 1910.

  Triclema lucretia (Smith & Kirby), see lucretilis.

lucretia (Smith & Kirby), 1894.

Triclema lucretilis albipicta Talbot, 1935, Entomologist's mon. Mag. 71: 127.

\*Triclema lutzi Holland, 1920, Bull. Am. Mus. nat. Hist. 43: 229. Fig. and fig. genitalia, Stempffer, 1953, Annls Mus. R. Congo belge 27: 39.

Triclema lydia Hulstaert, see nigeriae.

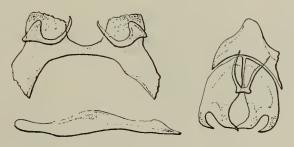


Fig. 171. Triclema obscura Druce, & genitalia.

\*Triclema maesseni Stempffer, 1957, Bull. Inst. Afr. noire 19: 223: fig. genitalia. Triclema marshalli (Bethune Baker), 1903. Fig. Bethune Baker, 1910.

\*Triclema nigeriae (Aurivillius), 1905. Fig. Aurivillius in Seitz, 1924. lydia Hulstaert, 1924. Fig. genitalia, Stempffer, 1953, Annls Mus. R. Congo belge 27: 38.

\*Triclema obscura H. H. Druce, 1910.

Triclema oculatus (Smith & Kirby), 1893.

\*Triclema phoenicis Karsch, 1893. Fig. and fig. genitalia, Bethune Baker, 1910.

\*Triclema rufoplagata Bethune Baker, 1910, and fig. genitalia, Bethune Baker, 1910.

Triclema subnitens (Bethune Baker), see fasciatus.

Triclema tisamenus (Holland), 1891. Fig. Bethune Baker, 1910.

#### Genus PHLYARIA Karsch

Phlyaria Karsch, 1895, Ent. Nachr. 21: 302. Type-species: Lycaena cyara Hewitson, by original designation.

Cupido Schrank (partim); Aurivillius, 1898; 358; 1924; 460.

Eyes shortly and densely pilose, palpi long, ascending, second segment long, clothed below with adpressed scales and some short stiff bristles, third segment short, very slender at its base, swollen in the middle and with a blunt apex; antennae slender, a little longer than half the length of the costa, with a well differentiated fusiform club; thorax clothed below with long white silky hair, legs long and slender;  $\delta$  fore leg with tibia shorter than the femur, tarsus unsegmented, finely spinose below.

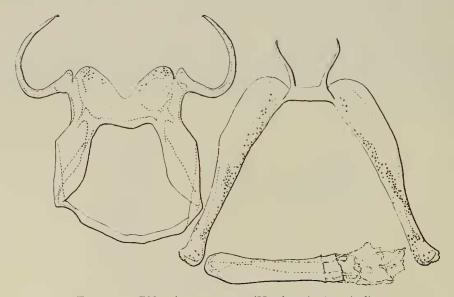


Fig. 172. Phlyaria cyara cyara (Hewitson), & genitalia.

Wing shape. Fore wing subtriangular, outer margin very slightly convex; hind wing suboval, a short filiform tail at the end of vein 2, anal angle well marked.

Wing venation (Text-fig. 322). Fore wing with 11 veins; 11 confluent with 12 for part of

its length.

Male genitalia (Text-fig. 172). Uncus reniform with rounded lateral angles; subunci long and slender, bent at two-fifths from base, apex unguiculate (in situ the subunci are folded back under the uncus); tegumen reduced; vinculum wide above, narrow below; lower fultura in the form of a furea with two slender curved arms; valves narrow, long, slightly swollen at the base, apex spatulate with strongly serrated edge; penis cylindrical, slightly swollen at its base; apex of uncus and lower edge of valves clothed with many fine hairs.

The genital armatures of *stactalla* and *cyara* are identical, for which reason I regard the former as a subspecies of the latter. The genitalia of *heritsia* are of the same type as those of *cyara*, but the valves are broader and shorter with long sharp apical teeth. The genitalia of *chibonotana* and *heritsia* again are identical, the former being a subspecies of the latter.

The early stages of cyara and heritsia have been described by T. H. E. Jackson (Trans. R. ent. Soc. Lond. 1937: 228).

# LIST OF SPECIES OF Phlyaria

\*Phlyaria cyara cyara (Hewitson), 1876.

\*Phlyaria cyara f. tenuimarginata (Grünberg,) 1908.

\*Phlyaria cyara stactalla Karsch, 1895. Fig. Aurivillius in Seitz, 1924.

\*Phlyaria heritsia heritsia (Hewitson), 1876 (♂). virgo (Butler), 1896 (♀).

\*Phlyaria heritsia chibonotana (Aurivillius), 1910. Fig. Gabriel, 1939, Ruwenzori Exp.

Phlyaria heritsia intermedia Tite, 1958, Entomologist 91: 115, fig. Phlyaria virgo (Butler), see heritsia.

#### Genus URANOTHAUMA Butler

Uranothauma Butler, 1895, Proc. zool. Soc. Lond.: 631; Pinhey, 1949:111; Swanepoel, 1953:76. Type-species: Uranothauma crawshayi Butler, by original designation. Cupido Schrank (partim); Aurivillius, 1898; 359; 1924:461.

Eyes densely and shortly pilose; palpi long, ascending, second segment very long, laterally compressed, clothed below with adpressed white scales and long stiff black bristles, above with black scales, third segment slender, cylindrical; antennae slender, a little more than half the length of the costa, club well differentiated, flattened;  $\delta$  fore leg, femur clothed with long hair, tibia slightly shorter than the femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing with costa slightly arched, apex slightly rounded, outer margin slightly convex and scalloped, hind wing with costa almost straight, outer margin slightly scalloped, a short triangular tail at the end of vein 2, a small indistinct lobe at the anal angle.

Wing venation (Text-fig. 323). Fore wing with 11 veins, 10 and 11 free from the upper edge of the cell, the latter being bent towards, and almost touching 12. Androconial scales are present in nearly all the species of *Uranothauma*, either grouped in a large velvety patch as in *crawshayi*, or in the form of long internervular rays as in *falkensteini*.

Male genitalia (Text-fig. 173). Uncus composed of two oval lobes fused to the tegumen and separated by the concave margin of the latter, subunci long and curved, gradually tapered; tegumen reduced; vinculum broad above, narrow below; lower fultura furca-like with two curved, tapering arms; valves oblong with spatulate and slightly serrate apices; penis oblong, slightly swollen at its base, vesica with a shagreened appearance; apices of uncus and valves pilose.

With the exception of *nubifer*, all the species of *Uranothauma* have male genitalia like those of *crawshayi*, only differing in the shape of the valves, which furnish good specific characters and enable one to separate *artemenes* on the one hand, and on the other, to associate *lunifer* with *poggei*. In *nubifer* the uncus lobes are smaller than in other species, the subunci shorter, the tegumen is larger and its distal margin less concave, and the upper part of the vinculum is expanded in the form of a triangle directed cephalad.

The early stages of *nubifer*, *delatorum* and *falkensteini* have been described by Jackson, (1937, *Trans. R. ent. Soc. Lond.* **86**: 229–230). The caterpillars live on species of Mimosacae, for *nubifer*, see also Pinhey, 1949: 111.

### LIST OF SPECIES OF Uranothauma

\*Uranothauma antinorii (Oberthür), 1883. Fig. genitalia, Stempffer, 1938, Mission Omo, 4: 190.

Uranothauma antinorii f. albicans Talbot, 1935, Entomologist's mon. Mag. 71: 147, fig.

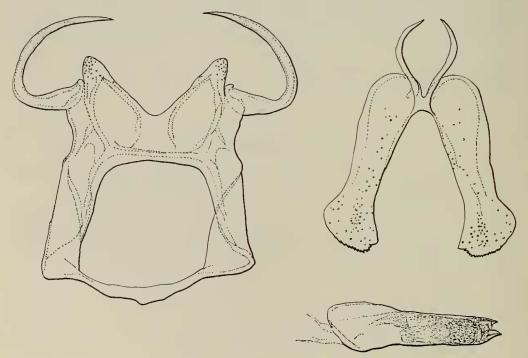


Fig. 173. Uranothauma crawshayi Butler, & genitalia.

Uranothauma antinorii f. felthami (Stevenson), 1934, Occ. Pap, natn Mus. Sth. Rhod. 16. Fig. Stevenson, 1940, J. ent. Soc. sth. Afr. 3.

Uranothauma antinorii f. magnificans Stoneham, 1937, Bull. Stoneham Mus.

Uranothauma antinorii f. Q splendens Stoneham, 1937, Bull. Stoneham Mus. **34** : 3.

\*Uranothauma artemenes (Mabille), Feb. 1880. Fig. Mabille 1885: 7. Fig. genitalia, Stempffer, 1938, Mission Omo 4: 193. auratus (Butler), April, 1880.

Uranothauma auratus (Butler), see artemenes.

- \*Uranothauma cordatus (E. Sharpe), 1891 (♂). Fig. \( \rightarrow \) and genitalia, Stempffer, 1938, Mission Omo 4: 188.
- \* Uranothauma crawshayi Butler, 1895. crawshayinus Aurivillius, in Seitz, 1925: 473.

Uranothauma cuneatum Tite, 1958, Entomologist 91: 117, fig. and fig. genitalia.

\*Uranothauma delatorum Heron, 1909. Fig. genitalia, Stempffer, 1938, Mission Omo 4: 191.

\*Uranothauma falkensteini (Dewitz), 1879. Fig. genitalia, Stempffer, 1938. l.c. : 191.

Uranthauma falkensteini f. albescens Stoneham, 1937, Bull. Stoneham Mus.

Uranothauma falkensteini Q-f. umbra Talbot, 1935, Entomologist's mon. Mag.

\*Uranothauma nubifer (Trimen), June 1895. Fig. genitalia, Stempffer, 1938, Mission Omo 4: 191.

pelotus (Karsch), October 1895.

Uranothauma nubifer f. distinctesignatus (Strand), 1911. Fig. Aurivillius in Seitz, 1925.

Uranothauma pelotus (Karsch), see nubifer.

\* Uranothauma poggei poggei (Dewitz), 1879. Fig. Aurivillius in Seitz, 1925.

\*Uranothauma poggei lunifer Rebel, 1914.

- \*Uranothauma vansomereni Stempffer, 1951, Bull. Soc. ent. Fr. 56: 128, fig.
- \*Uranothauma williamsi Carcasson, 1961, Occ. Pap. Coryndon meml Mus. 7: 21, fig. and fig. genitalia.

### Genus CACYREUS Butler

Cacyreus Butler, 1898, Proc. zool. Soc. Lond. 1897: 845; Pinhey, 1949: 111; Swanepoel, 1953: 67. Type-species: Papilio lingeus Cramer, by original designation. Hyreus Hübner, 1826, Verz. bek. Schmett.: 70 (invalid junior homonym).

Cupido Schrank (partim); Aurivillius, 1898; 360; 1924: 463.

Lycaena Fabricius (partim); Murray, 1935: 146.

Eyes densely and shortly pilose, palpi long, ascending, second segment laterally compressed, clothed below with scales and long stiff hair, third segment slender, ending in a sharp point, antennae slender, about half the length of the costa, club elongate and flattened; thorax rather

robust, clothed below with white hair, of fore leg, femur clothed below with long white hair, tibia shorter than the femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing subtriangular, apex angular, outer margin slightly convex; hind wing suboval, anal angle well marked, a short filiform tail at the end of vein 2.

Wing venation (Text-fig. 324). Fore wing with 11 veins; 10 and 11 free from the upper edge of the cell; 12 is slightly curved towards 11, but not touching it.

Male genitalia (Text-fig. 174). Uncus composed of two small subtriangular lobes with rounded apices fused to the lateral angles of the tegumen, each bearing a small projecting tubercle with rounded apex crowned with stiff hairs; subunci very robust with massive bases, curved, and ending in an open claw; tegumen large with a very convex posterior margin, giving the dorsal structures a trilobed appearance; vinculum broad above, narrow below; lower fultura furca-like with slender arms; valves broadly oval at the base; distally spatulate, the lower edge bearing sharp and irregular teeth which vary somewhat in shape in different specimens; penis short, stumpy, vesica with fine cornuti; uncus and distal portions of the valves pilose.

In their external characters (colour and markings of the wings) darius, virilis and audeoudi so closely resemble lingeus that they have for a long time been considered varieties of it; their genitalia, however, show quite clearly that they are separate species.

In darius (Text-fig. 175) the dorsal structures resemble those of lingeus, but the two processes of the valve are widely separate at their apices and the upper one, which is much longer than the lower one, is broad and heel-shaped.

In 1938 (Mission Omo 4: 196) I figured the male genitalia of virilis and andeoudi. Examination of these figures will show that in these two species the valves resemble those of lingeus, but the dorsal structures are very different: no tubercles on the uncus lobes, subunci longer and less robust, and in particular the posterior margin of the tegumen, instead of being markedly convex, is concave and so reduced to a narrow band medially.

In ethiopicus and dicksoni (for reference to figures see list of species) the tegumen is similarly reduced to a narrow band in its median portion, but tubercles on the

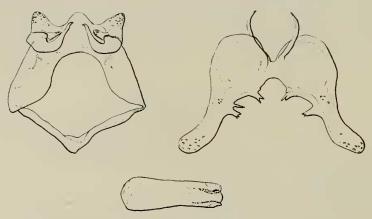


Fig. 174. Cacyreus lingeus lingeus (Cramer), & genitalia.

uncus are developed to form two lobes, which are longer in *dicksoni*, and the valves are elongate, slightly spatulate apically and smooth in outline.

In 1938 (l.c.: 199) I also figured the genitalia of *palaemon* and *marshalli*. In these the terminal margin of the tegumen is deeply concave, the uncus tubercles have become long digitate sclerites, the valves are oblong, massive in *marshalli* but deeply cleft apically in *palaemon*, the lower process much longer than the upper one.

The early stages of lingeus, palaemon and marshalli have been described by Murray (1935: 148), by Jackson (1937, Trans. R. ent. Soc. Lond. 86: 230) and by Dickson (1944, J. ent. Soc. sth. Afr. 7: 96; 1945, 8: 150; and 1947, 10: 127). The caterpillar of lingeus feeds on various species of Labiatae; those of palemon and marshalli on Geraniaceae. According to Jackson the colour of the caterpillar of lingeus varies considerably and always matches the colour of the species of Labiatae on which it feeds.

## LIST OF SPECIES OF Cacyreus

- \*Cacyreus audeoudi Stempffer, 1936, Bull. Soc. ent. Fr. 41: 284. Genitalia, Stempffer, 1938, Mission Omo 4: 197.
- \*Cacyreus darius (Mabille), 1877. Fig. Mabille, 1885: 7.
- \*Cacyreus dicksoni Pennington, 1962, J. ent. Soc. sth. Afr. 25: 283, fig. and fig. genitalia.
- Cacyreus ericus (Fabricius), see lingeus.
- Cacyreus ethiopicus Tite, 1961, Entomologist 94: 112, fig. and fig. genitalia.
- \*Cacyreus lingeus lingeus (Cramer), 1781. ericus (Fabricius), 1793.
- \*Cacyreus lingeus ciliaris (Aurivillius), 1910.
- \*Cacyreus marshalli Butler, 1898. Genitalia, Stempffer, 1938, Mission Omo 4: 200.

lingeus ♀ (Wallengren), 1857.

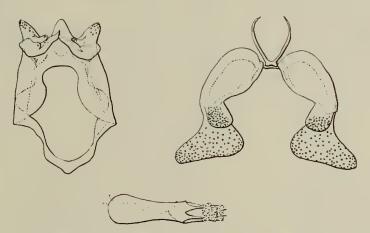


Fig. 175. Cacyreus darius (Mabille), 3 genitalia.

\*Cacyreus palemon palemon (Cramer), 1782. Fig. genitalia, Stempffer, 1950, Revue fr. Ent. : 145.

tespis (Herbst).

Cacyreus palemon f. fracta (Grünberg), 1912.

\*Cacyreus palemon f. ecaudata Stempffer, 1950, Revue fr. Ent. 17: 145.

\*Cacyreus palemon ghimirra Talbot, 1935, Entomologist's mon. Mag. 71: 148. Fig. genitalia, Stempffer, 1938, Mission Omo 4: 198; 1950, Revue fr. Ent. 17: 145.

Cacyreus tespis (Herbst), see palemon.

\*Cacyreus virilis (Aurivillius), 1924. Genitalia, Stempffer, 1938, Mission Omo 4:196.

#### Genus CASTALIUS Hubner

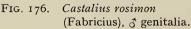
Castalius Hübner, 1819, Verz. bek. Schmett (5): 70; Pinhey, 1949: 112; Swanepoel, 1953: 62. Type-species: Papilio rosimon Fabricius (Indo-Malayan), designated by Scudder, 1875, Proc. Am. Acad. Arts Sci. 3: 135.

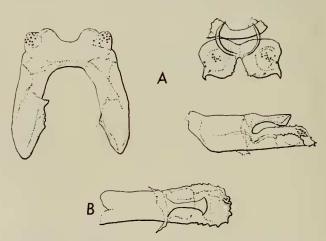
Cupido Schrank (partim); Aurivillius, 1898: 363, 1924: 466.

Lycaena Fabricius (partim); Murray, 1935: 155.

Eyes glabrous; palpi long, ascending, slightly divergent; second segment long, laterally, compressed, clothed below with long white adpressed scales; third segment long, slender acuminate; antennae slender, a little longer than half the length of the costa, club elongate, flattened; thorax clothed below with white hair; & fore leg, femur clothed with white hair, tibia shorter than femur, tarsus unsegmented, finely spinose below.







Castalius hintza hintza (Trimen), 3 genitalia. Fig. 177. A, Bulawayo; B, Kaikai Hills, N. of Ngami.

Wing shape. Fore wing subtriangular, hind wing oval, a thread-like tail about 3 mm. long, at the end of vein 2.

Wing venation (Text-fig. 325). Fore wing with 11 veins; 11 bent and touching 12 at one point.

Male genitalia (Text-fig. 176). Uncus composed of two oblong lobes with rounded apices, subunci long, slightly curved, with blunt apices, folded under the uncus; tegumen very much reduced, dorsally on the whole somewhat resembling species of holarctic Lycaeninae, vinculum very wide; lower fultura curtain-shaped and fused to the penis, which cannot be extracted without tearing open the apex of the fultura; valves subtriangular, the two processes slightly separated at their lightly serrated apices, near the base on the inner surface there is a rounded, semimembranous process; penis elongate, cylindrical in its internal portion, gradually tapering in its external portion to a sharply pointed apex, uncus and apices of valves pilose.

The male genitalia of the ethiopian species generally placed in the genus *Castalius* are entirely different from those of *rosimon*, the type-species; and they also display such a lack of uniformity that it is desirable to deal with each one individually.

C. hintza. Male genitalia (Text-fig. 177): uncus composed of two small semicircular lobes fused to the lateral angles of the tegumen; no subunci; tegumen very large with a rounded depression in its terminal margin; vinculum very wide above; lower fultura of two curved arms; valves much reduced, disc-shaped, with a small hook at the apex; penis highly specialized, short, massive, the external portion divided into two processes, one short, spatulate, the other long, gutter-shaped, with slightly serrated edges and enclosing a sheaf of imbricated spines; uncus and middle of valves pilose. The wing venation also differs slightly from that of rosimon inasmuch as in the fore wing vein 11 comes very near 12 but does not touch it.

C. calice. Male genitalia (Text-fig. 178): uncus composed of two small oval lobes fused to the lateral angles of the tegumen; subunci short, massive, only slightly curved, with a widened, strongly dentate apex; tegumen rather large, its posterior margin only slightly concave, lower fultura in the form of a small lamella with slightly spatulate apex; valves very large, the upper process folded under the lower one and ending in a finely spinose rounded apex, lower process connected to the vinculum by a membrane, its lower edge bearing teeth which are irregular in shape and size, its apex widely falcate; penis minute, slightly curved, gradually tapering; uncus pilose, valves with longer hair.

C. gregorii. Male genitalia differ from those of calice only by the heavier upper process of the valves. Probably these two species are only two races of the same species.

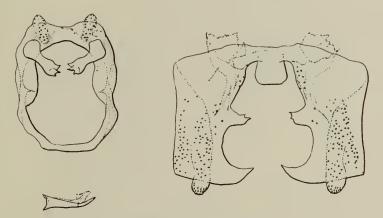


Fig. 178. Castalius calice calice (Hopffer), & genitalia.

C. cretosus. Male genitalia (Text-fig. 179): resembling those of calice but the valves are not so large, the upper process, which is folded under the lower one, ending in a long sharp point clothed with strong spines, the lower process, also connected to the vinculum by a membrane, armed with a tooth in the middle of its lower edge.

C. kaffana. Male genitalia (Text-fig. 180): closely resembling those of cretosus except that the terminal point of the upper process of the valves is shorter and more robust, and the apex of the lower process differs slightly in shape.

C. melaena. Male genitalia (Text-fig. 181): uncus composed of two rounded lobes fused on either side of the median concavity of the tegumen; subunci long, bent close to their massive base and gradually tapering to the apex; tegumen large, the dorsal part of the vinculum prolonged towards the eighth abdominal segment by a rounded expansion, ventral part narrow;

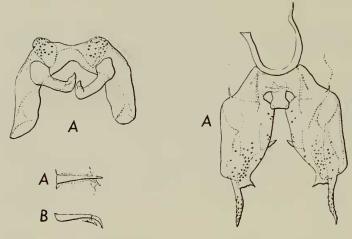


Fig. 179. Castalius cretosus cretosus Butler, & genitalia. A, Sudan-Abyssinia border;
B, Elgon, 3500 m., penis, lateral.

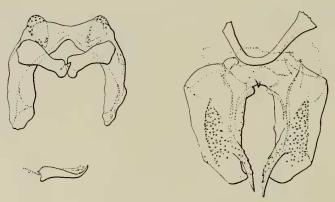


Fig. 180. Castalius kaffana Talbot, & genitalia.

lower fultura well developed, in the shape of a St. Andrew's cross whose lower arms are fused to the valves; valves oblong, widely connected to the vinculum by a membrane, the upper process folded under the lower one, short, curved and ending in a sharp point, the lower process having strongly serrated edges; penis minute, uncus and middle of valves pilose.

C. griqua. Male genitalia identical with those of melaena, with which it is probably conspecific.

C. carana. Male genitalia (Text-fig. 182): uncus composed of two small flattened lobes fused to the tegumen; subunci long, bent near the base, tapering to an apex which bears an open claw; tegumen and vinculum large; no lower fultura; valves oblong, the upper process folded under the lower one and both ending in a sharp point; penis of normal size, elongate, external portion short, slender, vesica with numerous cornuti, uncus and distal portions of valvae pilose.

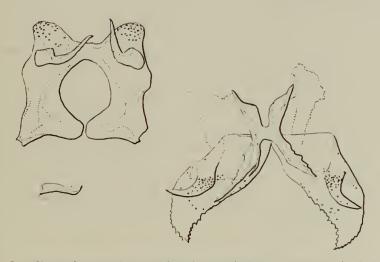


Fig. 181. Castalius melaena melaena (Trimen), & genitalia, tegumen, penis (Delagoa Bay), valves (Rhodesia-Congo border).

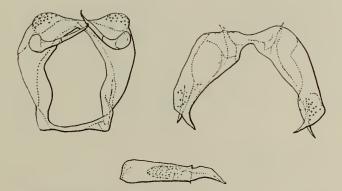


Fig. 182. Castalius carana carana (Hewitson), & genitalia.

C. ertli. Male genitalia (Text-fig. 183): uncus composed of two small flattened lobes fused to the tegumen; subunci very long with massive bases, only slightly curved, tapering gradually to their apices; tegumen strap-shaped; vinculum wide above, narrow below; lower fultura consists of a small spatulate lamella; valves oblong, upper process folded under the lower one and ending in a slender point with serrated edges, lower process widely connected to the vinculum by a membrane and having a rounded apex; penis of normal size and resembling that of carana; uncus pilose, hairs on valves more numerous and longer than on the uncus.

C. margaritaceus. Male genitalia (Text-fig. 184): uncus composed of two small oval lobes fused to the tegumen; subunci long, almost straight, with massive bases and tapering gradually to their apices; terminal margin of tegumen smoothly concave; vinculum wide above, narrow below; lower fultura consists of a small triangular lamella; valves very large, the upper process, which is folded under the lower one, ending in a long sharp spinose point, the lower process strongly dentate, the shape of the teeth varying in individual specimens; penis as in carana; uncus and valves pilose.

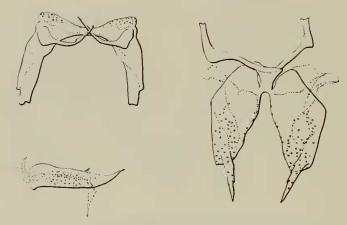


Fig. 183. Castalius ertli Aurivillius, & genitalia.

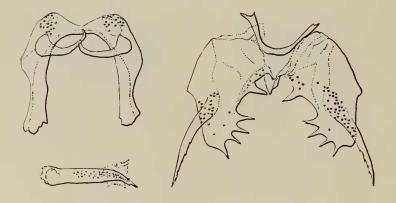


Fig. 184. Castalius margaritaceus Sharpe, & genitalia.

Comparison of the genitalia of the above species with those of *rosimon*, can only lead to the conclusion that the Ethiopian species of *Castalius* have no true relationship with the type-species *rosimon*.

Besides rosimon, I have examined only four Indo-Malayan species, viz: decidia Hewitson, ethion Doubleday, roxus Godart and elna Hewitson. Their male genitalia also differ considerably from those of rosimon and they show no resemblance to the so-called Castalius of Africa.

Fruhstorfer (Seitz, 9:883) has already pointed out the great diversity that is found in the male genitalia of species of this genus and yet he considered that the genus Castalius, as he understood it, was a valid systematic group. I cannot agree with him. It is evident that the species of Castalius auct. do not constitute a natural genus, but that they form a heterogeneous collection of groups of species whose convergent characters are of minor importance, e.g. the type of venation, which is commonplace, and the undeniable similarity of external appearance, with the white ground-colour of the upperside more or less tinged with blue, and with black markings, and especially a strong black streak parallel to the costa on the underside of the fore wing. I do not consider that this black streak has any particular systematic value, being only a recurrent character found also in species of Tarucus and in Azanus isis Drury, which have genitalia very distinctive from those of C. rosimon. In my opinion, the generic name Castalius should be restricted to rosimon and its Indo-Malayan congeners, if there are any. The Ethiopian species should be divided into four groups, probably with the status of genera:—r. hintza; 2. calice, gregorii, cretosus and kaffana; 3. melaena and griqua; 4. carana, ertli and margaritaceus. At present however, I refrain from naming these groups as genera, prefering to wait till the related Indo-Malayan species have received comprehensive treatment.

The early stages of hintza and cretosus have been described by Jackson (1937, Trans. R. ent. Soc. Lond. 86: 231-2): for hintza see also Pinhey (1949: 112).

#### LIST OF SPECIES OF Castalius

Castalius analogramma Bethune Baker, see cretosus nodieri.

- \*Castalius calice calice (Hopffer), 1855. Fig. Hopffer, 1862.
- \*Castalius calice gregorii Butler, 1894.

Castalius calice (Wallengren), nec Hopffer, see melaena.

\*Castalius carana carana (Hewitson), 1876.

Castalius carana kontu (Karsch), 1893. Fig. H. H. Druce, 1910.

\*Castalius cretosus cretosus Butler, 1876.

Castalius cretosus lactinatus Butler, 1886.

Castalius cretosus nodieri (Oberthur), 1883.

analogramma Bethune Baker, 1911.

- \*Castalius cretosus usemia Neave, 1904.
- \*Castalius ertli Aurivillius, 1907.
- \*Castalius hintza hintza (Trimen), 1864. Fig. Trimen, 1887. rosimon (Wallengren), 1857, nec Fabricius, 1775. Castalius hintza resplendens Butler, 1876.

\*Castalius kaffana Talbot, 1935, Entomologist's mon. Mag. 71: 149, fig.

\*Castalius margaritaceus E. Sharpe, 1892.

Castalius margaritaceus f. phasma Talbot, 1935, Entomologist's mon. Mag. 71: 148, fig.

\*Castalius melaena melaena (Trimen), 1887. Fig. Aurivillius in Seitz, 1924. calice (Wallengren), 1857; melama and melas Aurivillius ex errore Seitz: 467.

\*Castalius melaena griqua Trimen, 1887.

Castalius melaena interruptus Gabriel, 1954, Exp. S.W. Arabia, 1937/38: 381. Castalius melama (Aurivillius), see melaena, 1924: 467.

Castalius melas (Aurivillius), see melaena.

Castalius rosimon (Wallengren), see hintza.

#### Genus TARUCUS Moore

Tarucus Moore, 1881, Lep. Ceylon 1:81; Bethune Baker, 1918:269-296; Swanepoel, 1953:65. Type-species: Hesperia theophrastus Fabricius, by original designation.

Cupido Schrank (partim); Aurivillius, 1898: 363; 1924: 468.

Lycaena Fabricius (partim); Murray, 1935: 152.

Cacyreus Butler (partim); Pinhey, 1949: 119

Eyes glabrous, vertex clothed with white hair; palpi long, strongly ascending, second segment long, laterally compressed, clothed with white erect scales, third segment slender, cylindrical, clothed with black scales (see Bethune Baker, 1917, pl. 20, figs 33–34): antennae slender, about three-fifths the length of the costa, club elongate, well differentiated; thorax fairly robust, clothed below with long white hair;  $\eth$  fore leg, femur slightly flattened, tibia about as long as femur, tarsus unsegmented, black and white-annulated, spinose below.

Wing shape. Fore wing subtriangular, hind wing oval with a short filiform tail at the end

of vein 2.

Wing venation (Text-fig. 326): Fore wing with 11 veins, 11 curved, and touching 12 at one

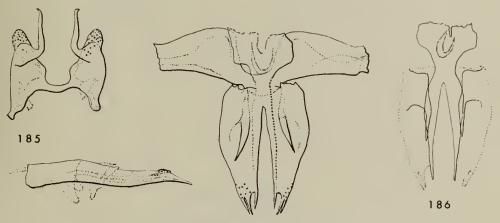
point in its length.

Male genitalia (Text-fig. 185). Uncus composed of two small lobes with rounded apices fused to the lateral angles of the tegumen; subunci long, bent at right-angles, the basal part massive, the free arm tapering gradually to the claw-shaped apex; tegumen rather large, the terminal margin deeply excised so as to leave only a narrow median band; vinculum very wide above; lower fultura peculiar in that the arms are enlarged to form wide curtains, which are fused to form a conical hood that passes above the penis, the apex of the cone directed towards the eighth abdominal segment (Bethune Baker gave the name "tectorius" to this structure); valves elongate with three sharp teeth of varying lengths at the apices, and bearing towards the base on the inner surface of the upper process a digitate, sharply pointed sclerite which runs parallel to the valve itself (the "virgae excitatae" of Bethune Baker); penis very elongate, sheathed by an anellus fused to the tectorius, tapering gradually in its external portion to the sharp pointed apex, vesica echinate; uncus and apices of valves pilose.

Bethune Baker (1917: 271) has interpreted the structures of the male genitalia of the species of *Tarucus* in a way which differs from my own interpretation. Instead of considering the "tectorius" as a modification of the lower fultura, he considers that it is a peculiar vestigial organ, part of a primitive structure which consisted of two groups, one surrounding the anus, the other protecting the sexual organs. On

the other hand he admits that the "virgae excitatae" take the place of the furca of other species of Lampidinae. I cannot agree with this interpretation. I have for a long time studied the geographical distribution of Tarucus in the Ethiopian region and dissected very numerous male genitalia of species of this genus. Not content with single whole mounts, in side view, I have dissected out the parts and cut through the top of the tectorius to extract the penis. I have come to the conclusion that the tectorius is not an independent structure, not a second vinculum, but simply a prolongation of the lower process of the valves, as there is no suture between the two structures. In other African Lampidinae one comes across a similar fusion of the distal portions of the arms of the lower fultura, or at least a curtain-like expansion of its arms. In my opinion the virgae excitatae do not correspond to the furca; they are simply extensions of the valves originating at different levels according to the species, and furthermore they are not found in all species of Tarucus. In theophrastus the virgae excitatae (Text-fig. 186) arise from the upper process, in species of the rosacea group (mediterraneae Bethune Baker) they are fused to the lower process. The valves of some Ethiopian species of Castalius (e.g. melaena Trimen) also have sclerites comparable to the virgae excitatae of Tarucus. To illustrate my contention I have drawn in the accompanying diagram (Text-fig. 186) with solid lines the lower process of the valves, the tectorius (cut open at its apex) and the virgae excitatae, and have indicated the general outline of the valves by dotted lines. I believe that my interpretation of these structures in the genitalia of Tarucus is more tenable than that of Bethune Baker.

In accordance with his genitalia studies, Bethune Baker divided the species of *Tarucus* into four groups and he figured the male genitalia of all the species known in 1918, except those of *thespis* Linnaeus. As I have since illustrated the genitalia of all the species that I have described, figures are here given only of those species not yet illustrated.



Figs 185-186. Tarucus theophrastus (Fabricius) & genitalia. 185, Kantara; 186, W. Darfur.

- Group I: virgae excitatae well developed, fused to the lower process of the valve; tectorius present.
  - T. rosacea Austaut; Bethune Baker, 1918: 282, pl. 16, fig. 7; Stempffer, 1943, Annls Soc. ent. Fr. 1942: 121; fig. 2.
  - T. balkanicus Freyer; Bethune Baker, 1918: 278, pl. 15, fig. 3; Stempffer, 1943, Annls Soc. ent. Fr. 1942: 121, fig. 3.
  - T. legrasi Stempffer, 1948, Revue fr. Ent. 15: 195, fig. 4.
  - T. kulala Evans (Text-fig. 187).

The armature of *kulala* resembles that of *rosacea* except that the subunci are rather less robust and the valves narrower with the apex not bent back. *T. kulala* is certainly a distinct species, easily separated by the pattern of the underside.

Group II: Virgae excitatae absent, tectorius poorly developed.

- T. grammicus Gr. Smith; Bethune Baker, 1918: 284, pl. 16, fig. 10.
- T. sybaris Hopffer; Bethune Baker, 1918: 285, pl. 16, fig. 9.
- T. quadratus Grant; Bethune Baker, 1918: 286, pl. 16, fig. 11.

Group III: Virgae excitatae fused to the upper process of the valve (theophrastus) or absent (ungemachi); tectorius well developed.

- T. theophrastus Fabricius; Bethune Baker, 1918: 287, pl. 16, fig. 12; Stempffer, 1943, Annls Soc. ent. Fr. 1942: 121, fig. 1.
- T. ungemachi Stempffer, 1943: 121, fig. 4.

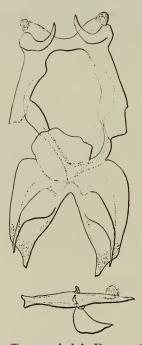


Fig. 187. Tarucus kulala Evans, & genitalia.

Group IV: virgae excitatae absent, tectorius present, uncus and subunci well developed.

T. bowkeri Trimen; Bethune Baker, 1918: 295, pl. 17, fig. 15.

T. thespis L.; Bethune Baker, 1918: 296.

This fourth group of species departs rather markedly from the others. The venation differs slightly in that vein 11 of the fore wing is anastomosed with vein 12 for some distance, whereas in the other species the two veins only touch at a point. The external appearance is also different, the pattern of the underside being heavier; in respect of the genitalia, in thespis (Text-fig. 188) the uncus consists of two narrow subtriangular lobes with rounded apices, the subunci are bent, very long and slender, with a terminal hook; the tegumen is reduced to a narrow band, vinculum moderately broad above, narrow below, tectorius well developed, valves fusiform, very elongate, broadly fused to the vinculum and apically rounded; penis elongate, very swollen at the level of the floor of the genital cavity, then suddenly narrowed and ending in a sharp point; vesica shagreened in appearance; lobes of the uncus and apices of the valves pilose.

The early stages of *T. rosacea* have been described by Chapman and Buxton (1919, *Entomologist's mon. Mag.* 55: 163–173), those of *T. sybaris* by Murray (1935, 153), Gowan C. Clark (1942, *J. ent. Soc. sth. Afr.* 5: 115–118), and Pinhey (1949, : 112); those of *T. thespis* by Dickson (1944, *J. ent. Soc. sth. Afr.* 7: 20–29).

The caterpillars of thespis feed on species of Rhamnaceae, those of rosacea and sybaris on species of Zizyphus. In Europe theophrastus has been found on Zizyphus vulgaris Lam.

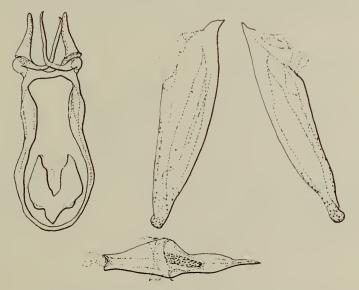


Fig. 188. Tarucus thespis (Linnaeus), & genitalia.

The genus *Tarucus* includes several species which it is almost impossible to differentiate by their external appearance, but their male genitalia are distinctive enough to leave no doubt as to the validity of the species. It would be interesting to examine systematically the genitalia of all the specimens of *Tarucus* which are to be found in various collections so as to decide the habitats of the known species and possibly discover new species.

## LIST OF SPECIES OF Tarucus

\*Tarucus balkanicus (Freyer), 1844.

- \*Tarucus bowkeri (Trimen), 1883. Fig. Trimen, 1906.
- \*Tarucus grammicus (Gr. Smith), 1893. louisae E. Sharpe, 1898.

\*Tarucus kulala Evans, 1955, Entomologist 88: 185.

\*Tarucus legrasi Stempffer, 1948, Revue fr. Ent. 15: 194, fig.

Tarucus louisae E. Sharpe, see grammicus.

Tarucus mediterraneae Bethune Baker, see rosacea.

Tarucus pitho (Linnaeus), see thespis.

\*Tarucus quadratus Grant, 1899. Fig. Bethune Baker, 1918.

- \*Tarucus rosacea (Austaut), 1885. Fig. Bethune Baker, 1918 (as mediterraneae). mediterraneae Bethune Baker, 1918.
- \*Tarucus sybaris sybaris (Hopffer), 1855. Fig. Hopffer, 1862.

\*Tarucus sybaris linearis (Aurivillius), 1924.

\*Tarucus theophrastus (Fabricius), 1793.

\*Tarucus thespis (Linnaeus), 1764. Fig. Trimen, 1887. pitho (2) (Linnaeus), 1764.

\*Tarucus ungemachi Stempffer, 1943, Annls Soc. ent. Fr. 1942: 121, fig.

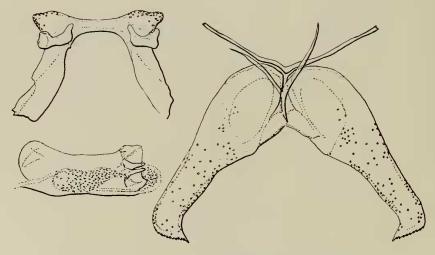


Fig. 189. Lampides boeticus (Linnaeus), & genitalia.

#### Genus LAMPIDES Hübner

Lampides Hübner, 1819, Verz. bek. Schmett (5) 70; Pinhey, 1949: 115; Swanepoel, 1953: 82.

Type-species: Papilio boeticus Linnaeus, by selection of Grote (1873, Bull. Buffalo Soc. nat. Sci. 3: 179).

Cupido Schrank (partim); Aurivillius, 1898; 367; 1924:471.

Lycaena Fabricius (partim); Murray, 1945: 161.

Eyes clothed with long dense hair; palpi long, ascending, second segment very long, laterally compressed, clothed below with close-set long white scales and with a row of long stiff black hairs, third segment short, horizontal, with blunt apex; antennae slender, white-annulated, a little longer than half the length of the costa, club elongate, clavate; thorax robust, clothed below with long white hair; 3 fore leg, femur clothed with very long white hair, tibia shorter than femur, tarsus unsegmented.

Wing shape. Fore wing triangular, outer margin slightly convex, hind wing oval, with a filiform tail at the end of vein 2.

Wing venation (Text-fig. 327). Fore wing with 11 veins.

Male genitalia (Text-fig. 189). Uncus composed of two subtriangular lobes with rounded apices fused to the lateral angles of the tegumen, subunci short, flattened into lamellae, slightly curved, apices blunt; tegumen reduced to a narrow median band, vinculum wide above, very narrow below, no saccus; lower fultura Y-shaped, with long slender arms; valves oblong with broad bases and serrated apices which are slightly recurved; penis short, massive, apical part with two short blunt-ended processes which are probably homologous with the carina of the penis of Syntarucus; vesica armed with numerous cornuti; uncus and distal halves of valves pilose.

The early stages of boeticus have often been described:—Murray (1935: 162); Jackson (1937, Trans. R. ent. Soc. Lond. 86: 234); Dickson, C. G. E., (1944, J. ent. Soc. sth. Afr. 7: 96; 1945, 8: 150; and 1947, 10: 127); Pinhey, (1949: 117). As in Europe the caterpillar is polyphagous and lives in the pods of many wild and cultivated species of Leguminosae.

## LIST OF SPECIES OF Lampides

\*Lampides boeticus (L.), 1767.

#### Genus CYCLYRIUS Bulter

Cyclyrius Butler, 1897, Proc. zool. Soc. Lond. 1896: 830. Type-species: Polyommatus webbianus Brulle (species restricted to the Canary Islands), by original designation. Cupido Schrank (partim); Aurivillius, 1924: 471.

Eyes densely clothed with fairly long hair; palpi long, ascending, second segment long, laterally compressed, clothed above and below with white scales and below also bearing long black stiff hairs, third segment rather short, slender, acuminate and clothed with black scales; antennae slender, about three-fifths the length of the costa, club elongate clavate, well differentiated; thorax fairly robust, clothed below with white hair; of fore leg, tibia shorter than the femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing subtriangular, apex angular, hind wing oval, no tail.

Wing venation (Text-fig. 328). Fore wing with 11 veins, 11 runs close to 12 but does not touch it.

Male genitalia (Text-fig. 190) (see also Stempffer, 1938, Mission Omo 4: 201, fig. 23): Uncus composed of two small lobes narrowly fused to the tegumen on either side of its median con-

cavity; subunci rather short, almost straight, tapering gradually from their base to the blunt apices; tegumen reduced to a rather narrow median strip; vinculum wide above, narrow below; lower fultura consisting of two slender curved arms arising from the base of the valves; valves oblong, proximal portion oval, apex serrated, the size of the teeth increasing gradually from the upper to the lower process, the lower edge deeply excised near the apex; penis of the same structure as in the genus Syntarucus, i.e. short, massive with carina penis and cuneus, the latter bifid; uncus and lower processes of valves pilose.

As can be seen by comparing figures, the male genitalia of *C. webbianus* are closely analogous to those of Ethiopian *Syntanucus* and American *Leptotes*.

In his generic description, Butler included in *Cyclyrius* the species aequatorialis Sharpe and juno Butler; Aurivillius (1924:471–473) also included in what he called the *Cyclyrius* group, tsomo Trimen, noquasa Trimen, kisaba Joicey & Talbot, notobia Trimen and crawshayi Butler. Later on other species or subspecies were placed in this genus by other authors, viz: boma Bethune Baker, ruandensis Joicey & Talbot, vulcanica Joicey & Talbot, marungensis Joicey & Talbot, wollastoni Bethune Baker and major Joicey & Talbot.

I have examined the male genitalia of all these species or subspecies. As I have shown (1943, Annls Soc. ent. Fr. 1942: 125) they have nothing in common with those of C. webbianus; crawshayi is a Euchrysops (Stempffer, 1938, Mission Omo 4: 204, fig. 26) and all the others belong to the genus Harpendyreus. They must be excluded from the genus Cyclyrius, which they resemble only fortuitously and in the markings on the underside of the hind wing.

On the other hand mandersi H. H. Druce, from Mauritius, described as a Nacaduba, is a true Cyclyrius not only in its venation but also in its male genitalia, of which I give a drawing (Text-fig. 191): it is true that the subunci are finely spinose, but the penis has the highly characteristic form of Cyclyrius, Syntarucus and Leptotes. It seems impossible to have any doubts as to the close relationship of these three genera

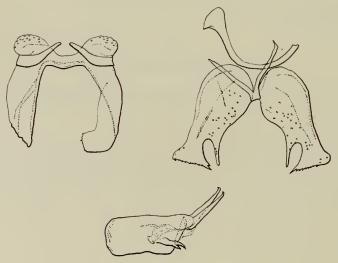


Fig. 190. Cyclyrius webbianus (Brullé), & genitalia.

and of their common ancestry. They are rare examples amongst Lycaenidae, of a stock common to Indo-Malaysia, Africa and to sonoran and neotropical America. This interesting fact had already been noted by Seitz, who (1937, Macrolep. World 5: 739) writes "By this, however, we cannot explain other symptoms, such as the occurrence of otherwise African genera on the high ridge of the Andes (genus Cyclyrius Butler)". It is interesting to note that whereas the underside markings of Leptotes cassius Cramer, L. theonus Lucas and L. marina Reakirt recall species of Syntarucus, those of Leptoters callanga Dyar and L. andicola Godman & Salvin are much more like those of C. webbianus.

The early stages of *C. mandersi* have been described by Manders (1908, *Trans. ent. Soc. Lond.* **1907**: 446).

# LIST OF SPECIES OF Cyclyrius

- \*Cyclyrius mandersi (H. H. Druce), 1907.
- \*Cyclyrius webbianus (Brulle), 1850 (palaearctic species : Canary Isles).

## Genus SYNTARUCUS Butler

Syntarucus Butler, 1900, Proc. zool. Soc. Lond.: 929; Pinhey, 1949: 115; Swanepoel, 1953: 8.

Type-species: Papilio pirithous L., 1767 (Papilio telicanus Lang, 1789), by monotypy.

Cupido Schrank (partim); Aurivillius, 1898: 364; 1924: 470.

Lycaena Fabricius (partim); Murray, 1935: 189.

Eyes densely pilose; vertex clothed with erect black hair; palpi long, ascending, second segment long, laterally compressed, clothed beneath with long stiff white and black hair, third segment slender, acuminate; antennae slender, a little longer than half the length of the costa,

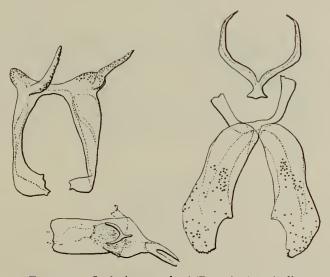


Fig. 191. Cyclyrius mandersi (Druce), & genitalia.

club oval, flattened, well differentiated, thorax moderately robust, clothed below with long white silky hair; of fore leg, tibia shorter than femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing subtriangular, apex angular, outer margin slightly convex; hind wing oval, with a short filiform tail at the end of vein 2, anal angle not well marked.

Wing venation (Text-fig. 329). Fore wing with 11 veins, 11 approximate to but not touching vein 12.

Male genitalia (Text-fig. 192). Uncus composed of two small oval lobes narrowly fused to the posterior margin of the tegumen; subunci long, slightly curved, tapering gradually to blunt apices, not unguiculate; tegumen reduced to a narrow median band of which the posterior margin is roundly concave; the rather wide upper part of the vinculum bent at right angles to the narrow lower portion; lower fultura composed of two slender curved arms fused together at their base and attached to the base of the valves; valves suboval ending in a long curved point which bears a small tooth on its inner edge; penis short, massive in its internal protion; vesica enclosing a large hook-shaped cornutus (cuneus of Stitz): the external portion of the penis ends in two long sharp points curved towards the ventral surface (carina penis of Petersen). During the act of copulation the pars inflabilis, accompanied by the cuneus, is unsheathed and passes above (not between) the points of the carina penis; uncus and distal portions of valves pilose.

In 1935 (Mission Omo 2: 219, 240) I published a revision of the African species of Syntarucus, in which I showed that Syntarucus plinius Fabricius, often recorded from Africa by various authors, was in fact an exclusively Indo-Malaysian species, but that, on the other hand there existed in Africa other very common species, which till then had been confused with pirithous, all of which had conspicuously different genitalia. Since then Tite (1958, Entomologist 19: 189) has isolated three other species.

The male genitalia of all species resemble those of *pirithous*, though the subunci and the valves supply excellent specific characters which allow one to determine the species rapidly and accurately, in contrast to their external appearances which are

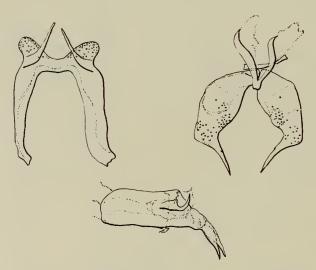


Fig. 192. Syntarucus pirithous (Linnaeus), & genitalia.

confusing. The penis of all has the same characteristic form and has the carina penis and cuneus typical of the genus. References to figures of genitalia are included in the list of species given below.

It is worth noting that, whereas most of the species of *Syntarucus* are widespread throughout Africa and often found flying together, five have very restricted distributions. These are *S. socotranus*, found only on Sokotra, *S. rabefaner*, restricted to Madagascar, *S. terrenus* in São Thomé and *mayottensis* and *casca* of the Comoro Islands. In the description of *Cyclyrius* I have already called attention to the close affinities existing between that genus and *Syntarucus* and the American genus *Leptotes*.

The early stages of *pirithous* have been described by Murray (1935: 190). In S. Africa the caterpillar feeds on various species of Leguminosae, e.g. *Plumbago capensis*, *Burkea africana*, *Indigofera* spp., *Mundulea sericea* and *Medicago sativa* L. (Leguminosae). Recently Mr. K. M. Pennington has informed me that at Delagoa Bay the caterpillar of *pulcher* feeds on *Sesbania sesban*.

# LIST OF SPECIES OF Syntarucus

- \*Syntarucus babaulti Stempffer, 1935, Mission Omo 4: 235, fig. and fig. genitalia.
- \*Syntarucus brevidentatus Tite, 1958, Entomologist 91:189, fig. and fig. genitalia.
- \*Syntarucus casca Tite, 1958, Entomologist 91: 190, fig. and fig. genitalia.
- \*Syntarucus jeanneli Stempffer, 1935, Mission Omo 4: 232, fig. and fig. genitalia.
- \*Syntarucus marginalis (Aurivillius), 1924. Fig. genitalia, Stempffer, 1935, Mission Omo 4.
- \*Syntarucus mayottensis Tite, 1958, Entomologist 91: 191, fig. and fig. genitalia.
- \*Syntarucus pirithous pirithous (Linnaeus), 1767. Fig. Gerhard, 1853. Fig. genitalia, Stempffer, 1935, Mission Omo 4. telicanus (Lang), 1789.

Syntarucus pirithous insulanus (Aurivillius), 1909.

- \*Syntarucus pulcher (Murray), 1874. Fig. genitalia, Stempffer, 1935, Mission Omo 4.
- \*Syntarucus rabefaner (Mabille), 1877. Fig. and fig. genitalia, Stempffer, 1935.
- \*Syntarucus socotranus (Grant), 1899. Fig. and fig. genitalia, Stempffer, 1935. Syntarucus telicanus (Lang), see pirithous.

Syntarucus terrenus Joicey & Talbot, 1926, Entomologist 59: 224.

#### Genus HARPENDYREUS Heron

Harpendyreus Heron, 1909, Trans. zool. Soc. Lond. 19: 158; Swanepoel, 1935: 72. Typespecies: Harpendyreus reginaldi Heron, by monotypy.

Cupido Schrank (partim); Aurivillius, 1898: 368; 1924: 462, 472.

Lycaena Fabricius (partim); Murray, 1935: 162, 185.

Eyes densely clothed with long hair; palpi long, ascending, second segment long, laterally compressed, clothed above with black scales, below with white scales and long stiff black hair,

third segment slender, acuminate, clothed with white scales on the inside and black scales on the outside; antennae slender, about three-fifths the length of the costa, club oval, flattened, well differentiated; thorax fairly robust, clothed below with long silky hair;  $\eth$  fore leg, femur clothed with long grey hair, tibia slightly shorter than the femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing subtriangular, apex angular margin convex, hind wing oval, no tail, anal angle rounded.

Wing venation (Text-fig. 330). Fore wing with 11 veins.

Male genitalia (Text-fig. 193). Uncus composed of two oval lobes fused to the posterior edge of the tegumen, each lobe bearing near its apex a small prominent tubercle crowned with long strong stiff bristles (N.B. as this tubercle is rather soft, it is easily displaced when the genitalia are mounted; sometimes it is pushed sideways as in the case in the figure, sometimes it is pushed upwards, projecting above the posterior margin of the tegumen so that it is difficult to prepare mounts that are strictly comparable in respect of all details); subunci long, slender, curving, tapering gradually from the base to the pointed apex which bears a wide-open claw; tegumen very large, prolonged towards the eighth abdominal segment by a wide rounded expansion; vinculum narrow, almost at right-angles to the tegumen when seen in side view, lower fultura composed of two long curved arms fused together at their bases and arising from the origin of the valves; valves oblong, their general shape reminding one of a human leg including calf, ankle and foot, the internal surface bearing near its base a small protuberance which possibly corresponds to a connection between the valves and the tergal elements of the genitalia; penis short, massive, vesica with numerous cornuti; uncus densely clothed with long fine hair, distal portion of the valves sparsely clothed with short hairs.

The male genitalia of all the species or subspecies of Harpendyreus examined are of the same type as those of H. reginaldi except those of H. notobia (see below). Their specific characters are found mainly in the shape of the valves, but these

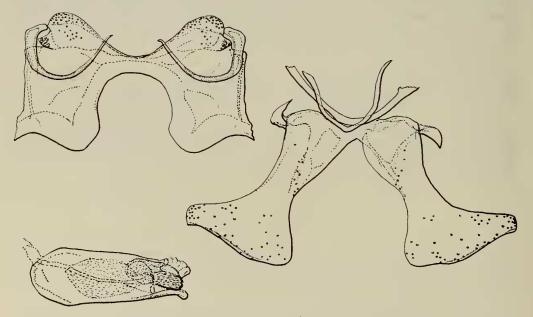


Fig. 193. Harpendyreus reginaldi Heron, 3 genitalia.

characters are sometimes so poorly defined that it is difficult to decide whether one is dealing with a distinct species, a geographical race or even an individual variation. The species can be grouped according to variation in the valves, but as in certain species I had only a few specimens for dissection, I cannot rest satisfied with such an arrangement. To illustrate what I mean I give a figure (Text-fig. 194) of the male genitalia of *major*, which had been described as a subspecies of *noquasa*, but which is certainly a valid species.

H. notobia Trimen (Text-fig. 195). In this species uncus, subunci, tegumen and penis are of the same type as in reginaldi, but the two arms of the lower fultura are strongly curved and their apices are connected by a membrane which forms a complete ring around the penis, and the valves are very long, digitate, without any distal widening.

Judging by their male genitalia the species of *Harpendyreus* form a homogeneous genus. The markings on the underside of the hind wing of *reginaldi* remind one of *Uranothauma antinorii* Oberthur. All the other species have a white transverse streak, more or less pronounced, like the one in *Cyclyrius webbianus* and it is this trivial unimportant character that led some authors to include them, rather arbitrarily, in the genus *Cyclyrius*.

In Eastern Africa *Harpendyreus* is distributed from Kivu, Ruanda and Ruwenzori to Natal, Transvaal, Basutoland and the Cape, but it is discontinuous, the various scattered species and subspecies being restricted to specific mountain masses, though clearly having a common origin.

The early stages of *notobia* and *noquasa* have been described by Murray (1935: 163, 186).

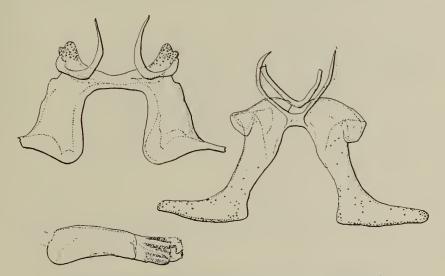


Fig. 194. Harpendyreus major (Joicey & Talbot), & genitalia.

# LIST OF SPECIES OF Harpendyreus

- \*Harpendyreus aequatorialis aequatorialis (Sharpe), 1892. Fig. genitalia, Stempffer, 1938, Mission Omo 4: 202.
- \*Harpendyreus aequatoralis sharpiae (Butler), 1900.
- \*Harpendyreus aequatoralis vulcanica (Joicey & Talbot), 1924, Bull. Hill Mus. Witley 1:547.
- \*Harpendyreus argenteostriata Stempffer, 1961: 65, fig. and fig. genitalia.

\*Harpendyreus juno juno (Butler), 1897.

- \*Harpendyreus juno boma (Bethune Baker), 1926, Ann. Mag. nat. Hist. (9) 17: 402.
- \*Harpendyreus major (Joicey & Talbot), 1924, Bull. Hill Mus. Witley 1:548.

\*Harpendyreus marlieri Stempffer, 1961: 64, fig. and fig. genitalia.

- \*Harpendyreus marungensis marungensis (Joicey & Talbot), 1924, Bull. Hill Mus. Witley 1: 547.
- \*Harpendyreus marungensis wollastoni (Bethune Baker), 1926, Ann. Mag. nat. Hist. (9) 17: 402. Fig. Gabriel, 1939, Ruwenzori Exp.
- \*Harpendyreus meruanus meruanus (Aurivillius), 1910.

\*Harpendyreus meruanus kisaba (Joicey & Talbot), 1921.

- \*Harpendyreus meraunus ruandensis (Joicey & Talbot), 1924, Bull. Hill Mus. Witley 1: 548.
- \*Harpendyreus notobia (Trimen), 1868. Fig. Trimen, 1887.

\*Harpendyreus reginaldi Heron, 1909. Fig. Gabriel, 1939.

\*Harpendyreus tsomo tsomo (Trimen), 1868. Fig. Trimen, 1887.

\*Harpendyreus tsomo noquasa (Trimen), 1887. Fig. Butler, 1898.

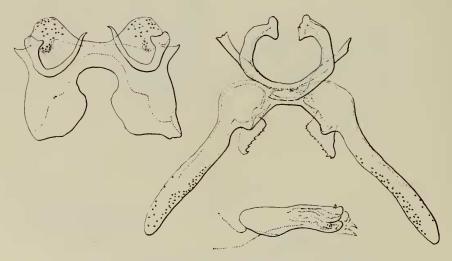


Fig. 195. Harpendyreus notobia (Trimen), & genitalia.

# Genus PSEUDONACADUBA Stempffer

Pseudonacaduba Stempffer, 1943, Ann. Soc. ent. Fr. 1942: 130; Pinhey, 1949: 186; Swanepoel, 1953: 77. Type-species: Lycaena aethiops Mabille, by original designation. Cupido Schrank (partim); Aurivillius 1898: 367; 1925: 471.

Eyes densely pilose; from black with two narrow white lateral lines; palpi long, ascending, second segment long, laterally compressed, clothed below with long erect bristles, third segment fairly long, very slender, ending in a pointed apex; antennae slender, about three-fifths the length of the costa; club flattened, well differentiated; of fore leg, femur clothed with long grey hair, tibia shorter than the femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing subtriangular, apex angular, outer margin convex; hind wing oval, no tail, anal angle well marked.

Wing venation (Text-fig. 331). Fore wing with 11 veins; 11 bent and confluent with 12 for part of its length.

Male genitalia (Text-fig. 196). Uncus crescentic with sharp pointed horns, thin in the median region; subunci very robust, bent close to their massive base and bearing a distinct apical hook; tegumen fairly large, its lateral margin markedly tooth-shaped; vinculum broad, with a long slender saccus; lower fultura consists of two divergent arms; valves oblong with rounded apices fused together by their lower edges in the basal half; penis elongate, gradually tapering in its internal portion, slightly dilated in its external portion, which ends in a sharp point, the meatus runs almost along its whole length and its edges bear spinules in its apical part; uncus and apices of valves long pilose.

In *Pseudonacaduba sichela* the male genitalia (Text-fig. 197) resemble those of *aethiops* except that the apices of the uncus, instead of tapering to a point, are crowned with a kind of volute ending in a hook, the subunci are shorter, and the valves more widely fused together, and with an obliquely truncated apex. In the figure the vesica, bearing fine cornuti, is exerted.

Karsch (1895, Ent. Nachr., 21: 297) placed sichela in the genus Orthomiella de Nicéville. Aurivillius (1898: 357), followed by most other authors of faunistic papers, included aethiops and sichela in the genus Nacaduba Moore.

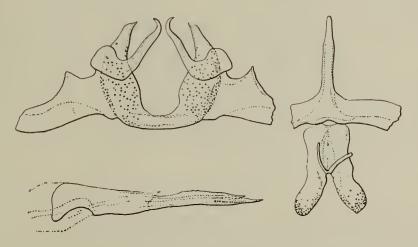


Fig. 196. Pseudonacaduba aethiops (Mabille), & genitalia.

I have compared the genitalia of aethiops and sichela with those of Orthomiella pontis sinensis and O.p. rovorea and several species of oriental Nacaduba. They had no similarity, and for this reason I erected the genus Pseudonacaduba.

In 1938 (Trans. R. ent. Soc. Lond. 1938: 125, 146) Corbet published a revision of of the Malayan species of the Nacaduba group, in which he stated "The genus Petrelaea Toxopeus (1929, Tijds. Ent. 72: 242), type Petrelaea dana varia Toxopeus (1929)...includes P. dana de Nicéville and certain African forms in the sichela group".

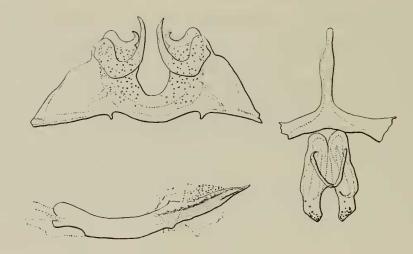


Fig. 197. Pseudonacaduba sichela (Wallengren), & genitalia.

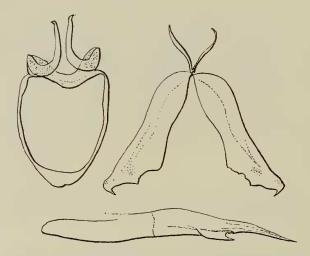


Fig. 198. Petrelaea dana dana (de Nicéville), o genitalia.

At that time, having no material of P. dana, I accepted Corbet's opinion. Since then, I have been able to dissect three males from the British Museum (N.H.) and to figure the male genitalia (Text-fig. 198). A glance at this figure will show they bear little resemblance to the genitalia of sichela: the uncus is much simpler, the vinculum very narrow and devoid of a saccus; the valves bigger and well separated, and the penis disproportionately large in relation to the dorsal structures. I consider therefore that there is good reason to retain the genus Pseudonacaduba for the two African species in spite of the undeniable superficial resemblance between dana and sichela.

## LIST OF SPECIES OF Pseudonacaduba

\*Pseudonacaduba aethiops (Mabille). Fig. Aurivillius in Seitz, 1925.

melania (Capronnier), 1889; stratola (Holland), 1891.

Pseudonacaduba dexamene (Druce), see sichela.

Pseudonacaduba docilis (Butler), see sichela.

Pseudonacaduba melania (Capronnier), see aethiops.

\*Pseudonacaduba sichela sichela (Wallengren) 1857. Fig. Aurivillius in Seitz,

dexamene (Druce), 1887; docilis (Butler), 1888.

Pseudonacaduba sichela reticulum (Mabille), 1877. Fig. Mabille, 1885: 87. Pseudonacaduba stratola (Holland), see aethiops.

### Genus *LEPIDOCHRYSOPS* Hedicke

Lepidochrysops Hedicke, 1923, Dt. ent. Z. 1923: 226 (nom. nov. pro Neochrysops Bethune Baker, 1923, nec Szilady, 1922); Pinhey, 1949: 117; Swanepoel, 1953: 89, 112. Typespecies: Papilio plebeius ruralis parsimon Fabricius, through Art. 67 (i) of Int. Com. zool. Nomenclature (replacement name).

Neochrysops Bethune Baker, 1923 (invalid junior homonym).

Cupido Schrank (partim); Aurivillius, 1898, 372; 1927: 479 et seq.

Lycaena Fabricius (partim); Murray, 1935: 164 et seq.

Catochrysops auct. plur., nec Boisduval, 1832.

Head rather small; eyes rather densely covered with short hair; palpi of medium size, second segment rather long, laterally compressed, clothed with white scales and some black bristles near the apex, third segment rather short, slender, acuminate; antennae slender, more than half the length of the costa, club elongate; thorax robust, clothed below with white silky hair; & fore leg, femur pilose, tibias as long as the femur, tarsus unsegmented, finely spinose below; mid and hind legs with a pair of short spurs at the apex of the tibia close to the metatarsus.

Wing shape. Fore wing subtriangular, apex angular, outer margin very slightly convex; hind wing oval, tailless in parsimon, but tailed in most species.

Wing venation (Text-fig. 332). Fore wing with 11 veins.

Male genitalia (Text-fig. 199). Uncus composed of two small lobes ("cheeks" of Bethune Baker) fused to the lateral angles of the tegumen; subunci long, curving, tapering gradually to the apex; tegumen consists of a median strip excised on its posterior edge; vinculum fairly wide; lower fultura comprising two small divergent arms fused to the base of the valves, and

a conical anellus fused to and sheathing the penis completely; valves very elongate with suboval bases, each valve being slightly dilated near its apex, which is recurved in the form of a short hook with slightly serrated lower edge; penis elongate, subcylindrical the tip slightly dilated ovoid and divided; vesica with fine cornuti; uncus, anterior edge of tegumen, lower edge and upper portions of the valves all pilose.

Although the genus *Lepidochrysops* contains a large number of species, the male genitalia are remarkably uniform throughout the genus. Three species alone, *niobe*, *lacrimosa* and *ariadne* differ in having shorter, wider valves and the apex of the penis distinctly hook-shaped. The genitalia of some species differ so slightly from one another that it is difficult to determine them by their genitalia alone. This situation is complicated further by the fact that the apices of the valves vary considerably in individuals of the same species (Text-fig. 200, *L. methymna* valves) and are sometimes asymmetrical. Bethune Baker (1923) described and illustrated the genitalia of a large number of species. References to these are to be found below.

The early stages of the following species have been described. L. procera and L. patricia by Murray (1935: 167, 172); L. puncticilia, L. bacchus and L. ortygia by Dickson (J. ent. Soc. sth. Afr. 1945, 7: 150; 1947, 9: 127); L. asteris, L. nisbe and L. ariadne by Pennington (J. ent. Soc. sth. Afr. 1946, 8: 23, 24).

# LIST OF SPECIES OF Lepidochrysops

Lepidochrysops acholi (Bethune Baker), see victoriae.

\*Lepidochrysops aethiopia (Bethune Baker), 1923, fig. and fig. genitalia. nyasae (Bethune Baker) 1923, fig.

Lepidochrysops albilinea Tite, 1959, Entomologist 92: 159, fig. Lepidochrysops ansorgei Tite, 1959: 160.

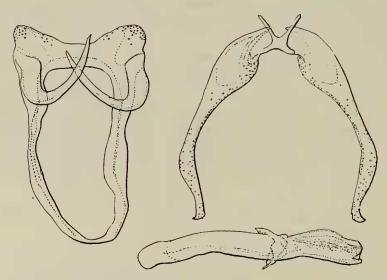


Fig. 199. Lepidochrysops parsimon parsimon (Fabricius), & genitalia.

Lepidochrysops arabicus Gabriel, 1954, S.W. Arabia Exp. 1937-38: 379, fig.

\*Lepidochrysops ariadne (Butler), 1898. Fig. genitalia, Bethune Baker, 1923.

\*Lepidochrysops asteris (Godart), 1824. Fig. genitalia, Bethune Baker, 1923. celaeus (Trimen) (in part), 1866; caffrariae (Trimen), 1887.

Lepidochrysops asteris (Trimen) (in part), see ortygia.

\*Lepidochrysops australis Tite, 1964, Entomologist 97: 6, fig. genitalia. ortygia Trimen, 1887 (in part).

\*Lepidochrysops azureus (Butler), 1879. Fig. Mabille, 1885-1887.

\*Lepidochrysops bacchus Riley, 1938, Trans. R. ent. Soc. Lond. 87: 243, fig. Lepidochrysops badhami van Son, 1956, Ann. Transv. Mus. 22: 508. Lepidochrysops barnesi Pennington, 1953, J. ent. Soc. sth. Afr. 16: 108, fig. Lepidochrysops brabo (Hulstaert) see skotios.

\*Lepidochrysops budama van Someren, 1957, J. ent. Soc. sth. Afr. 20: 65, fig. Lepidochrysops butha (Strand), 1911.

Lepidochrysops caerulea Tite, 1961, Entomologist 94: 21, fig.

Lepidochrysops caffrariae (Trimen), see asteris.

Lepidochrysops carsoni (Butler). Fig. and fig. genitalia, Bethune Baker, 1923. Lepidochrysops celaeus (Cramer), see parsimon.

Lepidochrysops celaeus (Trimen), see in parte asteris and trimeni.

\*Lepidochrysops chloauges (Bethune Baker), 1923. Fig. and fig. genitalia, Bethune Baker, 1923.

\*Lepidochrysops cinerea cinerea (Bethune Baker), 1923. Fig. and fig. genitalia, Bethune Baker, 1923.

menna (Hulstaert), 1924; theodota (Hulstaert), 1924.

\*Lepidochrysops cinerea kitale (Stempffer), 1937, Bull. Soc. ent. Fr. 42: 284. Lepidochrysops cinerea lunulifer (Ungemach), 1932, Mem. Soc. Sci. nat. Phys. Maroc 32: 92.

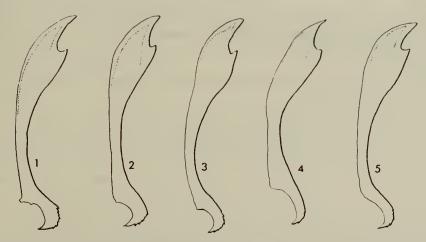


Fig. 200. Lepidochrysops methymna methymna (Trimen) valves showing variation, 1-3, Cape Town; 4-5, Kalk Bay.

Lepidochrysops cinerea imperialis (Stoneham), 1938, Bull. Stoneham Mus. 36:2.

Lepidochrysops cinerea princeps (Stoneham), 1938: 2.

\*Lepidochrysops coxii Pinhey, 1945, Proc. Trans. Rhod. Scient. Ass. 40: 61, fig.

\*Lepidochrysops cupreus (Neave), 1910. Fig. and fig. genitalia, Bethune Baker, 1923.

\*Lepidochrysops delicata (Bethune Baker), 1923, fig. and fig. genitalia.

\*Lepidochrysops desmondi Stempffer, 1951, Bull. Soc. ent. Fr. 56: 70, fig. genitalia.

\*Lepidochrysops dollmani (Bethune Baker) 1923, fig. and fig. genitalia.

\*Lepidochrysops elgonae elgonae Stempffer, 1950, Revue fr. Ent. 17: 147. Fig. van Someren, 1957, J. ent. Soc. sth. Afr. 20.

\*Lepidochrysops elgonae moyo van Someren, 1957, J. ent. Soc. sth. Afr. 20: 73,

Lepidochrysops exclusa (Trimen), see peculiaris.

Lepidochrysops flavisquamata Tite, 1959, Entomologist 92: 161, fig.

Lepidochrysops fulvescens Tite, 1961, Entomologist 94: 24, fig.

Lepidochrysops fumosa (Butler), see letsea.

\*Lepidochrysops gigantea (Trimen) 1898. Fig. and fig. genitalia, Bethune Baker, 1923.

\*Lepidochrysops glauca glauca (Trimen), 1887. Fig. and fig. genitalia, Bethune Baker, 1923.

\*Lepidochrysops glauca swinburnei Stevenson, 1939, Proc. Trans. Rhod. Scient. Ass. 34: 42.

\*Lepidochrysops grahami (Trimen), 1893. Fig. genitalia, Bethune Baker, 1923.

\*Lepidochrysops grandis Talbot, 1937, Trans. R. ent. Soc. Lond. 86: 65, fig.

\*Lepidochrysops guichardi Gabriel, 1949, Proc. R. ent. Soc. Lond. (B.) 18: 213,

\*Lepidochrysops hawkeri (Talbot), 1929, Bull. Hill Mus. Witley 3: 140, fig.

Lepidochrysops hypoleucus (Butler), see peculiaris.

\*Lepidochrysops hypopolia (Trimen), 1887. Fig. and fig. genitalia, Bethune Baker, 1923.

\*Lepidochrysops ignota (Trimen), 1887. Fig. Trimen, 1906. Fig. genitalia, Bethune Baker, 1923.

\*Lepidochrysops intermedia intermedia (Bethune Baker), 1923, fig. and fig. genitalia.

\*Lepidochrysops intermedia cottrelli Stempffer, 1954, Bull. Soc. ent. Fr. 59:

\*Lepidochrysops inyangae Pinhey, 1945, Proc. Trans. Rhod. scient. Ass. 40:64, fig.

Lepidochrysops irvingi Swanepoel, 1948, J. ent. Soc. sth. Afr. 11: 193, fig. Lepidochrysops jacksoni van Someren, 1957, J. ent. Soc. sth. Afr. 20: 67, fig.

\*Lepidochrysops jansei van Someren, 1957: 75, fig.

\*Lepidochrysops jefferyi (Sweistra), 1909.

Lepidochrysops kilimandjarensis (Strand), 1909.

Lepidochrysops koena (Strand), 1911.

Lepidochrysops labwor van Someren, 1957, J. ent. Soc. sth. Afr.: 68, fig.

- \*Lepidochrysops lacrimosa lacrimosa (Bethune Baker), 1923, fig. and fig. genitalia.
- \*Lepidochrysops lacrimosa major (Bethune Baker), 1923.
- \*Lepidochrysops lerothodi (Trimen), 1904. Fig. genitalia, Bethune Baker, 1923.
- \*Lepidochrysops letsea (Trimen), 1870. Fig. genitalia, Bethune Baker, 1923. fumosa (Butler), 1885.
- \*Lepidochrysops leucon (Mabille), 1879. Fig. Mabille, 1885-87.

Lepidochrysops loewensteini Swanepoel, 1951, J. ent. Soc. sth. Afr.: 57, fig.

- \*Lepidochrysops longifalces Tite, 1961, Entomologist 94: 22, fig. and fig. genitalia. Lepidochrysops lotana Swanepoel, 1962, J. ent. Soc. sth. Afr. 25: 291, fig. Lepidochrysops loveni (Aurivillius), see parsimon.
- \*Lepidochrysops lukenia van Someren, 1957, J. ent. Soc. sth. Afr.: 69, fig.
- \*Lepidochrysops mashuna (Trimen). Fig. and fig. genitalia, Bethune Baker, 1923.

Lepidochrysops menna (Hulstaert), see cinerea.

\*Lepidochrysops methymna methymna (Trimen), 1862. Fig. and fig. genitalia, Bethune Baker, 1923.

Lepidochrysops methymna dicksoni Tite, 1964, Entomologist 97: 7.

\*Lepidochrysops mpanda Tite, 1961, Entomologist 94: 23, fig.

Lepidochrysops nacrescens Tite, 1961: 24, fig.

Lepidochrysops naidina (Butler), 1885.

\*Lepidochrysops neavei (Bethune Baker), 1923, fig. and fig. genitalia.

\*Lepidochrysops negus (Felder), 1865. Fig. and fig. genitalia, Bethune Baker, 1923.

Lepidochrysops negus f. wau (Wichgraf), 1921.

Lepidochrysops negus (Karsch), see quassi.

\*Lepidochrysops neonegus neonegus (Bethune Baker), 1923, fig. and fig. genitalia.

variegata (Bethune Baker), 1923.

- \*Lepidochrysops neonegus borealis van Someren, 1957, J. ent. Soc. sth. Afr. 20: 64, fig.
- \*Lepidochrysops nevillei (Bethune Baker), 1923, fig. and fig. genitalia.
- \*Lepidochrysops nigeriae Stempffer, 1957, Bull. Inst. fr. Afr. noire 19: 225, fig. Lepidochrysops nigritia Tite, 1959, Entomologist 92: 162, fig.
- \*Lepidochrysops niobe (Trimen), 1862. Fig. Trimen, 1866. Fig. genitalia, Bethune Baker, 1923.

Lepidochrysops nyasae (Bethune Baker), see aethiopia.

\*Lepidochrysops nyika Tite, 1961, Entomologist 94: 23, fig.

\*Lepidochrysops oreas Tite, 1964, Entomologist 97: 4, fig. genitalia. ortygia Trimen 1887 (in parte).

Lepidochrysops orontius (Hulstaert), see skotios.

\*Lepidochrysops ortygia (Trimen), 1887. Fig. Trimen, 1906. Fig. genitalia, (Bethune Baker), 1923, fig. 15. asteris (Trimen), 1870, in parte.

\*Lepidochrysops pampolis (H. H. Druce), 1905. Fig. genitalia, Bethune Baker,

\*Lepidochrysops parsimon parsimon (Fabricius), 1775. Fig. genitalia, Bethune Baker, 1923.

celaeus (Cramer), 1782; loveni (Aurivillius), 1922.

Lepidochrysops parsimon f. albicans (Hulstaert), 1924, Revue zool. afr. 12: 143.

Lepidochrysops parsimon abyssiniensis (Strand), 1911.

\*Lepidochrysops parsimon anerius (Hulstaert), 1924, Revue zool. afr. 12: 139. Lepidochrysops parsimon kivuensis (Joicey & Talbot), 1921, Bull. Hill Mus. Witley 1: 99, fig.

\*Lepidochrysops parsimon oculus (Ungemach), 1932, Mem. Soc. Sci. nat. Phys.

Maroc 32: 92.

Lepidochrysops parsimon (Wallengren), see patricia.

\*Lepidochrysops patricia (Trimen), 1887. Fig. Trimen, 1906. Fig. genitalia, Bethune Baker, 1923.

parsimon (Wallengren), 1875 (nec Fabricius, 1775).

\*Lepidochrysops peculiaris (Rogenhofer), 1891. Fig. and fig. genitalia, Bethune Baker, 1923.

perpulchra (Holland), 1892; hypoleucus (Butler), 1894; exclusa (Trimen), 1894.

\*Lepidochrysops pephredo (Trimen), 1899. Fig. Trimen, 1906. Fig. genitalia, Bethune Baker, 1923.

Lepidochrysops perpulchra (Holland), see peculiaris.

Lepidochrysops phasma (Butler), see quassi.

\*Lepidochrysops plebeja plebeja (Butler), 1898. Fig. genitalia, Bethune Baker, 1923: 31.

\*Lepidochrysops plebeja proclus (Hulstaert), 1924, Revue zool. afr. 12: 137.

\*Lepidochrysops polydialecta (Bethune Baker), 1923, fig. and fig. genitalia. Lepidochrysops praeterite Swanepoel, 1962, J. ent. Soc. sth. Afr. 25: 293, fig.

\*Lepidochrysops procera (Trimen), 1893. Fig. genitalia, Bethune Baker, 1923.

\*Lepidochrysops pterou pterou (Bethune Baker), 1923.

\*Lepidochrysops pterou lilacina (Ungemach), 1932, Mem. Soc. Sci. nat. Phys. Maroc 32: 92.

\*Lepidochrysops pterou suk van Someren, 1957, J. ent. Soc. sth. Afr. 20: 61, fig.

\*Lepidochrysops puncticilia (Trimen), 1883. Fig. Trimen, 1887. Fig. genitalia, Bethune Baker, 1923.

\*Lepidochrysops quassi (Karsch), 1895. Fig. H. H. Druce, 1910. Fig. genitalia, Bethune Baker, 1923.

negus (Karsch), 1893; phasma (Butler), 1901.

\*Lepidochrysops reichenowi (Dewitz), 1879.

Lepidochrysops rhodesendae (Bethune Baker), 1923. Fig. and fig. genitalia, Bethune Baker, 1923.

Lepidochrysops ringa Tite, 1959, Entomologist 92: 163, fig.

Lepidochrysops ruthica Pennington, 1953, J. ent. Soc. sth. Afr. 16: 107, fig.

\*Lepidochrysops skotios (H. H. Druce), 1905. Fig. genitalia, Bethune Baker, 1923.

orontius Hulstaert, 1924; brabo (Hulstaert), 1924.

\*Lepidochrysops solwezi (Bethune Baker), 1923, fig. and fig. genitalia.

\*Lepidochrysops stormsi (Ribbe), 1892. Fig. Aurivillius, 1898. Fig. genitalia, Bethune Baker, 1923.

\*Lepidochrysops subvariegata Talbot, 1935, Entomologist's mon. Mag. 71: 150, fig.

Lepidochrysops swanepoeli Pennington, 1948, J. ent. Soc. sth. Afr. 11: 164, fig. Lepidochrysops sylvius (Hulstaert), see synchrematiza.

\*Lepidochrysops synchrematiza (Bethune Baker), 1923. Fig. and fig. genitalia sylvius (Hulstaert), 1924.

\*Lepidochrysops tantalus (Trimen), 1887. Fig. Trimen, 1906. Fig. genitalia, Bethune Baker, 1923.

Lepidochrysops theodota (Hulstaert), see cinerea.

\*Lepidochrysops trimeni (Bethune Baker), 1923, fig. and fig. genitalia. celaeus (Trimen) (in parte).

\*Lepidochrysops vansoni Swanepoel, 1949, J. ent. Soc. sth. Afr. 12: 123, fig. Lepidochrysops variegata (Bethune Baker), see neonegus.

Lepidochrysops vera Tite, 1961, Entomologist 94: 25, fig.

\*Lepidochrysops victoriae victoriae (Karsch), 1895. Fig. H. H. Druce, 1910. Fig. genitalia, Bethune Baker, 1923. acholi (Bethune Baker), 1906.

\*Lepidochrysops victoriae vansomereni Stempffer, 1951, Bull. Soc. ent. Fr. 56:

\*Lepidochrysops violetta Pinhey, 1945, Proc. Trans. Rhod. scient. Assoc. 40:66, fig.

Lepidochrysops wykehami Tite, 1964, Entomologist 97: 5, fig. and fig. genitalia.

### Genus EUCHRYSOPS Butler

Euchrysops Butler, 1900, Entomologist 33; 1; Pinhey, 1949:118; Swanepoel, 1953:113, 114. Type-species: Hesperia cnejus Fabricius (Indo-Malayan species) by original designation.

Cupido Schrank (partim); Aurivillius, 1898: 373; 1925: 473, 483, 484. Lycaena Fabricius (partim); Murray, 1935: 165, 167, 169, 170.

Eyes glabrous; palpi long, ascending, second segment laterally compressed clothed below with long white scales, third segment short, acuminate; antennae slender, about half the length of the costa, club elongate, gradually swollen, clavate; thorax clothed below with white silky hair;  $\Im$  fore leg, femur clothed with white hair, tibia shorter than femur, tarsus unsegmented.

Wing shape. Fore wing subtriangular; hind wing oval, a very short linear tail at the end of vein 2, anal angle rounded.

Wing venation (Text-fig. 333). Fore wing with 11 veins.

Male genitalia (Text-fig. 201). Uncus composed of two small lobes ("cheeks" of Bethune Baker) fused to the margin of the tegumen; subunci long, bent about one-third from origin, tapering gradually to the apex which bears a wide-open claw, tegumen fairly large, the posterior edge with a rounded depression so deep that it leaves only a narrow median strip; vinculum narrow in the sternite; lower fultura Y-shaped and bearing an anellus which sheathes the penis; valves elongate oval at the base, narrow in the middle, the upper part deeply notched just before the crescentic apex, whose lower edge is slightly serrate; penis elongate, the internal portion subcylindrical, the external portion dilated at the apex; vesica with numerous fine cornuti, uncus bearing long, fine hair, valves with stiffer hair especially on the lower edge in the basal half.

The male genitalia of all the *Euchrysops* are like those of *E. cnejus*, and most of them have been figured by Bethune Baker (1923). They differ little from those of *Lepidochrysops*, and I find difficulty in defining the limits of these two genera. On the other hand they are far removed from *strabo* Fabricius, type-species of the genus *Catochrysops* Boisduval, in which Fruhstorfer (1924 in Seitz, *Macrolep.* 9:921) surprisingly included species belonging to two other quite different genera, namely *cnejus* (*Euchrysops*) and *pandava* (*Chilades*).

In the following list it will be noticed that two species are included, on account of the structure of their male genitalia, which are commonly included in genera only remotely related to *Euchrysops*. These are *unigemmata* Butler, described as a *Zizera* but transferred later by Butler to *Chilades*, and *crawshayi* Butler described as a *Scolitantides*, but placed by Aurivillius in *Harpendyreus*.

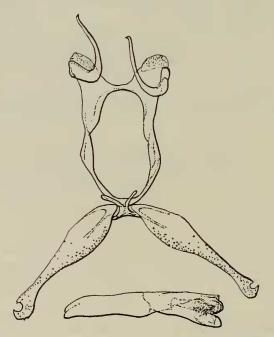


Fig. 201. Euchrysops cnejus (Fabricius), & genitalia.

The early stages of some species of *Euchrysops* have been described, viz. *E. crawshayi* by Jackson (1937, *Trans. R. ent. Soc. Lond.* 86: 234), *E. malathana* by Farquharson (1921, *Trans. ent. Soc. Lond.* 1921: 377), by Jackson (1937: 235) and by Pinhey (1949: 118), *E. osiris* by Jackson (1937: 236) and Pinhey (1949: 119).

# LIST OF SPECIES OF Euchrysops

\*Euchrysops abyssinica (Aurivillius), 1922.

\*Euchrysops albistriatus (Capronnier), 1889. Fig. genitalia, Bethune Baker, 1923.

Euchrysops anubis (Snellen), see osiris.

Euchrysops asopus (Hopffer), see malathana.

Euchrysops asteris (Snellen), see barkeri.

\*Euchrysops barkeri (Trimen), 1893. Fig. genitalia, Bethune Baker, 1923. asteris (Snellen), 1872; tiressa (Karsch), 1985; osiris Q (Trimen), 1887.

Euchrysops browni Stempffer, see unigemmata.

\*Euchrysops brunneus Bethune Baker, 1923, fig. genitalia.

Euchrysops conguensis (Mabille), see malathana.

\*Euchrysops crawshayi (Butler), 1899. Fig. genitalia, Stempffer, 1938, Mission Omo 4: 204.

subdita (Smith), 1902; crawshayinus (Aurivillius), 1925.

Euchrysops crawshayinus (Aurivillius), see crawshayi.

Euchrysops cuprescens (E. Sharpe), see osiris.

- \*Euchrysops cyclopteris (Butler), 1876. Fig. and fig. genitalia, Bethune Baker, 1923.
- \*Euchrysops decaryi Stempffer, 1947, Revue fr. Ent. 14: 139, fig. genitalia.
- \*Euchrysops dolorosa (Trimen), 1887. Fig. Trimen, 1906. Fig. genitalia, Bethune Baker, 1923.

Euchrysops fescennia Hulstaert, see subpallida.

Euchrysops horus (Stoneham), 1938, Bull. Stoneham Mus. 36: 3.

- \*Euchrysops jacksoni Stempffer, 1952, Bull. Soc. ent. Fr. 57: 117. Fig. and fig. genitalia.
- \*Euchrysops kabrosae (Bethune Baker), 1906. Fig. and fig. genitalia, Bethune Baker, 1923.

Euchrysops kama (Trimen), see malathana.

Euchrysops katangae Bethune Baker, 1923, fig. and fig. genitalia.

Euchrysops latruncula (Grünberg), 1910.

Euchrysops lois (Butler), 1896.

\*Euchrysops malathana malathana (Boisduval), 1833. Fig. genitalia, Bethune Baker, 1923.

asopus (Hopffer), 1855; kama (Trimen), 1862; conguensis (Mabille), 1877.

Euchrysops malathana nilotica (Aurivillius), 1904.

\*Euchrysops mauensis mauensis Bethune Baker, 1923, fig. and fig. genitalia. Euchrysops mauensis abyssiniae Storace, 1950, Doriana 1 (7): 1.

\*Euchrysops migiurtinensis Stempffer, 1946, Revue fr. Ent. 13: 17, fig. and fig. genitalia.

\*Euchrysops nandensis nandensis (Neave), 1904. Fig. genitalia, Bethune Baker, 1923.

Euchrysops nandensis abyssiniae (Storace), 1953, Doriana 1 (35): 3.

\*Euchrysops osiris osiris (Hopffer), 1855. Fig. Hopffer, 1862. Fig. genitalia, Bethune Baker, 1923.

anubis (Snellen), 1872; phoa (Snellen), 1872; pyrrhops (Mabille), 1877; cuprescens (E. Sharpe), 1898.

Euchrysops osiris australis Hulstaert, 1924, Revue zool. afr. 12: 136.

Euchrysops osiris orientalis Hulstaert, 1924, Revue zool. afr. 12: 136.

Euchrysops osiris ♀ (Trimen), nec Hopffer, see barkeri.

Euchrysops philbyi Gabriel, 1954, S.W. Arabia Exp. 1937-38: 386, fig.

Euchrysops phoa (Snellen), see osiris.

Euchrysops pyrrhops (Mabille), see osiris.

\*Euchrysops reducta reducta Hulstaert, 1924, Revue zool. afr. 12: 134. Fig. genitalia, Stempffer, 1961, Annls Mus. R. Afr. centr. 94: 67.

Euchrysops reducta niveocincta Ungemach, 1932, Mem. Soc. Sci. nat. Phys. Maroc 32: 93.

\*Euchrysops severini Hulstaert, 1924, Revue zool. afr. 12: 134.

Euchrysops subdita (Smith), see crawshayi.

\*Euchrysops subpallida subpallida Bethune Baker, 1923, fig. and fig. genitalia. fescennia Hulstaert, 1924.

Euchrysops subpallida major Bethune Baker, 1923.

Euchrysops tiressa (Karsch), see barkeri.

\*Euchrysops unigemmata (Butler), 1895. browni Stempffer, 1954.

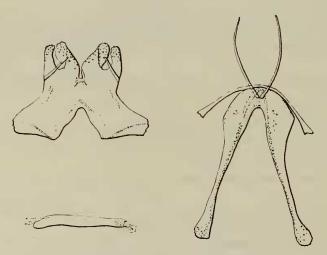


Fig. 202. Eicochrysops eicotrochilus Bethune Baker, & genitalia.

### Genus EICOCHRYSOPS Bethune Baker

Eicochrysops Bethune Baker, 1924, Ann. Mag. nat. Hist. (9) 14: 132; Pinhey, 1949: 119; Swanepoel, 1953: 70. Type-species: Eicochrysops eicotrochilus Bethune Baker, by original designation.

Cupido Schrank (partim); Aurivillius, 1898: 376; 1925: 487. Lveaena Fabricius (partim); Murray, 1935: 151, 152, 178.

Eyes smooth; palpi long, ascending, second segment long, laterally compressed, clothed below with adpressed scales and long erect bristles, third segment slender, acuminate; antennae slender, longer than half the length of the costa, club fusiform; thorax clothed below with long white silky hair; 3 fore tarsus unsegmented.

Wing shape. Fore wing subtriangular, outer margin slightly convex; hind wing oval, tailless in the type-species.

Wing venation (Text-fig. 334). Fore wing with 11 veins.

Male genitalia (Text-fig. 202). Uncus has four lobes, the median pair rolled up in the shape of a cornet with irregularly toothed edges, the lateral pair nodose at their apices; subunci straight, slender, almost filiform; tegumen very large, hood-shaped, so that, when viewed in profile in the natural position, the lateral lobes of uncus appear as if situated below the median lobes; vinculum very narrow; lower fultura formed of two long slender arms; anellus absent; valves long, narrowly digitate, with spatulate apices; penis very small, elongate, cylindrical; vesica with fine cornuti; uncus and lower edges of valves bearing long hair.

The male genitalia of all the species examined closely resemble those of eicotrochilus and it is easier to distinguish the species by external characters than by their genitalia. In pusillus, distractus, hippocrates and sanguigutta the hind wing has a filiform tail at the end of vein 2, which character would suffice to exclude them from Eicochrysops if their male genitalia did not resemble so closely those of eicotrochilus. I illustrate (Text-fig. 203) under higher magnification half the uncus and tegumen of hippocrates to show more clearly the peculiar shape of these parts. E. hippocrates, incidentally has been referred, in faunistic works, to various different genera, including Cupido, Everes and Cupidopsis.

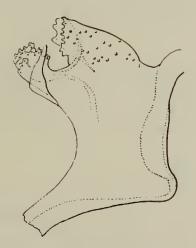


Fig. 203. Eicochrysops hippocrates (Fabricius), & genitalia.

In spite of the differences in the external appearance of its species the genus *Eicochrysops* seems very homogeneous.

The early stages of nandianus and messapus have been described by Jackson (1937, Trans. R. ent. Soc. Lond. 86: 236), by Dickson (1945, J. ent. Soc. sth. Afr. 7: 150) and by Pinhey (1948: 119).

Aurivillius (1898: 376 and 1925: 487) places scintilla Mabille close to E. sanguigutta, but the only characters common to these two species are their habitat (Madagascar) and their commonplace venation.

Male genitalia of scintilla (Text-fig. 204). Uncus composed of two small rounded lobes fused to the tegumen on either side of the median concavity of the posterior edge; no subunci; tegumen and vinculum large; lower fultura with long slender divergent arms; valves oblong, oval, apex rounded and bearing a strong spine bent back towards the body of the valve, penis very robust, base dilated and heart-shaped; vesica with numerous cornuti and some strong spines; uncus and distal portions of valves pilose.

The male genitalia of scintilla are unlike any that I know among Ethiopian species; the penis with its heart-shaped base reminds one of the penis of palaearctic species of Maculinea, but that is the only point of resemblance. This species seems to me to be unique and its place in the classification is uncertain. In any case it cannot be included in Eicochrysops, nor in Nacaduba where it has been placed by Lathy (Encycl. ent. B. II Lep. 2: 40, 1926).

# LIST OF SPECIES OF Eicochrysops

Eicochrysops antoto (Strand), 1911. Fig. Aurivillius in Seitz, 1925. Eicochrysops coeruleoarcuatus (Saalmuller); see sanguigutta. Eicochrysops delicatula (Mabille), see hippocrates.

\*Eicochrysops distractus (de Joannis), 1913.

\*Eicochrysops dudgeoni Riley, 1929, Trans. ent. Soc. Lond. 77: 497, fig.

\*Eicochrysops eicotrochilus Bethune Baker, 1924, Ann. Mag. nat. Hist. (9) 14:132.

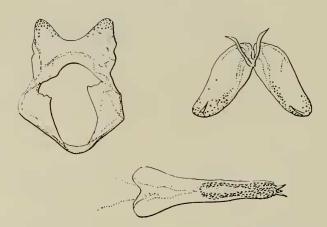


Fig. 204. Genus? scintilla Mabille, & genitalia.

- \*Eicochrysops fontainei Stempffer, 1961, Annls Mus. R. Afr. cent. 94: 68, fig. and fig. genitalia.
- \*Eicochrysops hippocrates (Fabricius), 1793. Fig. Mabille, 1885–87. delicatula (Mabille), 1877.
- \*Eicochrysops mahallakoena mahallakoena (Wallengren), 1857. Fig. Trimen, 1870.
- \*Eicochrysops mahallakoena trisignatus (Strand), 1911.
- \*Eicochrysops masai (Bethune Baker), 1905.
- \*Eicochrysops messapus (Godart), 1823. Fig. Aurivillius in Seitz, 1925. Eicochrysops messapus f. sebagadis (Guerin), 1847.
- \*Eicochrysops nandianus (Bethune Baker), 1906.
- \*Eicochrysops pauliani Stempffer, 1950, Naturaliste malgache 2: 131, fig.
- \*Eicochrysops pusillus (Ungemach), 1932, Mem. Soc. Sci. phys. Nat. Maroc 32: 94, 96, fig.
- \*Eicochrysops rogersi Bethune Baker, 1924, Ann. Mag. nat. Hist. (9) 14: 133.
- \*Eicochrysops sanguigutta (Mabille), 1879. Fig. Mabille, 1885-7. coeruleoarcuatus (Saalmuller), 1884.

Eicochrysops sapphirinus (Stoneham), 1938, Bull. Stoneham Mus., No. 36: 3.

## Genus incertus

Lycaena scintilla Mabille, 1877, Bull. Soc. ent. Fr. 5 (7): lxxii. quadriocularis Saalmuller, 1884.

#### Genus CUPIDOPSIS Karsch

Cupidopsis Karsch, 1895, Ent. Nach. 21: 297; Pinhey, 1949: 120; Swanepoel, 1953: 114. Type-species: Lycaena iobates Hopffer, by original designation.

Cupido Schrank (partim); Aurivillius, 1898: 377; 1925: 489.

Lycaena Fabricius (partim); Murray, 1935: 178.

Eyes smooth; palpi long, ascending, second segment long, laterally compressed, densely clothed with white adpressed scales, third segment shorter, slender, acuminate; antennae slender, white-annulated, about half the length of the costa, club clavate, well differentiated, thorax clothed below with long white silky hair;  $\eth$  fore leg, femur clothed with white hair, tibia shorter than the femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing subtriangular, outer margin convex, hind wing oval, with a filiform tail at the end of vein 2.

Wing venation (Text-fig. 335). Fore wing with only 10 veins.

Male genitalia (Text-fig. 205). Uncus composed of two small flattened lobes fused to the tegumen on either side of the median concavity, subunci robust, bent at about two-fifths from base, tapering suddenly a little before the apex, which is not hooked; tegumen large, hood-shaped, vinculum fairly broad with a rounded saccus; arms of lower fultura slender, strongly recurved; valves oblong, broadly fused to one another at the base and with rounded apices, their internal lamina bearing a kind of hook-like expansion, with a serrated apical edge; penis long, robust, slightly curved with a short external portion; vesica with two long rows of cornuti; uncus, hook-like expansion and apices of valves, pilose.

The male genitalia of cissus and mauritanica are almost identical with those of iobates. As cissus has no tail on its hind wing, Karsch (1895: 297) included it in the genus Neolycaena de Niceville (type-species sinensis Alpheraky). In my opinion this arrangement cannot be correct as, judging from the male genitalia, the genus Neolycaena belongs to the subfamily Theclinae, whereas cissus belongs to the Lampidinae; moreover the presence or absence of a tail in the hind wing has no generic value.

The early stages of *cissus* have been described by Jackson (1937, *Trans. R. ent. Soc. Lond.* **86**: 237). The caterpillar lives in the pods of *Eriosema cordifolium* Hochst (Leguminosae); it does not seem to be myrmecophilous.

# LIST OF SPECIES OF Cupidopsis

\*Cupidopsis cissus cissus (Godart), 1823. Fig. Mabille, 1885–87. Fig. genitalia, Stempffer, 1938, Mission Omo 4.

Cupidopsis cissus f. aberrans (Butler), 1878.

Cupidopsis cissus f. albiradiatus (Stoneham), 1938, Bull. Stoneham Mus., No. 36: 3.

Cupidopsis cissus f. immaculatus (Stoneham), 1938, l.c. No. 36: 3.

Cupidopsis cissus catharina (Trimen), 1862.

\*Cupidopsis iobates iobates (Hopffer), 1855. Fig. Hopffer, 1862. siwani (Trimen), 1862.

Cupidopsis iobates f. conjungens (Strand), 1911.

Cupidopsis iobates ochreopuncta (Aurivillius), 1925.

Cupidopsis iobates uranochroa Ungemach, 1932, Mem. Soc. Sci. nat. phys. Maroc 32: 97, fig.

Cupidopsis mauretanica Riley, 1932, Ann. Mag. nat. Hist. (10) 10: 141, fig. Cupidopsis siwani (Trimen), see iobates.

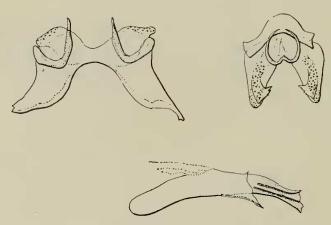


Fig. 205. Cupidopsis iobates iobates (Hopffer), & genitalia.

## Genus THERMONIPHAS Karsch

Thermoniphas Karsch, 1895, Ent. Nachr. 21: 303. Type-species: Thermoniphas plurilimbatus Karsch, by original designation.

Cupido Schrank (partim); Aurivillius, 1898: 377; 1925: 493.

Eyes smooth; palpi long, second segment laterally compressed, clothed below with long erect white scales, third segment shorter, slender, acuminate, clothed with small black adpressed scales, antennae slender, white annulated, longer than half the length of the costa, club flattened; thorax clothed below with silky white hair; of fore leg, tibia shorter than the femur, tarsus unsegmented, clothed below with stiff hair.

Wing shape. Fore wing subtriangular, outer margin convex; hind wing oval, a short filiform tail at the end of vein 2.

Wing venation (Text-fig. 336). Fore wing with 11 veins; 11 runs close to 12 and sometimes touches it at one point.

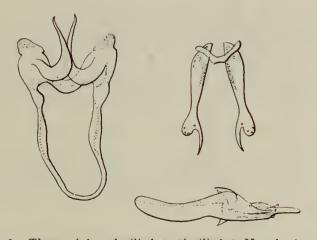


Fig. 206. Thermoniphas plurilimbata plurilimbata Karsch, & genitalia.

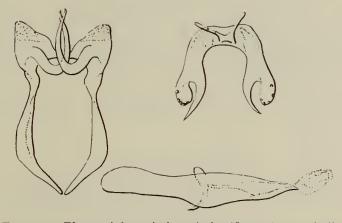


Fig. 207. Thermoniphas micylus micylus (Cramer), of genitalia.

Male genitalia (Text-fig. 206). Uncus composed of two small suboval lobes like those in Lepidochrysops and Euchrysops fused on either side of the tegumen; subunci long, robust, curved, tapering evenly to the apex which ends in a shallow hook; tegumen fairly large, reduced on the median line to a narrow strip; vinculum narrow, lower fultura formed of two small divergent arms to which is fused an anellus that sheathes the penis; valves very narrow, upper process with rounded apex, lower process ending in a slightly recurved point; penis elongate, swollen in the middle, slightly curved; vesica with fine cornuti; uncus clothed with long fine hair, a few short hairs on the lower edges of the valves, their apices bearing four bristles, of which the one on the upper process is much longer and stronger than the others.

The species comprising the genus *Thermoniphas* have been described in a variety of genera, e.g. *Cupido*, *Oboronia* and *Everes*. Bethune Baker (1923: 277) even relegated *micylus* (Text-fig. 207) and *togara* (Text-fig. 208) to the genus *Lycaenopsis*,

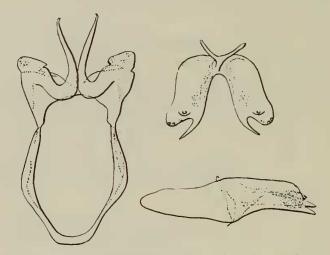


Fig. 208. Thermoniphas togara togara (Plötz), o genitalia.

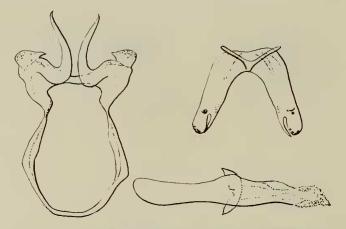


Fig. 209. Thermoniphas distincta (Talbot), of genitalia.

which is hard to understand, since he had examined their genitalia. In 1943 (Ann. Soc. ent. Fr. 1942: 131) I suggested that micylus (Text-fig. 207) should be transferred to Thermoniphas. In 1956 (Annls Mus. R. Congo belge 38: 52) I revised the genus as known at that date and figured the male genitalia of nearly all the species. On the basis of the evidence thus presented, the homogeneity of the genus is clear, and also its affinity to Lepidochrysops and Euchrysops on the one hand and to Oboronia on the other. A remarkable generic character peculiar to the genus Thermoniphas is the presence of the long stout bristles that arise from the apex of the valves.

# LIST OF SPECIES OF Thermoniphas

- \*Thermoniphas alberici (Dufrane), 1945, Bull. Soc. ent. Belg. 81:123. Fig. genitalia, Stempffer, 1956.
- \*Thermoniphas albocaerula Stempffer, 1956: 50, fig. and fig. genitalia. Thermoniphas bibundana (Grünberg), 1910.
- \*Thermoniphas caerulea Stempsfer, 1956: 49, fig. and fig. genitalia.
- \*Thermoniphas distincta (Talbot), 1935, Entomologist's mon. Mag. 71: 149. Fig. genitalia, Stempffer, 1956: 43.
- \*Thermoniphas fontainei Stempffer, 1956: 47, fig. and fig. genitalia.
- \*Thermoniphas fumosa Stempffer, 1952, Bull. Soc. ent. Fr. 57: 119, fig. and fig. genitalia.
- Thermoniphas kamitugensis (Dufrane), 1945, Bull. Annls Soc. ent. Belg. 81: 122.
- \*Thermoniphas kigezi Stempffer, 1956: 48, and fig. genitalia.
- \*Thermoniphas leucocyana Clench, 1961, Ann. Carnegie Mus. 36; 56, fig. and fig. genitalia.
- \*Thermoniphas micylus micylus (Cramer), 1780. Fig. genitalia, Stempffer, 1956: 41.
- \*Thermoniphas micylus colarata (Ungemach), 1932, Mem. Soc. Sci. nat. Phys. Maroc 32: 97. Fig. and fig. genitalia, Stempffer, 1956, 42.
- \*Thermoniphas plurilimbata plurilimbata Karsch, 1895.
- \*Thermoniphas plurilimbata rutshurensis (Joicey & Talbot), 1921, Bull. Hill Mus. Witley 1:99. Fig. genitalia, Stempffer, 1956, 40.
- \*Thermoniphas stempfferi Clench, 1961, l.c.: 52, fig. and fig. genitalia.
- \*Thermoniphas togara togara (Plötz), 1880. Fig. and fig. genitalia, Stempffer, 1956: 45.
- \*Thermoniphas togara bugalla Stempffer & Jackson, 1962, Proc. R. ent. Soc. Lond. (B.) 31:35.

### Genus OBORONIA Karsch

Oboronia Karsch, 1893, Berl. ent. Z. 38: 229. Type-species: Plebeius punctatus Dewitz, 1879 (Lycaena elorea Staudinger, Karsch, 1893, nec Papilio elorea Fabricius 1793; Obornia staudingeri Hemming 1960), selected by Hemming, 1960.

Cupido Schrank (partim); Aurivillius, 1898; 380; 1925; 493.

In his description of the genus, Karsch does not designate a type-species, but gives as included species elorea Staudinger (with a reference to Staudinger's figure in Exot. Schmett., pl. 94) and ornata Mabille. In the Ent. Nachr., 21: 297, 1895, the same author erects for ornata the genus Athysanota and under Oboronia he writes "zu Oboronia gehoren punctata Dewitz = elorea Staudinger nec Fabricius, gussfeldti Dewitz, etc." Hemming (1960, Annot. Lep. 1: 35) pointed out that there was no such nominal species as Lycaena elorea Staudinger, gave the name Oboronia staudingeri to the species depicted by Staudinger and then selected Oboronia staudingeri as type-species of the genus Oboronia. The oldest name for this species is, however, Plebeius punctatus Dewitz, 1879.

Eyes smooth; palpi long, ascending, second segment long, laterally compressed clothed below with long white scales and stiff hair, third segment short, acuminate; antennae slender, about half the length of the costa, club very elongate and flattened; thorax clothed below with white hair;  $\eth$  fore leg, femur slightly flattened, tibia much shorter than femur, tarsus unsegmented.

Wing shape. Fore wing subtriangular, costa and outer margin strongly convex, hind wing oval, a short linear tail at the end of vein 2.

Wing venation (Text-fig. 337). Fore wing with 11 veins; 10 and 11 free from the upper edge of the cell. Aurivillius (1898: 380) points out that the differences in the venation between Thermoniphas and Oboronia are slight and inconstant; this is true, for in certain specimens of punctatus vein 11 bends towards 12 and is sometimes confluent with it.

Male genitalia (Text-fig. 210). Uncus composed of two small subtriangular lobes with rounded apices fused to the lateral angles of the tegumen; subunci long, bent about two-fifths from base, tapering evenly from base to apex which ends as a shallow hook; tegumen with a rounded median hollow on posterior margin; vinculum fairly wide; lower fultura with two short divergent arms to which is fused an anellus that sheathes the penis; valves very elongate, widened at their base, then digitate, the lower process slightly serrated at its apex, penis elongate, internal portion cylindrical, external portion slightly dilated and divided into two processes; vesica with numerous cornuti; uncus and apices of valves pilose.

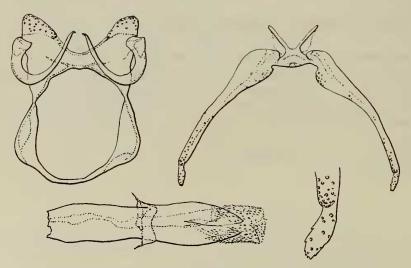


Fig. 210. Oboronia punctatus punctatus (Dewitz), & genitalia.

The male genitalia of all the species examined, except *liberiana*, closely resemble those of punctatus, the slight differentiating specific characters being found in variations in the shape of the apex of the valves. In O. liberiana, (fig., Stempffer, 1950, Bull. Inst. fr. Afr. noire 12:406) the dorsal structures are very characteristic; tegumen very wide with only a small rounded median notch, the uncus a narrow strip bordering the posterior edge of the tegumen and interrupted at the notch, subunci long, bent, very robust. But the vinculum, lower fultura, valves and penis are of the type common to the other species of Oboronia.

The larva of O. punctatus has been described by Poulton (1911, Proc. ent. Soc. Lond. 1911: ci) and by Lamborn (1913, Trans. ent. Soc. Lond. 46: 489). It lives in the flowers of Costus afer and ants of the genus Pheidole construct shelters over them from plant debris.

### LIST OF SPECIES OF Oboronia

- \*Oboronia albicosta (Gaede), 1915.
- \*Oboronia bueronica Karsch, 1895. Fig. Aurivillius in Seitz, 1925.
  - Oboronia elorea (Staudinger), see punctatus.
- \*Oboronia gussfeldti (Dewitz), 1879.
- \*Oboronia liberiana Stempffer, 1950, Bull. Inst. fr. Afr. noire 12: 405, fig. and fig. genitalia.
- \*Oboronia pseudopunctatus Strand, 1912.
- \*Oboronia punctatus punctatus (Dewitz), 1879. elorea (Staudinger) nec Fabricius; staudingeri Hemming, 1960.
- \*Oboronia punctatus arctimargo Hulstaert, 1924, Revue zool. afr. 12: 143.
- \*Oboronia punctatus jacksoni Stempffer, 1943, Ann. Soc. ent. Fr. 1942: 132. Oboronia staudingeri Hemming, see punctatus.

## Genus ATHYSANOTA Karsch

Athysanota Karsch, 1895, Ent. Nachr. 21: 297. Type-species: Lycaena ornata Mabille, by monotypy.

Cupido Schrank (partim); Aurivillius, 1898: 381; 1925: 494.

Eyes smooth; palpi long, ascending, second segment long, laterally compressed, clothed below with long erect scales, which are white at the base and black at the apex, third segment slender, cylindrical, acuminate; antennae slender, about half the length of the costa, club very elongate, not well differentiated, apex pointed; thorax clothed below with white hair; 3 fore leg, tibia much shorter than the femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing subtriangular, costa and outer margin strongly convex, hind wing oval, no tail.

Wing venation (Text-fig. 338). Fore wing with 11 veins.

Male genitalia (Text-fig. 211). Uncus composed of two small subquadrangular lobes fused to the lateral angles of the tegumen; subunci very long, slender, bent about one-third from origin, apex ending as a shallow hook; tegumen with a rounded depression in its posterior margin, vinculum rather broad; lower fultura comprising two small divergent arms fused to an anellus that sheathes the penis; valves digitate, a little dilated in the middle, lower process

with a recurved apex which bears a few spines, its edge deeply serrate; penis cylindrical in its internal portion, the external portion dilated and divided into two processes; vesica with fine cornuti; uncus and valves pilose.

Both in external appearance, and the structure of the male genitalia, A. ornata is closely related to the species of Oboronia.

## LIST OF SPECIES OF Athysanota

\*Athysanota ornata ornata (Mabille), 1890.

pseudosoyauxi (Ehrmann), 1894.

Athysanota ornata flava Holland, 1920.

Athysanata ornata vestalis Aurivillius, 1895.

Athysanota pseudosoyauxi (Ehrmann), see ornata.

## Genus CHILADES Moore

Chilades Moore, 1881, Lep. Ceylon 1:76. Type-species: Papilio laius Cramer (an Indo-Malayan species), by original designation.

Cupido Schrank (partim); Aurivillius, 1898: 375, 381; 1925: 485, 490, 491.

Eyes with very short, scarcely visible hair; palpi long, ascending, second segment long, strongly compressed laterally, clothed below with long erect bristles, third segment short, slender, antennae slender, about half the length of the costa club distinct, but not very stout, with pointed tip; thorax clothed below with long white silky hair; of fore leg, femur clothed

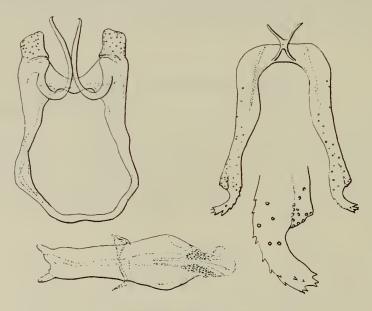


Fig. 211. Athysanota ornata ornata (Mabille), & genitalia.

with white hair, tibia shorter than the femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing subtriangular, costa slightly convex, apex rounded, outer margin convex; hind wing oval, no tail.

Wing venation (Text fig. 339).

Male genitalia (Text-fig. 212) (see also Bethune Baker, 1913, Trans. ent. Soc. Lond. 1913 pl. 4 and Stempffer, 1937, Bull. Soc. ent. Fr. 42: 214). Uncus composed of two long digitate processes with rounded apices; subunci long, bent in an acute angle close to their massive bases, apices hooked, tegumen reduced as in all species of the subfamily Plebeiinae; arms of the 9th sternite united dorsally to form a pseudotergum (for an account of this structure, peculiar to the Plebeiinae, see Bayard (1933, Bull. Soc. fr. Microsc. 3: 4), and Stempffer, (1937, Bull. Soc. ent. Fr. 42: 213, fig. A); lower fultura in the form of a furca with very long slender arms; valves fusiform, the upper process connected in its middle to the pseudotergum by a weakly sclerotized membrane, traces of which can be seen in the dorsal region, ending in a finely toothed comb, lower process with a rounded apex; penis elongate, slightly curved, apex pointed; uncus and lower process of valves densely pilose.

I place in the genus *Chilades* the following African species, of which I have been able to examine the male genitalia, *eleusis*, *nigeriae*, *elicola*, *kedonga* and *parrhasius*, their genitalia being of the same pattern as in *laius*; those of *eleusis*, *nigeriae* and *elicola* are indeed so similar that they may be races of a single species. In *kedonga* (see Text-fig. 213) the valves are shorter and broader, but the other parts are very like those of *laius*.

I have not been able to examine the genitalia of sanctithomae and alberta, and only refer them to the genus Chilades with reserve.

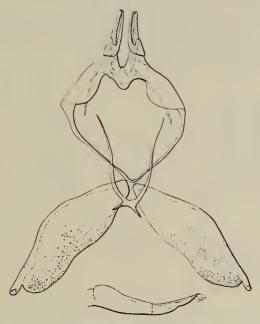


Fig. 212. Chilades laius (Cramer), & genitalia.

## LIST OF SPECIES OF Chilades

Chilades alberta Butler, 1901.

Chilades contracta (Butler), see parrhasius.

\*Chilades eleusis eleusis (Demaison), 1888. Fig. Aurivillius in Seitz, 1925. Fig. Genitalia, Stempffer, 1936, Livre jubilaire Bouvier: 324. podorina (Mabille), 1890; pharaonis (Staudinger), 1894.

Chilades eleusis strigatus (Aurivillius), 1925.

- \*Chilades elicola (Strand), 1911. Fig. Aurivillius in Seitz, 1925.
- \*Chilades kedonga (Gr. Smith), 1898. Fig. Butler, 1899. pulchristriata (Bethune Baker), 1905.
- \*Chilades nigeriae (E. Sharpe), 1902.
- \*Chilades parrhasius (Fabricius), 1793. contracta Butler, 1880.

Chilades pharaonis (Staudinger), see eleusis.

Chilades podorina (Mabille), see eleusis.

Chilades pulchristriata (Bethune Baker), see kedonga.

Chilades sanctithomae (E. Sharpe), 1893.

## Genus FREYERIA Courvoisier

Freyeria Courvoisier, 1920, Dt. ent. Z. Iris 34: 234; Pinhey, 1949: 120; Swanepoel, 1953: 80 Type-species: Lycaena trochylus Freyer, by monotypy.

Cupido Schrank (partim); Aurivillius, 1898: 376; 1925: 489, 495.

Lycaena Fabricius (partim); Murray, 1935: 150.

Eyes smooth; palpi long, ascending, second segment long, laterally compressed, clothed below with long stiff hair, third segment long, slender, cylindrical, acuminate; antennae slender, three-fifths the length of the costa, club fusiform, well differentiated; of fore leg, tibia as long as the femur, tarsus unsegmented, finely spinose below.

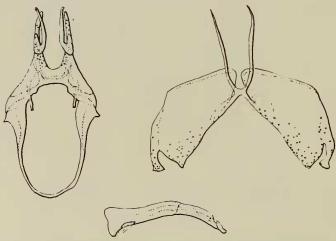


Fig. 213. Chilades kedonga (Grose Smith), & genitalia.

Wing shape. Fore wing subtriangular, costa and outer margin convex; hind wing oval, no tail. According to Courvoisier I.c., Freyeria differs from Chilades in the absence of androconia. Wing venation (Text-fig. 340). Fore wing with 11 veins, 11 is bent towards 12 but not confluent with it.

Male genitalia (Text-fig. 214). Resembling closely those of Chilades, but the two processes of the uncus are less slender and shorter; subunci long, with massive bases, bent at an acute angle about one-third from origin, apex ending in a shallow hook; tegumen, pseudotergum and furca similar to those of Chilades; valves fusiform, typical of Plebeiinae, connected to the pseudotergum by a thin membrane, upper process ending in a comb, lower process with a rounded apex, penis elongate, slightly curved, widely open towards its slender apex, vesica with fine cornuti, uncus and valves especially the lower process, pilose.

The male genitalia of *minuscula* (Text-fig. 215) are like those of *trochylus* but differ as follows: Uncus a little more massive and with the apices of the lobes slightly recurved to form a hook; subunci less robust, especially at the base; comb of valves a little broader.

The species of *Freyeria* closely resemble those of *Chilades*. Both genera are distributed throughout almost the whole of the Old World. *Chilades* ranges from Egypt to the New Hebrides. *F. trochylus* is found in Europe in the Balkans and its range extends as far as Ferghana in Central Asia and in Africa to the Cape.

The caterpillar of F. trochylus has been observed by Pinhey (1949: 120) in Rhodesia where it feeds upon Heliotrope and Indigofera.

# LIST OF SPECIES OF Freyeria

- \*Freyeria minuscula (Aurivillius), 1909.
- Freyeria parva (Murray), see trochylus.
- \*Freyeria trochylus (Freyer), 1844. parva (Murray), 1874.

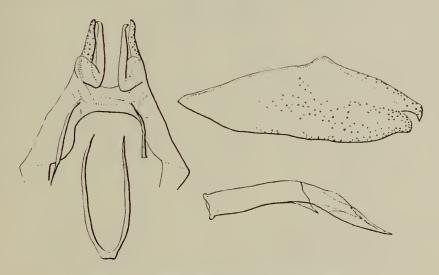


Fig. 214. Freyeria trochylus (Freyer), & genitalia.

## Genus AZANUS Moore

Azanus Moore, 1881, Lep. Ceylon 1:79; Pinhey, 1949:113; Swanepoel, 1953:58. Type-species: Papilio ubaldus Cramer, by original designation.

Cupido Schrank (partim); Aurivillius, 1898:364; 1925:468.

Lycaena Fabricius (partim); Murray, 1935:158.

Eyes densely pilose; palpi long, ascending, second segment long, laterally compressed, clothed below with long stiff white hair, third segment short, acuminate; antennae slender, about two-thirds the length of the costa, club oval, flattened well differentiated; thorax clothed below with long white silky hair;  $\delta$  fore leg, femur clothed with long white hair, tibia slightly shorter than the femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing subtriangular, apex angular, outer margin convex from the apex to the end of vein 4, then straight; inner margin straight, distinctly shorter than the costa; hind wing oval, no tail.

Wing venation (Text-fig. 341). Fore wing with 11 veins; 11 broadly anastomosed with 12. Male genitalia (Text-fig. 216). Uncus shaped like a breast-plate, its posterior margin deeply notched, each angle bearing a small, weakly sclerotized lobe; subunci short, massive, curved; tegumen reduced to a narrow band; vinculum narrow; lower fultura crescentic; valves very long, digitate, with an irregular distal edge and a slightly recurved apex; penis cylindrical, very long, curved, ending in a sharp point; vesica with fine cornuti; uncus clothed with long fine hair; coarser hair on the distal halves of the valves.

I have described and figured the genitalia of many species of *Azanus*. In all of them the uncus, subunci and tegumen are of the same shape as the corresponding

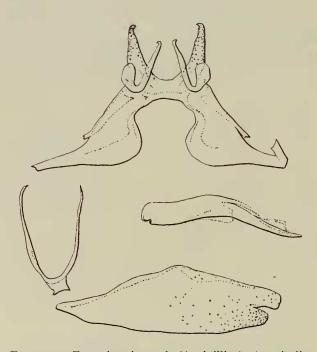


Fig. 215. Freyeria minuscula (Aurivillius), & genitalia.

parts in *ubaldus*, but the valves and the penis are rather more complicated in shape than in the type of the genus. The valves, digitate in *jesous*, apex deeply incised in *sitalces*, wider than in *ubaldus* in the other species, have the distal portion broken up into more or less slender processes in *isis* and *natalensis*. The penis is not so long and more robust in these other species than in *ubaldus*, the ventral surface of the external portion is grooved and ends in a slender point, and in the groove there is a more or less massive cuneus, which is bare in *sitalces* and *mirza*, bristling with spinules in *jesous*. *Azanus isis*, which on account of the pattern of the upperside

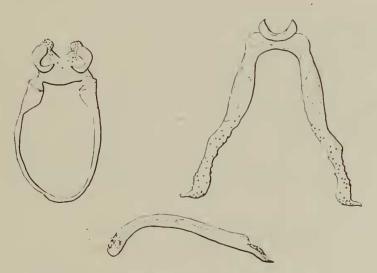


Fig. 216. Azanus ubaldus (Cramer), & genitalia.

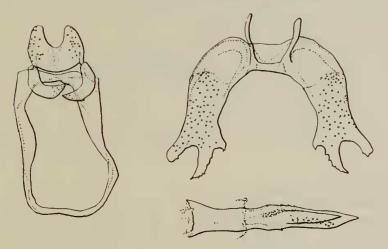


Fig. 217. Azanus isis (Drury), & genitalia.

markings has often been placed in *Castalius*, is quite unrelated to that genus; its genitalia (Text-fig. 217) are illustrated for comparison.

The early stages of *ubaldus*, *jesous* and *natalensis* have been described by Murray (1935: 159–160), Jackson (1937, Trans. R. ent. Soc. Lond. 86: 232–3) and Pinhey (1949: 114). The caterpillars feed on species of Mimosacae, e.g. Acacia abyssinica, A. stenocarpa and A. karroo.

## LIST OF SPECIES OF Azanus

Azanus agave (Walker), see jesous.

Azanus artemides (Stoll), see ubaldus.

Azanus benigna (Möschler), see moriqua.

Azanus camillus (Cramer), see isis.

Azanus coeruleoalbus (Goeze), see isis.

Azanus ethode (Walker), see ubaldus.

Azanus gamra (Lederer), see jesous.

Azanus isarchus (Fabricius), see isis.

\*Azanus isis (Drury), 1773.

coeruleoalbus (Goeze), 1779; camillus (Cramer), 1780; isarchus (Fabricius), 1793.

Azanus itea (Walker), see ubaldus.

\*Azanus jesous jesous (Guerin), 1847. Fig. genitalia, Stempffer, 1938, Mission Omo 4: 206.

gamra (Lederer), 1855; agave (Walker), 1870.

Azanus jesous soalalicus (Karsch), 1900.

Azanus macalenga (Trimea), see ubaldus.

\*Azanus mirza (Plötz) 1880. Fig. genitalia, Stempffer, 1938, Mission Omo 4: 209. ocidentalis Butler, 1888.

Azanus mirza f. deficiens Dufrane, 1953, Bull. Annls Soc. R. ent. Belg. 89: 55. Azanus mirza f. ornata Dufrane, 1953, l.c.: 55.

\*Azanus moriqua (Wallengren), 1857.

sigillata (Butler), 1876; benigna (Möschler), 1883.

\*Azanus natalensis (Trimen), 1887. Fig. Trimen, 1906. Fig. genitalia, Stempffer, 1938, Mission Omo 4: 209, (as sigillatus).

Azanus occidentalis Butler, see mirza.

Azanus rubropuncta Lathy, see sitalces.

Azanus sigillata (Butler), see moriqua.

\*Azanus sitalces (Mabille), 1899. Fig. genitalia, Stempffer, 1938, Mission Omo 4:207.

rubropuncta Lathy.

Azanus thebana (Staudinger), see ubaldus.

\*Azanus ubaldus (Cramer), 1782.

zena (Moore), 1865; artemides (Stoll), 1782; ethoda (Walker), 1870; itea (Walker), 1870; macalenga (Trimen), 1887; thebana (Staudinger), 1894.

Azanus zena (Moore), see ubaldus.

### Genus BREPHIDIUM Scudder

Brephidium Scudder, 1876, Bull. Buffalo Soc. nat. Sci. 3: 123; Swanepoel, 1953: 81. Typespecies: Lycaena exilis Boisduval (a Sonoran species) by original designation.

Cupido Schrank (partim); Aurivillius, 1898: 369; 1925: 473.

Lycaena Fabricius (partim); Murray, 1935: 150.

Eyes smooth; palpi long, second segment ascending, long, laterally compressed, clothed with white scales and bearing below long erect black bristles, third segment horizontal, slender, acuminate; antennae slender, about half the length of the costa, club oval, flattened, well differentiated; thorax clothed below with long white silky hair; 3 fore leg, femur clothed below with long white hair, tibia about as long as femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing subtriangular, apex rounded, outer margin convex; hind wing oval, no tail.

Wing venation (Text-fig. 342). Fore wing with 11 veins, 11 reduced to a short vein running from the upper edge of the cell to 12.

Male genitalia (Text-fig. 218). As Bethune Baker has already pointed out (1914, Trans. ent. Soc. Lond. 47: 330), the structure of the 3 genitalia of species of Brephidium differs considerably from that of all the other known Lycaenidae and it is difficult to establish the homologies of all the parts. Directly fused to the vinculum there is, on the posterior margin dorsally, an unpaired process of which the apex is deeply divided into four teeth, which may correspond to the uncus; on either side of this process the two lobes of the tegumen take an abnormal form; in the mount from which the figure was drawn they are spread out and flattened, but in situ they are convex and situated laterally; parallel to and arising from the external edge of these lobes there is a long process of which the rounded apex bears five strong rigid bristles which look like sharp-pointed thorns; vinculum narrow; the two slender arms of the lower fulture are fused, not to the base of the valves, but at two-thirds from their base, valves small, oval, with rounded apices, widely fused to each other along their lower edges; internal portion of penis bulbous, external portion beak-like, the two sharp points of the "beak" with finely serrate edges; tegumen clothed with long hair, especially on its anterior edge, short fine hair on the apices of the valves.

Besides exilis, the genus Brephidium includes another Sonoran species, B. pseudofea Morrison and the S. African B. metophis.

The male genitalia of B. metophis Wallengren (Text-fig. 219) are of the same type

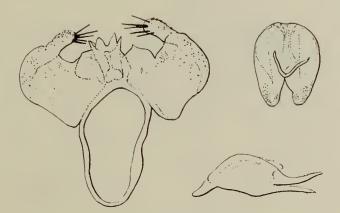


Fig. 218. Brephidium exilis (Boisduval), & genitalia.

as those of *exilis*, but the central dorsal process is more developed and is shaped like a wide blade with a scarcely denticulate distal edge; the lateral processes of the tegumen are thinner and bear at their apices two long rigid bristles; besides these the posterior edge of the tegumen bears two more short processes, strongly recurved and with apices divided into sharp-pointed teeth; the valves are pear-shaped; the penis is of the same shape as that of *exilis*.

I have also examined the male genitalia of B. pseudofea (= isopthalma Herrich Schaffer) and found it to be of the same type as in the two species mentioned above. This uniformity of structure shows the close relationship of these three species, whose venations are identical and whose external appearances are alike.

The geographical distribution of the genus *Brephidium* covers S. Africa (Cape of Good Hope, Natal and Delagoa Bay), the Sonoran region (Florida, Louisiana, New Mexico, Texas and California) and the northern part of the neotropical region (Mexico, West Indies, Central America and Venezuela). It is almost certain that this is a very ancient genus judging by the archaic structure of its male genitalia, e.g. the presence of rigid bristles and the beak-like apex of the penis, structures which elsewhere are only found in species of the genus *Zizula*, a genus also spread over the Old World and S. America.

# LIST OF SPECIES OF Brephidium

\*Brephidium metophis (Wallengren), 1860. Fig. Trimen, 1906.

#### Genus **ORAIDIUM** Bethune Baker

Oraidium Bethune Baker, 1914, Trans. ent. Soc. Lond. 47: 330; Swanepoel, 1953: 80. Typespecies: Lycaena barberae Trimen, by original designation.
Cupido Schrank (partim); Aurivillius, 1898: 369; 1925: 474.
Lycaena Fabricius (partim); Murray, 1935: 163.

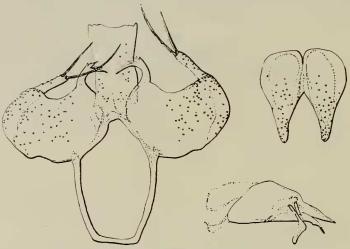


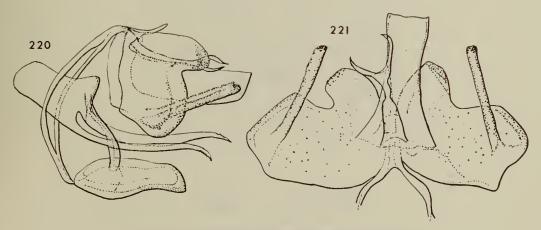
Fig. 219. Brephidium metophis (Wallengren), & genitalia.

Eyes glabrous, palpi long, ascending, second segment laterally compressed, clothed below with white scales and long black stiff hair, third segment long, slender; antennae slender, half the length of the costa, club oval, well differentiated; of fore leg, tibia with a spur and almost as long as femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing subtriangular, costa evenly convex, apex rounded, outer margin convex, hind wing oval, no tail.

Wing venation (Text-fig. 343). Bethune Baker (1914: 330) described the venation of the fore wing of barberae in these words, "Veins 6 and 7 arise from the upper apex of the cell and 7 is not stalked, 8 and 9 are absent, 11 is bent up to almost or quite touch 12". The venation of barberae is probably not stable and Bethune Baker must have examined an aberrant specimen. In the five specimens that I have examined the venation is as follows:—Fore wing with 10 veins; the cell short; 3 from before the lower angle of the cell; 4 from this angle; 5 equidistant from 6 and 4; 6 from the upper angle of the cell; 7 from rather far before this angle, ending in the apex; 8 and 9 absent; 10 from the upper edge of the cell; 11 reduced to a short cross-vein between 10 and 12.

Male genitalia (Text-fig. 220, side view; parts in situ Text-fig. 221) posteri-ventral view of the dorsal structures under higher magnification with the parts spread out and flattened (see also Bethune Baker, 1914, fig. 41). In general plant the structure is similar to that of the & genitalia of B. exilis. Directly fused to the vinculum there is a broad grooved blade stretching horizontally to the rear; above and parallel to this blade there is a process resembling a longhandled fork with two sharp-pointed prongs; lobes of tegumen very large, convex, each with a deep depression in its posterior edge, and a long digitate process on its inner surface which, instead of surrounding the external edge as in Brephidium, is directed to the rear so that its apex protrudes considerably beyond the posterior edge of the tegumen; vinculum narrow, lower fultura formed of two long slender arms which are fused to the valves, not at their base but at one-third from the base; valves small, oval, their lower margins fused throughout the first third from origin; penis very specialized, internal portion massive, saddle-shaped, external portion divided into two slender processes which diverge slightly apically where both the upper and lower edges are slightly serrate; posterior margin of tegumen clothed in long hair, especially in the middle, the digitate processes are also clothed with hair along the whole of their length and bear a pencil of long stiff hair at their apices, the lobes only sparsely clothed with short hair, distal portion of valves clothed with finer and shorter hair than that on the posterior edge and processes of the tegumen.



Figs 220-221. Oraidium barberae (Trimen), & genitalia.

The male genitalia of barberae, when examined in situ and without dissection, are confusing as the long hairs of the upper parts hide the remainder; it is for this reason probably that the presence of the "long-handled fork" escaped the notice of Bethune Baker, who would certainly have mentioned it had he seen it; his figure 41 gives a very imperfect idea of the structure of the genitalia.

The genus Oraidium is evidently closely related to Brephidium, but it is monospecific and is not found in the New World.

Desmond Murray (1935: 163) considered barberae to be a form of B. metophis in alluding to the instability of its venation. Comparison of the male genitalia of the two species renders such an hypothesis quite untenable.

## LIST OF SPECIES OF Oraidium

\*Oraidium barberae (Trimen), 1868.

# Genus ZIZEERIA Chapman

Zizeeria Chapman, 1910, Trans. ent. Soc. Lond. 43: 480; Pinhey, 1949: 121; Swanepoel, 1953: 74. Type-species: Lycaena karsandra Moore, by original designation. Cupido Schrank (partim): Aurivillius, 1898: 379; 1925: 495.

Lycaena Fabricius (partim); Murray, 1935: 183.

Eyes bearing short, very fine hair; palpi long, slightly ascending, second segment long, laterally compressed, clothed below with scales and long stiff hair, third segment shorter, slender, acuminate; antennae slender, white-annulated, more than half the length of the costa,

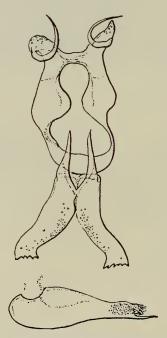


Fig. 222. Zizeeria karsandra (Moore), & genitalia.

club flattened, well differentiated; thorax clothed below with long white silky hair; legs: 3 fore leg, tibia shorter than the femur, tarsus unsegmented.

Wing shape. Fore wing subtriangular, costa evenly convex, apex rounded, outer margin convex, hind wing oval, no tail, anal angle obtuse.

Wing venation (Text-fig. 344) (see also Chapman, 1910, Trans. ent. Soc. Lond. 43, pl., 54, fig. 14): fore wing with 11 veins, 11 fused with 12 for part of its length.

Male genitalia (Text-fig. 222) (see also Chapman, 1910, Trans. ent. Soc. Lond. 43, pl. 55, fig. 20 and pl. 56, fig. 24). Uncus composed of two small lobes fused to the tegumen on either side of the median depression in its posterior edge, subunci long, slender, curved, tapering gradually to the apex which is not hooked; tegumen large, the median band strongly sclerotized on its anterior and posterior edges; vinculum broad; lower fultura formed of two robust arms fused to the base of the valves; valves fused together for the first quarter of their length, oblong, apices strongly serrate and truncate at right angles to the axis of the valves; penis shaped like an elongated flask ending in a long fine spine; vesica with numerous cornuti; uncus clothed with fine hair; thick stiff hair on the lower borders of the valves, especially near their bases, the distal portions more sparsely clothed with shorter weaker hair.

The male genitalia of knysna (Text-fig. 223) (see also Chapman, 1910, pl. 56, figs 22, 23) differ from those of karsandra solely by the shape of the valves, which are slightly wider and whose serrated apex is obliquely truncate. Steven Corbet (in litt.) considered karsandra and knysna as a collective species, karsandra inhabiting the eastern region (N.E. Arabia, Palestine, Egypt, E. Algeria and Sudan), knysna the western and southern regions (Spain, Canary Islands, Morocco, W. Algeria, S. Arabia, Sudan and tropical Africa down to S. Africa, Madagascar and Mauritius).

However the two species (or subspecies) have never been captured together and no specimens are known in which the valves have an intermediate shape.

The early stages of knysna have been described by Dickson (1944, J. ent. Soc. sth. Afr. 7:96). The caterpillar feeds on a species of Euphorbia. According to Pinhey (1945:121) in Rhodesia it feeds on Oxalis, Zornia, Medicago, Euphorbia and Amaranthus.

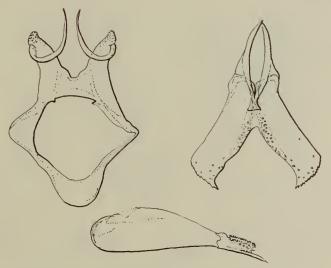


Fig. 223. Zizeeria knysna (Trimen), & genitalia

### LIST OF SPECIES OF Zizeeria

- \*Zizeeria karsandra (Moore), 1865.
- \*Zizeeria knysna (Trimen), 1862.

lysimon (Hübner), 1803, invalid homonym of lysimon (Stoll), 1790.

Zizeeria lysimon (Hübner), see knysna.

# Genus ZIZINA Chapman

Zizina Chapman, 1910, Trans. ent. Soc. Lond. 43: 482; Swanepoel, 1953: 75. Type-species:
Lycaena labradus Godart (an Indo-Australian species) by original designation.
Cupido Schrank (partim); Aurivillius, 1898: 378; 1925: 495.
Lycaena Fabricius (partim); Murray, 1935: 181.

Eyes clothed in short, very fine hair; palpi long, slightly ascending, second segment long, laterally compressed, clothed below with long stiff bristles, third segment very much shorter, slender, acuminate; antennae slender, somewhat more than half the length of the costa, club flattened, well differentiated; thorax clothed below with long white silky hair; of fore leg, tibia a little shorter than the femur, tarsus unsegmented.

Wing shape. Fore wing subtriangular, costa evenly convex, apex rounded, outer margin weakly convex, hind wing oval, no tail.

Wing venation (Text-fig. 345). Fore wing with 11 veins; 11 partially anastomosed with 12.

Male genitalia (Text-fig. 224). Uncus composed of two oval lobes fused to the tegumen on either side of the median depression in its terminal margin; subunci long, slender, curved, no apical hook; tegumen and vinculum large, lower fultura composed of two slender arms fused to the base of the valves; valves oblong, upper process much longer than the lower one and with digitate apex; penis ovoid, its extremity cylindrical with a slender sharp point; vesica with numerous cornuti; uncal lobes clothed with long fine hair, at the base of each valve there is a very long, very strong, sharp-pointed bristle and in the middle of the anterior edges of the valves there are a series of long bristles less robust than the one at the base.

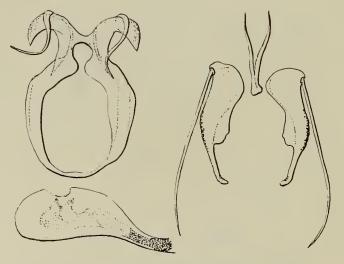


Fig. 224. Zizina labradus (Godart), & genitalia.

The male genitalia of Z. antanossa Mabille (Text-fig. 225) resemble closely those of labradus but they differ from them in the following particulars:—

- (1) The apex of the upper process of the valve is much shorter, more robust and sickle-shaped.
- (2) The basal bristle of the valve has a lanceolate apex.
- (3) The other valvular bristles are borne near the apex of the upper process and not along the middle part of the anterior edge.
- (4) The base of the penis is less swollen and the distal portion less elongated (the figures illustrate two different views of the penis, one of them with the vesica exserted after copulation).

The male genitalia of Z. antanossa are almost identical with those of Z. indica Murray, from India and Ceylon. According to Steven Corbet, antanossa of Madagascar and Africa, indica Murray, labradus Godart of Australia and oxleyi C. and R. Felder of New Zealand should be considered as forms of one collective species, otis Fabricius.

The two genera Zizeeria and Zizina are closely related and are the only genera in the subfamily Zizeerinae, which probably has a very ancient origin. This would explain its wide geographical distribution.

## LIST OF SPECIES OF Zizina

\*Zizina antanossa (Mabille), 1877. Fig. Mabille, 1885 : 87. publia (Hulstaert), 1924. Zizina publia (Hulstaert), see antanossa.

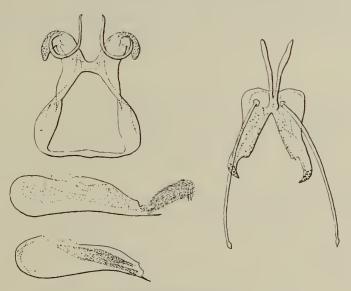


Fig. 225. Zizina antanossa (Mabille), 3 genitalia.

# Genus ZIZULA Chapman

Zizula Chapman, 1910: 483; Pinhey, 1949: 121; Swanepoel, 1958: 74. Type-species: Papilio hylax Fabricius = Lycaena gaika Trimen, by original designation.

Cupido Schrank (partim); Aurivillius, 1898: 378; 1925: 495.

Lycaena Fabricius (partim); Murray, 1935: 182.

Eyes smooth; palpi long, strongly ascending, second segment laterally compressed, clothed with scales and below with long stiff hair, third segment long, slender, acuminate; antennae white-annulated, about two-thirds the length of the costa, club ovoid, well differentiated; thorax clothed below with long white silky hair; abdomen long, extending well beyond the anal angle of the hind wing;  $\eth$  fore leg, tibia longer than the femur, tarsus short, unsegmented, strongly spinose below.

Wing shape. Fore wing subtriangular, elongate, costa evenly convex, apex rounded, outer

margin convex, hind wing oval, elongate, no tail.

Wing venation (Text-fig. 346). Fore wing with 11 veins; 11 reduced to a short cross-vein

between the upper edge of the cell and 12.

Male genitalia (Text-fig. 226) (see also Chapman, Trans, ent. Soc. Lond. 43: 493, pl. 59, figs. 33; and 35): uncus bifid, reminiscent of the Plebeiinae, the two lobes with rounded apices; subunci curved, robust, apex blunt, not hooked, tegumen large, vinculum narrow; lower fultura composed of a simple narrow blade, bent at right angles, bifid at its tip (in the mount from which the drawing was made the fultura had adhered to the penis); valves reduced, the upper process, which is folded over the lower one strongly sclerotized and ending in a long, curved digitate process crowned with a bundle of small spines, the lower process weakly sclerotized, and with a rounded apex; at the apex of the anterior edge of the valve there is an enormous long and rigid bristle, probably composed of an agglomeration of hairs because, under high magnification, it seems longitudinally channelled, and is wide at its apex, which is irregularly truncate; besides this large bristle there are 6 to 8 others, more slender and less rigid, but nearly equal in length (for the sake of the clarity only the points of insertion of these bristles are shown in the figure); penis short, massive, ending in two long beak-like points, the upper one with a smooth outer edge, and a grooved inner edge, the lower one with a slightly serrated outer edge; in situ the lower part lies in the groove of the upper part which, at its base, has on each side a small rugose triangular excrescence; uncal lobes and distal portion of the lower process of the valves pilose.

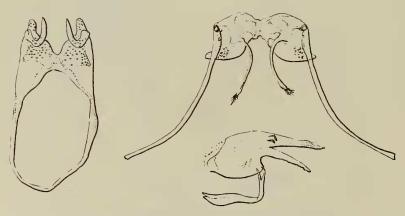


Fig. 226. Zizula hylax (Fabricius), & genitalia.

As I have already pointed out (1934, Bull. Soc. ent. Fr. 1933: 325–328) the genus Zizula includes, besides gaika, tulliola Godman & Salvin, whose male genitalia closely resemble those of gaika. Z. tulliola is found in Mexico, Guatemala, Venezuela and Brazil. Hence the habitat of the genus Zizula embraces India, Java, Sumatra, the Islands of the Indian Ocean, tropical and equatorial Africa and America, i.e. almost the whole of the ancient Gondwana.

I find it difficult to determine with certainty the exact relationship of Zizula. Like Zizina it has a huge bristle on the valve, but the tegumen and especially the penis, have a totally different structure. A penis of this type is, as far as I know, found only in Brephidium, whose habitat is also Southern Africa (metophis) and tropical America (exilis, pseudofea) but the dorsal structures of Zizula, which are of a simple type, are quite unlike the complex tegumen of Brephidium. Nevertheless the similarity in the structure of the penis, and their identical habitats, make me inclined to think that these two genera are somewhat related. Both are certainly of ancient origin, dating from before the breaking up of Gondwanza.

According to Pinhey (1949: 121) the caterpillar of gaika has been found on flowers of Oxalis species.

### LIST OF SPECIES OF Zizula

Zizula cleodora (Walker).

Zizula gaika (Trimen), see hylax.

\*Zizula hylax (Fabricius), 1775.

lysimon (Wallengren), 1857; gaika (Trimen), 1862; mylica Guenée, 1863; cleodora (Walker), 1870; pygmoea (Snellen), 1876; perparva (Saalmuller), 1889.

Zizula lysimon (Wallengren) see hylax.

Zizula mylica (Guenée), see hylax.

Zizula perparva (Saalmuller), see hylax.

Zizula pygmoea (Snellen), see hylax.

# Genus ACTIZERA Chapman

Actizera Chapman, 1910: 483; Pinhey, 1949: 121; Swanepoel, 1953: 121. Type-species: Lycaena atrigemmata Butler, selected by Hemming, 1929, Ann. Mag. nat. Hist. (10) 3: 220. Cupido Schrank (partim) Aurivillius, 1898: 379; 1925: 495. Lycaena Fabricius (partim) Murray, 1935: 183.

Eyes smooth; palpi long; ascending, second segment long, laterally compressed, clothed below with scales and long stiff hair, third segment long, slender, acuminate; antennae white-annulated, more than half the length of the costa, club ovoid, well differentiated; abdomen long, slender, extending beyond the anal angle; of fore leg, tibia as long as the femur, tarsus unsegmented, finely spinose below.

Wing shape. Fore wing subtriangular, outer margin convex, hind wing oval, no tail, anal angle obtuse.

Wing venation (Text-fig. 347). Fore wing with 11 veins, 10 and 11 free from the upper edge of the cell.

Male genitalia (Text-fig. 227) (see also Chapman, 1910, pl. 59, fig. 36 and pl. 60, fig. 39): uncus composed of two small lobes with rounded apices, tightly fused to the tegumen on either side of the shallow median depression of its posterior margin; subunci rather short, curved, about the same diameter from base to blunt apex, no terminal hook; tegumen large; lower fultura forked with two slender prongs; valves oblong, the distal portion digitate, bent upwards, apex rounded, a row of fine spines on the distal edge; valves attached below not only by their base, but also by an expansion of the upper process; penis elongate, swollen at its base, the external portion short; vesica with numerous spines; uncal lobes and lower edge of valves with long hairs.

The male genitalia of *lucida* (see Chapman, 1910, pl. 60, fig. 40) are similar to those of *atrigemmata*, differing by the serrated lower edges of the valves, the penis shorter and more swollen at its base. I have not been able to examine the type of *Zizera drucei* Bethune Baker, but I believe it to be the Madagascan race of *lucida*. A specimen from Madagascar which I dissected has an armature similar to that of *lucida*. In the genitalia of *stellata* Chapman 1910, (fig. 38) the uncal lobes are more elongate, the subunci longer and more slender, the distal portion of the valve less curved, the apical edge smooth, the penis much longer but less swollen at its base.

I do not think there is any real relationship between *Actizera* and the Zizeerinae, i.e. *Zizeeria* and *Zizina*. The only characters they have in common are their small size and a certain external similarity. The presence of a white streak on the underside of the hind wings gives no valid information as to their relationship; this character cannot have any systematic value as it varies in species of the same genus, and even in specimens of the same species, as for example, in the palaearctic genus *Agrodiaetus*.

The geographical distribution of *Actizera* (Madagascar, South and East Africa) is much less extensive than that of *Zizeeria*; maybe *Actizera* is a branch of the holarctic Glaucopsychinae in the Ethiopian fauna. Chapman included in *Actizera* the species described by Herrich Schaeffer as *Lycaena panagaea*, of Asia Minor, a species now generally assigned to the genus *Turanana* (Glaucopsychinae).

The caterpillar of A. lucida has been observed by E. C. G. Pinhey (1949: 122).

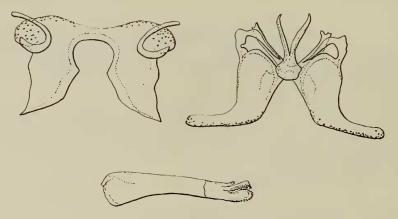


Fig. 227. Actizera atrigemmata (Butler), 3 genitalia.

#### LIST OF SPECIES OF Actizera

- \*Actizera atrigemmata (Butler), 1878. Fig. Mabille, 1885: 87.
- \*Actizera lucida lucida (Trimen), 1883. Fig. Trimen, 1906.

Actizera lucida drucei (Bethune Baker), 1906.

\*Actizera stellata (Trimen), 1883. Fig. Trimen, 1906.

#### Genus LYCAENA Fabricius

Lycaena Fabricius, 1807, Illiger. Mag. 6:285; Swanepoel, 1953:148. Type-species: Papilio phlaeas Linnaeus (a palearctic species), designated by Curtis, 1828, Brit. ent. 5: pl. 12.

Heodes Dalman: Aurivillius, 1898; 382; 1925: 497; Murray, 1953: 139.

Eyes smooth; palpi long, divergent, second segment long, laterally compressed, clothed with white scales and bearing below long black bristles, third segment more slender with obtuse apex; antennae slender, three-fifths the length of the costa, club elongate, fusiform; thorax clothed below with long white silky hair;  $\beta$  fore leg, femur clothed with long white hair, tibia shorter than the femur, tarsus unsegmented, finely spinose below, metatarsus of hind legs swollen.

Wing shape. Fore wing triangular, costa convex at its base, then straight, apex acute, outer margin slightly convex; hind wing suboval, somewhat produced at the anal angle in phlaeas but not in orus.

Wing venation (Text-fig. 348). Fore wing with 11 veins.

Male genitalia (Text-fig. 228). Uncus composed of two elongate oval lobes well separated from the tegumen, to which they are attached narrowly by their bases on either side of the median depression; subunci long, evenly curved, slightly swollen in the middle; tegumen reduced to a narrow band, vinculum rather narrow, with a tapering saccus; lower fultura composed of two lamellae obliquely truncated at their apices and fused by their bases to the base of the valves, valves oblong, widened apically with a rounded spinose apex; penis elongate,

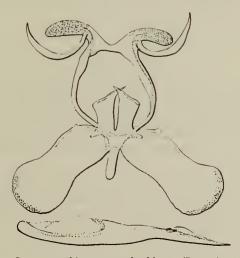


Fig. 228. Lycaena phlaeas pseudophlaeas (Lucas), of genitalia.

lower portion curved like a pistol-stock, widely open on its upper surface, apex pointed; vesica enclosing a short spine; uncus densely clothed with fine hair; just a few hairs on the upper edge of the valves near the apex.

The male genitalia of abottii (see Stempffer, 1938, Mission Omo 4:210) resemble so closely those of phleaas that it is permissible to wonder whether it is a true species or a geographical subspecies of phlaeas. Those of orus are of the same type except that the apices of the valves, instead of being rounded, are obliquely truncate and irregularly dentate, and the lower fultura consists of two narrow lamellae with pointed apices.

The biology of the European races of *phlaeas* is well known. The early stages of *orus* have been described by Murray (1935: 140, fig. 86) and Dickson (1947, *J. ent. Soc. sth. Afr.* 10: 127).

One can understand how, during the colder geographical periods, the genus *Lycaena* managed to reach South Africa by way of the Mountains of Abyssinia and East Africa. It is much more difficult to explain how it reached New Zealand, since in Asia it extends no further than the Himalayas and Sze Chuan being effectively replaced by *Heliophorus* in further Asia.

## List of Species of Lycaena

- \*Lycaena abottii (Holland), 1892. Fig. Holland, 1895. Lycaena arcas (Fabricius), see orus.
- \*Lycaena orus (Cramer), 1780.

arcas (Fabricius), 1787.

\*Lycaena phlaeas pseudophlaeas (Lucas), 1866.

phlaeas ethiopica (Poulton), 1921; phlaeas menelicki (Thierry Mieg), 1911.

Lycaena phlaeas ethiopica (Poulton), see phlaeas pseudophlaeas.

Lycaena phlaeas menelicki (Thierry Mieg), see phlaeas pseudophlaeas.

Lycaena phlaeas shima Gabriel, 1954, Exped. S.W. Arabia 1937-38: 38.

#### CONCLUSION

Now that I have set out in detail the generic characters of the Ethiopian Lycaenidae, the time has come to draw conclusions from my studies, and to attempt an outline of classification based on phylogeny, one which will, I hope, give a more accurate picture of the family than that given by the existing classification.

What are the essential characters that should form the basis of a classification? This poses the question that is at the root of the problem, because differing conclusions will result from differing choices of prime characters. Further, whatever these results may be, in terms of major taxonomic units, many of the latter will not be completely homogeneous, for not all the included species will display all the chosen characters; one can but hope to reach an approximation to a true phylogeny, because the fossil record which could prove or disaprove it, is lacking.

Aurivillius chose venation as the prime factor and I quote below the characters

he chose when he divided the Lycaenidae into sub-families (1914, in Seitz, 13: 297).

- A. Fore wing nearly always with 12 veins, of which veins 7 and 9 arise from vein 8 behind the apex of the cell.\* Rarely as in (Eresina and Iridana), vein 7 is wanting, so that only 11 veins are present, in which case either vein 10 arises from the stalk of vein 8 and 9, or vein 8 is semicircularly bent before its end. Eyes naked. Hind wing always rounded, without tail appendages, anal lobe or hairs tufts LIPTENINAE
- B. Fore wing nearly always with 10 or 11 veins, vein 7 and sometimes also vein 9 being absent; vein 10 always free from the anterior margin of the cell. Rarely (e.g. in *Aphnaeus*, *Phasis*, *Erikssonia* and the male of some species of *Iolaus*) 12 veins are present, in which case however, the hind wing is tailed, lobed or angled . . . . LYCAENINAE

Note the vagueness of the criteria: The Lipteninae have "nearly always" 12 veins, the Lycaenidae "nearly always" 10 or 11 veins. For additional distinguishing characters, Aurivillius has recourse to the pilosity of the eyes, and the shape of the hind wings. However, although it is true that all the Lipteninae have smooth eyes, this character reappears in many genera of Lycaeninae, and especially in certain Theclinae; further, many species of Lycaeninae (sensu Aurivillius) have rounded hind wings devoid of tails, lobes and hair tufts.

Aurivillius subdivided his two subfamilies into genera, usually on venational characters, sometimes on shape of wings, or the palpi or the antennae. It is undeniable that these characters are easily seen and so are convenient for the rapid determination of specimens, but in my opinion there are serious objections to the employment of venational characters as basic criteria, for example:—

- (1) At the species level, venational aberrations that are certainly of little importance occur fairly frequently in different individuals. I have given examples in *Anthene*, *Neurellipes* and *Triclema* (1944, *Revue fr. Ent.* 10:63) and I feel sure that the examination of long series of specimens would reveal their occurrence in other genera.
- (2) If we classify species into genera solely on minor venational differences we shall have to split up certain natural groups (e.g. *Iolaus*, *Phasis* and *Anthene*), of which the homogeneity on other grounds is evident.
- (3) In many cases, genera characterized by a uniform venation will include species whose male genitalia are completely dissimilar, indicating a very distant phylogenetic origin. The most striking example of such an artificial genus is *Cupido* (sensu Aurivillius) which includes species belonging to several distinct subfamilies, e.g. to Lampidinae, Plebeiinae, Zizeerinae, etc., but not a single true *Cupido*.

I agree with Warren (1947, Entomologist 80: 208 et seq.,) that we shall arrive at a more accurate classification by adopting as our basic characters (1) the shape of the legs, (2) the structure of the male genitalia. Warren makes use of the structure of the prothoracic legs to subdivide the superfamily Papilionoidea into three groups of families:—

<sup>\*</sup> An error of observation: It should read: veins 8 and 9 arise from vein 7.

- A. Primitive family group: Prothoracic legs functional in both sexes.
- B. Advanced family group: Prothoracic legs degenerate in the male, perfect in the female.
- C. Specialized family group: prothoracic legs degenerate in both sexes.

He subdivided Group A into Papilionidae, Pieridae and Lycaenidae. As characters of the last he gives: Prothoracic legs slightly smaller in male than in female; venation of fore wings specialized, veins being reduced in number (10 or 11); claws appendiculate and bifid.

This characterization of the Lycaenidae needs some qualification. It is probable that Warren was thinking solely of the holarctic fauna when he refers to "10 or 11 veins", for most of the Lipteninae, and a few genera of the Lycaeninae (sensu Aurivillius) have 12 veins in the fore wing. Also the prothoracic legs of the Lycaenidae have more than a "small" difference in size between the sexes. In almost all the genera of this family, the five segments of the fore tarsus of the male are fused to form one single segment, which is often rather slender, slightly curved, bears fine spines or short, stiff hair beneath, and has no distinct terminal claw. This seems to be the first stage of regressive evolution which culminates in the degenerate legs of the male of the Riodinidae. I think, the two families are more closely related than Warren assumes when he puts the Lycaenidae in his Group A and the Riodinidae in Group B.

As far as I know, the genera of the Lycaenidae which provide exceptions to the rule are :—Artopoetes Chapman, Coreana Tutt, Japonica Tutt, Liphyra Westwood, Euliphyra Holland, Aslauga Kirby, Paraslauga Bethune Baker, Egumbia Bethune Baker, Lachnocnema Trimen, Thestor Hübner and Theclopsis Godman & Salvin. The first three belong to the Far Eastern fauna, Liphyra is an Indo-Malayan genus and Theclopsis is a neotropical genus; as I am not sufficiently well acquainted with these genera I will say no more about them. The other six genera belong to the Ethiopian fauna, and I am of the opinion that they should be taken out of this family, since they differ from all the other genera by an extremely important character which best characterizes the family. Liphyra, Euliphyra, Aslauga and Paraslauga should form the subfamily Liphyrinae described by Bethune Baker (1924, Trans. ent. Soc. Lond. 1924: 199-238). In this subfamily, Bethune Baker also included the genus Teratoneura, but as I have pointed out in my description of that genus (p. 73), Bethune Baker must have made a mistake since the fore tarsus of the male of Teratoneura isabelle Dudgeon is quite ordinary and unsegmented. On the other hand I propose that the genus Egumbia, in which the fore tarsus of the male is five-segmented, and bears well developed terminal claws, be included in this subfamily.

When we compare the male genitalia of the four Ethiopian genera of the Liphyrinae we find the following similarities and variations:—

Tegumen:— Euliphyra: well developed, a slight notch in the posterior margin, a wide expansion of the anterior margin.

Aslauga: reduced in size, a rather deep notch in the posterior margin, a triangular expansion of the anterior margin.

Paraslauga: well developed, subtriangular, no expansion of the anterior margin.

Egumbia: fairly well developed, a rounded depression in the posterior margin, a rounded expansion of the anterior margin.

Subunci:— Long, robust and curved in *Euliphyra*, absent in the other genera. Vinculum:— Prolonged cephalad to form a large rounded saccus in all genera. Lower fultura:—Present in all four genera.

Valves:— Reduced in size in *Euliphyra*, well developed in the other genera.

Penis:— Small, elongate, subcylindrical in *Euliphyra*, more robust and enclosing numerous cornuti in the other genera.

Thus we see that, although the male genitalia are not identical, there is no radical divergence from the type. As a complementary character we may note that the fore wing always has 12 veins, of which vein 7 is stalked on 6 in *Euliphyra* but arises independently in the other genera. In my opinion the sub-family Liphyrinae is a homogeneous group, though the genus *Euliphyra*, both by its venation and by its male genitalia, seems more closely related to the Indo-Australian genus *Liphyra* than to the other three Ethiopian genera.

Of the Ethiopian genera with a segmented fore tarsus in the male, there remain Lachanocnema and Thestor, two genera that cannot be included in the Liphyrinae. Their male genitalia show at first glance one striking difference, viz: the presence in *Thestor* of two long, curved, pointed processes on the posterior margin of the uncus but, as I have already pointed out, such median expansions of the uncus have little systematic value; for instance, they recur in some species of palaearctic Theela and Chaetoprocta, which cannot for that reason alone be separated generically from allied species. Apart from this, the male genitalia of Lachnocnema and Thestor are very similar; tegumen subrectangular, subunci curved, long and rather slender, vinculum narrow and with a saccus, valves oblong with peculiar articulated processes, penis long, slightly curved and tapering gradually in its external portion; also the venation is almost identical, except that vein 7 of the fore wing arises independently in Lachnocnema, whereas it is stalked low down on vein 6 in Thestor. Finally, species of these two genera resemble one another to some extent in their outward appearance with their stout bodies and dull colours, which remind one of the Hesperidae. Bethune Baker (1924: 203) had already pointed out the close relationship of these two genera which I propose to unite in the subfamily Thestorinae.

We now have to deal with the genera in which the fore tarsus of the male is unsegmented, and these compose the greater part of the family. In order to group them into subfamilies, we shall have recourse in the first place to characters derived from their male genitalia.

In Alaena Boisduval, Telipna Aurivillius, Pentila Westwood, Ornipholidotos Bethune Baker and Liptenara Bethune Baker the ventral elements have undergone an important modification. The valves, instead of being independent organs articulated to the vinculum close to the saccus, have become mere expansions of, and more or less separated from, the vinculum, symmetrical in Alaena and Telipna, always asymmetrical in Pentila, Ornipholidotos and Liptenara. In these last three

genera the dorsal elements are also asymmetrical and subject to individual variations especially in Ornipholidotos. In Telipna and Ornipholidotos we note also the presence of paired organs articulated to the tergal-sternal suture, these being symmetrical in Telipna, asymmetrical in Ornipholidotos. In the females of Pentila and Ornipholidotos the ostium bursae is situated laterally in relation to the ostium oviductus and I suspect that the same holds good for Liptenara, but I have not had the opportunity to examine females of this genus. As a complementary character we note a certain similarity in the wing venation of the five genera: the fore wing has 12 veins, there is a short precostal vein in the hind wing and veins 3 and 4 are widely separated at their origins. In Pentila, Ornipholidotos and Liptenara the cell is much elongated and extends beyond the middle of the wing. This combination of characters seems to me to justify the grouping of these five genera into a subfamily, the Pentilinae. Such a subfamily would correspond broadly to the tribe Pentilini sensu Aurivillius. But this author, taking the presence of a precostal vein in the hind wing as the sole criterion, included in his tribe the genera Cooksonia Druce and Durbania Trimen, two genera which do not show in their male genitalia any of the distinctive characters enumerated above. A precostal vein being also found in Megalopalpus, a genus evidently far removed from the Pentilinae, I cannot accept its presence as a character of basic importance.

In another group of genera it is the tergal elements that have undergone important modifications: the tegumen is radically reduced; the uncus is composed of two large hemispherical lobes, clearly asymmetrical, fused directly to the vinculum and each armed at the apex with a long, curved, sharp-pointed process; the valves are elongate, simple in outline and apically slightly falcate; the penis is strongly curved and ends in a sharp point, its base closely wrapped in a kind of sheath borne on a pedicel attached to the base of the valves. This type of genitalia is very constant in the group, especially as regards the shape and asymmetry of the uncus, the valves differing only slightly from one species to another. I propose to erect for these two genera, *Mimacraea* and *Mimeresia*, the subfamily Mimacraeinae, (subfam. n.) although they differ in venational detail and, more particularly, in size and wing markings. *Mimacraea* more or less resembles *Bematistes*; *Mimeresia* resembles *Pseuderesia* with which it has long been confused.

Let us now consider the genera included by Aurivillius in his tribe Liptenini. We have taken out of this tribe Euliphyra, and Aslauga and Paraslauga, which we have put into the Liphyrinae, and Mimacraea, which we have put into the Mimacraeinae. On the other hand we have added Cooksonia and Durbania, withdrawn from the Pentilinae, in spite of the presence of a small precostal vein in the hind wing, because their very ordinary male genitalia in no way resemble those of the highly specialized Pentilinae. On comparing the male genitalia of the species of those genera of the Liptenini which I have examined, I found such a great variety of structure that I was able to find only one character common to them all, namely that the base of the penis is wrapped in a sheath borne on a more or less distinct pedicel attached to the base of the valves, an arrangement similar to that found in the Mimacraeinae. No doubt, there are other characters common to most of the genera, but no other common to

them all. The distribution of some of these, amongst the genera of the Lipteninae, are indicated below:—

- (I) Uncus crescentic, more or less excised along its posterior margin, surrounding a subtriangular tegumen, to which it is connected by a weakly sclerotized membrane which is translucent under the microscope: Cooksonia, Durbania, Euthecta, Larinopoda, Liptena, Micropentila (in part), Argyrocheila, Teratoneura, Iridana, Deloneura, Poultonia, Epitolina, Batelusia, Powellana, Neaveia, Phytala, Stempfferia, Epitola, Aethiopana, Neoepitola and Hewitsonia, but an uncus of this type is not found in Pseuderesia (s. str.), Citrinophila, Teriomima, Baliochila, Cnodontes, Eresinopsides, Eresina, Toxochitona and Pseudoneaveia, or at least not in all the species included in these genera.
- (2) Vinculum prolonged to form a more or less well developed saccus, which extends candad, not cephalad as is the case in most Lycaenidae. The saccus in this case is not an integral part of the vinculum, but is fused to it and is fairly easily detached in the course of dissection; it is large and triangular in Cooksonia, Durbania, Argyrocheila, Deloneura and Poultonia; more elongate in Liptena, Micropentila, Batelusia, Powellana, Neaveia, Stempfferia, Phytala, Epitola, Aethiopana and Hewitsonia; sometimes disproportionate and with a spatulate apex, e.g. in some species of Liptena and Micropentila; or even absent in some genera, e.g. Citrinophila, Baliochila etc.
- (3) The penis varies very much in shape; its external portion is gradually tapered in Cooksonia, Durbania, some Liptena, some Micropentila, Teratoneura. Iridana and Epitolina; bifid in Larinopoda; strongly S-shaped in Falcuna, Argyrocheila, some Citrinophila and some Eresina; provided with large dorsal expansions in Deloneura, Poultonia, Batelusia and some species of Epitola and Hewitsonia; disproportionately long, slender, and curved in Toxochitona; its apical part circinate in several species of Micropentila.
- (4) Finally there is the remarkable structure in *Baliochila* and *Eresinopsoides bichroma* Strand, that deserves mention. This consists of two processes with weakly sclerotized bases which are fused to the tegumen, and to the dorsal side of the penis, and bear either apical hooks or many stiff erect bristles along the whole of their length.

I have not succeeded in dividing the Liptenini (sensu Aurivillius) into homogeneous subfamilies, each having a distinctive type of male genitalia. On the other hand, in the present state of our knowledge, with the male genitalia of many species still not examined, the erection of a large number of subfamilies as an alternative solution, seems undesirable. I think that here we are dealing with many different lines of descent, some of which are very ancient and, in the course of a long evolution, have undergone a complete modification of their original external appearance. What can have been the factors which caused some of these species to resemble, in wing colour and markings, Eurema or other Pieridae? This is not the place to start a discussion on the causes and evolution of mimicry, but to note its existence. In the course of evolution there may have been convergent lines of development, which tended towards not only similarity of external appearances but even similarity of venation,

for, after all, the number of possible combinations of the veins is limited. Some of these fortuitous groups of species have up to now being treated as true genera, e.g. Teriomina, (sensu Aurivillius) Citrinophila and Eresina, but their male genitalia have retained, at least in part, their ancestral characters and their study reveals both the different origins of the species comprising the group, and the artificiality of these so-called genera. One may hope that when all the species of the Liptenini have been methodically examined, it may be possible to trace their phylogenetic relationships, and to group them in a natural classification. For the present however, I prefer to admit defeat rather than yield to a love of symmetry by forming arbitrary groups, whose systematic value would be no improvement on the present classification. Hence I propose to retain provisionally the division of the subfamily into two sections as done by Aurivillius, based on the relative positions of the origins of veins 6 and 7 of the fore wing, i.e. stalked or very close together, in the first group, far apart in the second group.

### Section 1.

Fore wing veins 6 and 7 stalked or very close together at point of origin.

Durbania Trimen. Fore wing 12 veins, vein 6 and the stem of veins 7+8+9 have separate origins; 11 arising nearer to 10 than to the base; hind wing short precostal vein present; 7 not stalked on 6; costa strongly convex at its base; 3 and 4 far apart.

Durbaniella van Son. Fore wing with vein II arising equidistant between 10 and base; hind wing with short precostal vein, cell about half as long as wing.

- Durbaniopsis van Son. Fore wing with vein II arising much nearer to vein IO than to base; hind wing with short precostal vein, cell much shorter than in Durbaniella, about one-third as long as wing.
- Pseuderesia Butler. Fore wing (P. eleaza) with 12 veins, vein 6 stalked on 7+8+9, 10 from the upper angle of cell in 3, stalked on 7+8+9 in Q; or (P. isca group) 10 from upper angle of cell; hind wing with 7 stalked on 6, 5 nearer to 6 than to 4, 3 and 4 far apart at origin.

Citrinophila Kirby. Fore wing with 12 veins; 6 and 7+8+9 have a common stem; 5 nearer to 6 than to 4; hind wing, 7 branched on 6, 5 nearer to 6 than

4; 3 and 4 far apart.

- Teriomima Kirby, Baliochila Stempffer & Bennett, Cnodontes Stempffer & Bennett, Euthecta Bennett. Fore wing with 12 veins; 6 and 7+8+9 have a common stem; 5 nearer to 6 than to 4; hind wing, 7 free; 5 nearer to 6 than to 4; 3 and 4 rather close together.
- Larinopoda Butler. Fore wing with 12 veins; 6 separated from 7+8+9; cell produced at the lower angle; hind wing, 7 free; cell short produced at the lower angle; 3 and 4 far apart; 5 nearer to 6 than to 4.
- Liptena Hewitson, Falcuna Stempffer & Bennett. Fore wing with 12 veins; 6 separated from 7+8+9; cell rather short, 5 nearer to 6 than to 4; hind wing, 7 free; 3 and 4 slightly separated, connate, or slightly stalked.

- Micropentila Aurivillius. Venation as on Liptena; differs from Liptena only in shape of palpi and antennae.
- Eresinopsides Strand. Fore wing with 11 veins; 6 and 7+9 have a common stem; hind wing, 7 stalked on 6; 3 and 4 connate.

  Eresina Aurivillius. Fore wing with II veins; 7+8 and 10 stalked on 6; hind
- wing 7 stalked on 6; 3 and 4 have separate origins.
- Toxochitona Stempffer. Fore wing with 11 veins, 7+8 stalked on 6; 10 from upper angle of cell. Hind wing with 7 stalked on 6; 6 and 7 widely separate.

  Argyrocheila Staudinger. Fore wing with 12 veins; 6 stalked on 7+8+9; outer
- margin deeply scalloped (in *undifera*); hind wing, 7 stalked on 6; 3 and 4 far apart.
- Teratoneura Dudgeon. Fore wing with 12 veins; 6 and stalk of 7+8+9 separate; 2 and 3 curved towards the inner margin; outer margin excised; hind wing, 7 free; 3 and 4 shortly stalked; costa concave.
- Iridana Aurivillius. Fore wing with 11 veins; 6 and 7 arise from the upper angle of the cell; 7 doubly curved near the apex; hind wing, 7 free; 3 and 4 connate.

### Section 2.

- Fore wing veins 6 and 7 widely separated at origin *Cooksonia* Druce. Fore wing with 12 veins; 6 widely separated from 7+8+9; 5 much nearer to 6 than to 4; hind wing, short precostal vein present; 7 free; 3 and 4 separate.
- Deloneura Trimen. Fore wing with 12 veins; 6 widely separated from 7+8+9; 5 and 6 arise from the upper angle of the cell; hind wing, cell short, truncate; 7 free; 3 and 4 slightly separate at their origins.

  Poultonia Neave. Fore wing with 12 veins; 6 widely separated from 7+8+9;
- 5 very close to 6; hind wing, 7 free; 5 a little nearer to 6 than to 4; 3 and 4 connate.
- Batelusia Druce. Fore wing with II veins; 6 widely separated from 7+8; 5 very close to 6; hind wing, 7 free; 5 nearer to 6 than to 4; 3 and 4 connate.
- Tumerepes Aurivillius. Fore wing with 12 veins; 6 widely separated from 7+8+9; 5 very close to 6; hind wing, 7 free; 5 nearer to 6 than to 4; 3 and 4 on a short common stem.
- Neaveia Druce. Fore wing with 12 veins; 6 separate from 7+8+9; 5 nearer to 6 than to 4; hind wing, 7 free; 5 nearer to 6 than to 5; 3 and 4 separate. Pseudoneaveia Stempffer. Venation as in Neaveia. Genitalia very different.
- Epitolina Aurivillius. Fore wing with 12 veins; 6 and 7+8+9 separate; hind wing, 7 free, 3 and 4 connate.
- Stempfferia Jackson. Fore wing with 12 veins; 6 and 7+8+9 separate; 5 nearer to 6 than to 4; 10 (in the male) shortly stalked on 7, 11 and 7 connate; 10 (in the female) connate with 7, and 11 slightly before 7; hind wing with 7 free, 3 and 4 short stalked.
- Phytala Westwood. Fore wing with 12 veins; 6 widely separated from 7+8+9:

5 nearer to 6 than to 4; II and I2 confluent; hind wing, 7 free, 3 and 4 on a short common stem.

Epitola Westwood. Fore wing with 12 veins; 6 separate from 7+8+9; 5 nearer to 6 than to 4; 11 and 12 free; hind wing, 7 free; 3 and 4 shortly stalked. Aethiopana Bethune Baker. Fore wing with 12 veins; 6 widely separated from

7+8+9+10; II and I2 free; hind wing, 7 free; 3 and 4 on a common stem. Neoepitola Jackson. Fore wing with 12 veins; 6 widely separated from 7+8+9; II stalked on IO; hind wing with 7 free, 3 and 4 on a common short stem.

Hewitsonia Kirby. Fore wing with 12 veins; 6 and 7+8+9 separate; 5 nearer to 6 than to 4; hind wing, 7 free; 3 and 4 on a common stem.

Powellana Bethune Baker. Fore wing with 12 veins; 6 widely separated from 7+8+9; 5 nearer to 6 than to 4; 11 stalked on 10; hind wing 7 free; 3 and 4 connate.

I propose to restrict the name Lipteninae to Section 1, and to adopt the name Epitolinae, a name already employed by several authors, for Section 2, which is much more homogeneous than Section 1, from the point of view of the male genitalia. In Section 2 (except in *Pseudoneaveia*) the uncus is crescentic, sometimes notched at the apex, the subunci are robust, curved, except in *Epitola hewitsoni* which lacks them completely, the valves have a smooth uniform outline, are elongate with convex upper edge, a nearly straight lower edge and a rounded apex. The penis bears a wide expansion on its dorsal surface in *Deloneura*, *Poultonia* and *Batelusia*, and also in some species of *Epitola* and *Phytala*.

In the Lycaeninae (sensu Aurivillius) we start with the genus Megalopalpus Rober, all the characters of which, specialized male genitalia, large palpi and venation, show its close relationship to the Indo-Malayan Gerydus and Allotinus. Its proper place is therefore in the subfamily Miletinae (= Gerydinae) as Aurivillius and Bethune Baker had already pointed out.

We now come to the extensive subfamily Theclinae, which is cosmopolitan and forms the bulk of the rich neotropical Lycaenid fauna. I propose to include in it the following genera; Deudorix Hewitson, Hypomyrina Druce, Actis Karsch, Kopelates H. H. Druce, Hypokopelates H. H. Druce, Pilodeudorix H. H. Druce, Diopetes Karsch, Virachola Moore, Myrina Fabricius, Oxylides Hübner, Syrmoptera Karsch, Hypolycaena Felder, Hemiolaus Aurivillius, Stugeta H. H. Druce, Pseudiolaus Riley, Trichiolaus Aurivillius, Dapidodigma Karsch, Tanuetheira H. H. Druce, Argiolaus H. H. Druce, Iolaphilus Stempffer & Bennett, Philiolaus Stempffer & Bennett, Iolaus Hübner, Aphniolaus H. H. Druce, Epamera H. H. Druce, Etesiolaus Stempffer & Bennett, Sukidion H. H. Druce, Leptomyrina Butler, and Capys Hübner.

Except in a few instances, with which I shall deal later on, the male genitalia of the above genera have the following characters: tegumen large, hood-shaped with a more or less deep rounded median depression in the posterior margin; uncus composed of two small lobes fused to the tegumen on either side of this depression, (but it is difficult to make out the suture by a superficial examination, so the lobes may then appear to be mere thickenings of the tegumen less translucid posteriorly). Very seldom the two uncal lobes fuse with one another so that the median depression

disappears, e.g. in some species of *Epamera*; subunci long, robust, usually curved, sometimes bearing apophyses; vinculum wide above, rather narrow below, the saccus generally small or absent; lower fultura absent in many genera (e.g. *Deudorix* and closely allied genera, *Capys*, *Hypolycaena* and *Leptomyrina*), ringshaped and encircling the penis, sometimes carried on pedicel in some genera of the *Iolaus* groups; the valves, reduced in size in *Deudorix* and the closely allied genera, *Capys*, *Oxylides*, *Syrmoptera*, *Hypolycaena* and *Leptomyrina*, are more fully developed in the genera of the *Iolaus* group; penis widely open on the dorsal surface of its internal portion, usually weakly curved, and often enclosing a voluminous cuneus or large spines.

It is remarkable that the male genitalia of the Theclinae are often extremely similar in species which outwardly are very different. This is particularly the case in the *Deudorix* group. The relatively simple pattern of the genitalia of this subfamily suggests that the Theclinae are of recent origin, in spite of their wide geographical distribution.

The species whose male genitalia differ from the typical structure just described belong to the genera and subgenera *Myrina*, *Dapidodigma*, *Argiolaus*, *Philiolaus* and *Etesiolaus*. Having already given detailed accounts of their male genitalia under the relevant genera there is no need to repeat them here. It is only necessary to mention that in all these it is the tergal parts that are modified, the sternal elements remaining typically Thecline.

It is fairly easy to pick out groups of closely allied genera in the Ethiopian Theclinae. The genera *Hypomyrina*, *Actis*, *Kopelates*, *Hypokopelates*, *Pilodeudorix*, *Diopetes* and *Virachola* are scarcely more than subgenera of *Deudorix*, *Capys* is closely allied to them. *Myrina* is rather isolated; *Syrmoptera* can hardly be separated from *Oxylides*; *Leptomyrina* is closely allied to *Hypolycaena*; *Dapidodigma* is isolated, but *Hemiolaus*, *Stugeta*, *Pseudiolaus*, etc. up to and including *Sukidion* can be regarded as belonging to this large *Iolaus* group.

Continuing the examination of the genera in the order adopted by Aurivillius, I propose to unite in the subfamily Aphnaeinae Swinhoe (1911, Lep. Indica 9: 156) the following genera:—Aphnaeus Hübner, Paraphnaeus Thierry Mieg, Apharitis Riley, Spindasis Wallengren, Lipaphnaeus Aurivillius, Choloroselas Butler, Zeritis Aurivillius, Desmolycaena Trimen, Axiocerses Hübner, Phasis Hübner, Alocides Hübner, Poecilmitis Butler, Chrysoritis Butler, Crudaria Wallengren, Erikssonia Trimen and Pseudaletis Druce.

The adoption of this grouping is justified on the basis of one striking character in the structure of the male genitalia, which is present in all the species I have examined, namely the semimembranous band connecting the median parts of the upper processes of the valves by passing and closely sheathing the penis. There is a trace of this structure in some species of *Actis* and *Kopelates*, in which the valves are also connected in their middle, but in these two genera, closely allied to *Deudorix*, it is less well developed and the lower fultura is absent, whereas the latter is always present in the Aphnaeinae, usually in the shape of a notched shield.

In Aphnaeus and Paraphnaeus the dorsal elements are subrectangular and hood-

shaped, the subunci long, robust, and curved, the two processes of the valves are separated at their apices, and the penis is very massive, and bears spines and numerous cornuti. In Apharitis and in most species of Spindasis the dorsal elements are divided into two lobes by a deep median depression in the posterior margin of the tegumen and the valves are not apically divided. In Lipaphnaeus, Chloroselas and Desmolycaena the dorsal structure form an oval, which is excised at the hind apex, the subunci long and slender (except in D. rogersi Riley in which the subrectangular shape reappears and the subunci are absent). In Zeritis the dorsal aspect is subtriangular, the anterior margin of the tegumen bears a rounded process, the subunci have a long apophysis and the lower fultura is composed of two long asymmetrical arms. In Axiocerses the dorsal aspect rather resembles that of Zeritis but the arms of the lower fultura are symmetrical. In Phasis, Aloeides, Poecilmitis and Chrysoritis the dorsal elements are more or less rectangular and always bear a rounded process on the anterior margin. The subunci are always long, and robust and bear a small apophysis. In *Crudaria* on the other hand the subunci are reduced in size. In Erikssonia the dorsal aspect resembles that of Axiocerses, but the subunci have no apophysis. I do not know Pseudaletis well, having examined only six species, in all of which the uncus is divided into four processes (in P. agrippina Druce the apex of each process is blunt, in P. clymneus Druce the apices are pointed) and lacks subunci. But the lower fultura, valves and penis are of Aphnaeine type, for which reason I include Pseudaletis in this subfamily, though with some misgiving. Except for Pseudaletis, all the genera of this subfamily have moreover one striking external character, namely the presence of metallic silvery golden or nacreous bands and spots on the underside of the wings. The palaearctic genus Cigaritis Boisduval, a close ally of Apharitis, should be included in this subfamily.

We must now consider the genus *Spalgis* Moore, the precise affinities of which I find difficult to decide. In his generic description, Moore says that it is allied to *Gerydus* (Seitz, 1923, *Gross-Schmett. Erde* 9:881). Fruhstorfer compares the wing markings of *Spalgis* with those of *Lycaenopsis* and their dull colour with that of the *Gerydinae*; as regards the male genitalia he says that the long furca resembles that of *Tarucus*, and that the uncus resembles that of *Everes*. He concludes that *Spalgis* is probably a very ancient group, possibly the stock from which both *Gerydus* and *Everes* were derived.

Colour and wing pattern are not very important characters in my opinion. The palpi of *Spalgis* have no resemblance whatever to those of *Gerydus*, the short stout antennae are unlike those of *Gerydus*, *Everes* or *Lycaenopsis*, and the genitalia in no way resemble those of *Lycaenopsis* or *Gerydus*. On the other hand the pentagonal uncus and short stout subunci are very much Everine in type. I conclude that, on the evidence available, the wisest course is to leave the genus *Spalgis* isolated in the subfamily Spalginae, as other authors have done.

The next group comprises the genera Cupidesthes Aurivillius, Anthene Doubleday, Neurypexina Bethune Baker, Neurellipes Bethune Baker and Triclema Karsch, which differ only in minor venational characters. Their male genitalia are so uniform in type that I think they ought to be considered merely as subgenera of

Anthene. The genitalia may be described as follows: Uncus composed of two small lobes fused to the tegumen on either side of the median depression in its posterior margin, subunci curved, sometimes robust, sometimes slender; tegumen a rather narrow notched band; vinculum always prolonged towards sternite 8 to form a saccus which is often well developed; lower fultura a furca with divergent arms and attached to the base of the valves, which are oval and more or less incised; penis elongate and widely open on the dorsal surface of its internal portion.

In external appearance the *Anthene* group bear some affinity to the Theclinae, but the male genitalia are intermediate between those of the Theclinae and the Lampidinae. However, the invariable presence of a furca and the poor development of the tegumen lead me to place the group in the Lampidinae, where they form a well marked tribe.

I would also include in the Lampidinae the following genera taken from the "omnibus" genus Cupido of Aurivillius:—Phlyaria Karsch, Uranothauma Butler, Cacyreus Butler, Tarucus Moore, Castalius Hübner, Lampides Hübner, Syntarucus Butler, Cyclyrius Butler, Harpendyreus Heron, Pseudonacaduba Stempffer, Lepidochrysops Hedicke, Euchrysops Butler, Eicochrysops Bethune Baker, Cupidopsis Karsch, Thermoniphas Karsch, Oboronia Karsch, Athysanota Karsch and Actizera Chapman. I must confess that it is not without some hesitation that I group together all these genera because the male genitalia are not of a perfectly uniform type. For instance, the male genitalia of Tarucus, Castalius and Eicochrysops differ in some particulars from the common plan, which may be summarized as follows:—Tegumen with a median depression in its posterior margin, sometimes reduced to a narrow band; uncus divided into two lobes; well developed subunci, lower fultura furca-shaped, and sometimes with an anellus; penis subcylindrical. Some pairs of genera, e.g. Phlyaria and Uranothauna, Syntarucus and Cyclyrius have a similar specialized penis, and in this show close relationship to the American Leptotes; others, e.g. Lepidochrysops, Euchrysops, Thermoniphas, Oboronia and Athysanota from a group characterized by their reduced tegumen, their digitate valves and the presence of an anellus.

As regards *Tarucus* with tectorius and virgae excitatae present in most species, *Castalius* which is only an artificial grouping of species with convergent external appearance, and *Eicochrysops* with its four-lobed dorsum, all three genera differ considerably from the typical Lampidinae and if I have included them in this subfamily it is solely because I could not find a more suitable place and they seemed more allied to the Lampidinae than to any other subfamily. To *Actizera*, which has some of the characteristics of the Glaucopsychinae, I give only a temporary home in the Lampidinae. It seems probable that in the future this subfamily will need subdivision into several homogeneous groups, but before that can be done a more thorough study of the Indo-Australian species on the basis of their male genitalia is needed.

There is no doubt that *Chilades* Moore and *Freyeria* Courvoissier should be included in the subfamily Plebeiinae, a subfamily not well represented in the Ethiopian and Indo-Malayan faunas but which includes many holarctic species.

Azanus Moore is characterized by its undivided uncus, which is more or less excised at its apex, its short robust subunci and its reduced tegumen, features typical of the sub-family Everinae, which it also resembles in venation viz. fore wing with II veins, with II and I2 confluent.

Although the genera *Brephidium* Scudder and *Oraidium* Bethune Baker include very few species I think they should be placed in a subfamily to themselves, the *Brephidinae*. Their male genitalia are of such a peculiar structure and differ so markedly from those of all other genera of Lycaenidae that they cannot be included in any of the other subfamilies.

I propose to restrict the subfamily Zizeerinae to the two genera Zizeeria Chapman and Zizina Chapman. It is characterized by an uncus divided into two slender lobes, slender curving subunci long slender curved valves with long stiff bristles, and a flask-shaped penis, with pointed apex and a vesica bearing numerous spines. Chapman included in the subfamily Zizeeriinae the genera Actizera Chapman and Zizula Chapman. I have above (p. 275) stated my reasons for taking Actizera out of the subfamily Zizeeriinae. As for Zizula their male genitalia have one feature found in the Zizeeriinae, namely the stiff valvular bristles, but they have also one feature in common with the Brephidiinae, namely a penis shaped like a bird's beak. The Zizeeriinae are found in the Indo-Malayan, Ethiopian and Palaearctic zones, the Brephidiinae in the Ethiopian, neotropical and Sonoran zones, the species of Zizula in Indo-Malayan, Ethiopian and neotropical zones. There is probably some relationship between these archaic groups which probably originated before the breaking up of Gondwanaland.

There remains only the genus Lycaena Fabricius, which has very few species in the Ethiopian fauna, but is well represented in the holarctic zone. It comprises the "Coppers" so familiar to British entomologists and must be included in a separate subfamily. As to the name to be given to this subfamily, I hesitate between Lycaeninae and Heodinae; although the former is the correct name in accordance with the Rules of Nomenclature, there is a risk that it might be confused with Lycaeninae, sensu Aurivillius, by those entomologists who are not specialists.

It is not difficult to foresee that criticisms will be levelled at my conclusions, indeed I am well aware of some that can be made. It is abundantly clear that most of the proposed subfamilies, namely the Liphyrinae, Thestorinae, Pentilinae, Mimacraeinae, Miletinae, Aphnaeinae, Plebeiinae, Everinae, Brephidinae, Zizeerinae and Lycaeninae are based on well defined structural characters. To distinguish the Lipteninae from the Epitolinae or the Lampidinae from the Theclinae is much more difficult. Why, one might ask, split up the African Lycaenid fauna into so many fragments when eight subfamilies suffer for the whole of the palaearctic fauna? Why erect a subfamily to include only two genera each with only a few species? Why propose a modification of the generally accepted sequence of genera when, on my own admission, the reasons for this are debatable.

The answer is that the African fauna, at any rate as far as the Lycaenidae are concerned, is far richer and much more varied than that of the palaearctic region. During the Tertiary period, Europe was subjected to so many large scale changes of

climate that its original fauna must have been replaced, at least partially on several occasions, the species present there now having been derived mostly from recent angarian immigrants. In Africa, on the other hand where the climate remained more stable, there are still ancient gondwanian elements which have continued to evolve there, or have come in from Indo-Australian regions via Arabia. It is this that has produced the astonishing diversity both of types and of highly specialized genera. Some of these archaic groups include only a small number of species that have, however, an exceedingly wide distribution, they are nevertheless very distinct, and it would be quite arbitrary to lump them, systematically, with a group of more recent and more numerous species. Classification can only take facts into account, it cannot be subservient to geometrical patterns. It cannot, for example, like an army commander divide its taxa into units of fixed size, like regiments, battalions and companies.

I could have limited the scope of this work to a bare compilation and the statement of facts. Instead, I have attempted to revise and bring up to date the systematics of the African Lycaenidae by the use of characters accepted in the study of the faunas of other regions. This first attempt on these lines cannot but be imperfect and, to some extent, hypothetical. I have not glossed over my doubts, hesitations and setbacks, but have called attention to them, believing it wiser so to do than to attempt a dogmatic solution in spite of them. The preparation of this work has occupied much of my time and has entailed much labour, I am fully satisfied if it attracts fruitful criticism and provides basis for a more comprehensive and detailed investigation.

#### A POSTSCRIPT

Mr. Harry K. Clench, of the Carnegie Museum, Pittsburgh, U.S.A. has been so very kind as to send me a copy of the volume on the Butterflies of Liberia, (1965, Mem. Am. ent. Soc. 19), to which he contributed the section on the Lycaenidae. It is most unfortunate that this should have been published at a moment when it was no longer possible for me to incorporate the important results of his research in the body of the present work. This has obliged me to review, in the form of a supplementary note, his ideas on the subject of the classification of the African Lycaenidae. And I find it a little disconcerting to have to disagree, on several matters of importance, with the views of a valued correspondent of long standing.

As a starting point Clench erects the superfamily Lycaenoidea, divided into four families: Liptenidae, Liphyridae, Lycaenidae and Riodinidae. I will not deal with the last of these as it is outside the scope of the present work.

If the Liphyridae are excluded from the Lycaenidae because of the segmented fore tarsi of the males of the former, it would be equally logical to exclude the Thestorinae from the Lycaenidae, because they too have segmented fore tarsi, a character which I regard as of primary importance.

Clench then subdivides each of the families on the basis of a variety of characters, without according consistent primacy to any single character. For example, he

uses turn and turn about the morphology of the fore tarsi of the male, the presence of a precostal vein on the hind wing, the arrangement of the small spines on the underside of the male fore tarsi, the relative lengths of the femur and tibia of the hind leg, the wing venation in general, the presence or absence of coremata, and even at times the food of the larvae. On the other hand he seems to attribute only a secondary degree of importance to the form of the male genitalia.

I find myself unable to agree with this method, for I believe it essential, in order to achieve a coherent system, to establish a kind of hierarchy—arbitrary perhaps, amongst characters, and to follow this throughout a family, only using characters of secondary importance in a supplementary sense. To me it does not seem reasonable, for example, to attach the same taxonomic importance to the spines on the fore leg as to the genitalia of the male.

However, so as not unduly to prolong this note, I propose to indicate below only those points upon which I do not find myself in agreement with Clench.

# Page 269:—Key to the subfamilies of the Liptenidae:

- Ventral spines of fused male fore tarsus in two dissimilar rows, a mesad row of erect,
  rather closely and evenly spaced spines and an exterior row of sparse, declivent,
  irregular spaced spines, hind leg with femur subequal to or slightly exceeding tibia

  Durbaniinae (extralimital)

I have already explained (p. 268) my reasons for excluding *Durbania*, and the allied recently erected genera, from the Pentilinae, because their genital armatures are simple and symmetric. As to the genus *Thestor* (which Clench does not mention again, since it is restricted to South Africa), I consider it has nothing to do with the Lipteninae, the fore tarsi of the male being segmented.

# Page 269:—Subfamily Pentilinae.

I see no objection to the generic treatment adopted by Clench. Indeed I fully support his creation of the genus *Ptelina* for *carnuta* Hewitson, having already indicated, following Bethune Baker, that this species should be eliminated from the genus *Telipna*. I would point out, however, that the male genitalia of *carnuta* are clearly analogous to those of *Alaena*, particularly in respect of its dorsal elements and the presence of subunci.

# Page 281:—Key to the Tribes of Lipteninae.

For ewing  $R_{3-5}$  arises distinctly before upper angle of cell, well separated from origin of M; male with coremata eversible ventral sacs bearing long, modified scales, the sacs just anterior to genital capsule . . . . . . . . . . . . . Epitolini

	R <sub>3_5</sub> arise from upp	er angle of	cell,	close t	to or	connate	with	(occa	asiona	lly lo	ng st	alked	
	with $M_1$ ) .											. 2	
2	Male with coremata	(as above)										Iridanini	
	No coremata .											Liptenini	

The presence of coremata in many species has been well known to me for many years. I make no mention of them in the present work for the following reasons:—

- (I) Within a single genus there may be some species with coremata, others without.
- (2) In the course of dissection in water, after maceration in potash, it often happens that, on applying pressure to the abdomen to force the extrusion of the genitalia, the coremata remain within the eighth segment, and escape detection through being mixed up with other abdominal debris. This depends I believe, in some degree upon the degree of decomposition sustained by the organs after death, in a hot and humid climate, and also perhaps upon the strength of the potash solution used.
- (3) In the process of mounting in Canada balsam, when pressure is put upon the cover slip, the coremata often break away from the vinculum and the scales are dispersed throughout the preparation and become almost invisible; for this reason I have hitherto not included the coremata in my preparations.

These technical details may seem trivial, and out of place. Forty years experience, however, has taught me that they are of importance. It is only too easy to draw false conclusions from a faulty mount.

Wishing to check the taxonomic importance of the coremata, in the sense in which Clench uses them, I have made a small series of fresh preparations of species already examined, paying particular attention to preserving the coremata, with the following results.

# Coremata present

# Coremata absent

# Lipteninae

Liptena o-rubrum Holland
Liptena modesta Kirby
Liptena despecta Holland
Liptena catalina Smith & Kirby
Micropentila brunnea Kirby
Micropentila dorothea Bethune Baker
Micropentila ugandae Hawker Smith
Micropentila bunyoro Stempffer & Bennett
Micropentila mpigi Stempffer & Bennett
Micropentila jacksoni Talbot
Micropentila cherereti Stempffer & Bennett
Teratoneura isabellae Dudgeon
Iridana unyoro Stempffer

Liptena opara immaculata Grünberg Liptena flavicans Smith & Kirby Liptena homeyeri Dewitz Liptena ideoides Dewitz Micropentila adelgitha Hewitson Argyrocheila undifera Staudinger Eresina rougeoti Stempffer Coremata present

Coremata absent

*Epitolinae* 

Phytala elais Doubleday Hewitson Phytala hyeltonides Aurivillius Stempfferia carcassoni Jackson Aethiopana honorius Fabricius Epitola ceraunia Hewitson Hewitsonia boisduvali Hewitson Poultonia ochrascens Neave Powellana cottoni Bethune Baker Epitolina dispar Kirby Epitolina catori Bethune Baker

There is no point in pursuing this matter further. The lists above clearly demonstrate that the presence or absence of coremata does not provide a character of valid generic significance, and even less of tribal value. In order to separate the Lipteninae from the Epitolinae we are forced back on the venational characters of Aurivillius' system. In fact, as I have already indicated, the two subfamilies are not sharply definable, *Iridana* providing a transition from one to the other.

Clench includes the Mimacraeinae within his Liptenini. I must confess that I do not understand why he fails to accord them subfamily rank, for they constitute a perfectly homogeneous, very specialized group in which the male genitalia are so uniform that they are rarely even of use in separating species. I cannot accept Pseuderesia eleaza in any way as providing a transition in genitalia characters between Mimeresia and the Lipteninae, in Mimeresia as in Mimacraea the uncus is formed of two very large semicircular asymmetric lobes, each bearing a long tapering similarly asymmetric horn on its terminal margin and the tegumen is much reduced. The genitalia indeed are without any exception asymmetric in every species. On the other hand in Pseuderesia eleaza, although admittedly the tegumen is equally reduced, the uncus is formed of two long perfectly symmetrical foliaceous processes, and there are neither semicircular lobes nor accessory horns; I freely admit that in eleaza the uncus is more deeply divided and the tegumen narrower than in the other species of pseuderesia, in consequence of which it occupies a somewhat isolated position in the genus, but I do not see any necessity to erect, for the other species of Pseuderesia, the new genus Eresiomera based otherwise on the shape of the fore wing and the pattern of the markings on the hind wings. If the shape of the wings is taken as a generic criterion, Epitola should be subdivided into at least three genera according to whether the four wings are falcate (posthumus), dentate (hewitsoni) or slightly convex (cercene). To accept wing pattern as a generic character is to return to the mistakes of earlier authors who put Azanus isis in Castalius and Euchysops crawshayi and most of the species of Harpendyreus in Cyclyrius.

In the same way, I see no merit in restricting the genus *Phytala* to its type species *P. elais*, and erecting for all the other species the new genus *Hypophytala*, on the basis of the large size of *elais* and the much smaller expanse of all the other species of *Phytala*, and the presence of a protuberance on the dorsal surface of the penis in *hyettoides*, which is lacking in *elais*. In my view this protuberance is not a good generic character, since it is to be found in many other Epitolinae, for example, *Deloneura millari* Trimen, *Poultonia ochrasceus* Neave, *Batelusia zebra* H. H. Druce, *Epitola posthumus* Fabricius, *Epitola urania* Kirby and others.

Page 320 :—Family Liphyridae.

Clench unites in this family the subfamilies Liphyrinae and Gerydinae and the Indo-Malaysian Poritiinae. The last is not well known to me, but the association of the other two seems debatable, since they differ fundamentally in their morphology. In the Liphyrinae the male fore tarsus is segmented and the genitalia differ little from those of the Lycaenidae. In the Gerydinae the fore tarsi are unsegmented, and in *Gerydus*, *Allotinus* and *Megalopalpus* the hind wing has a precostal vein, and the very specialized genitalia are different from those of the Lycaenidae.

Page 321:—Subfamily Liphyrinae.

The author divides this subfamily into two tribes, the Liphyrinae and the Lachnocnemini. The former corresponds to my subfamily Liphyrinae; but the presence here of *Lachnocnema*, which apart from the segmented fore tarsi, bears no resemblance whatever to the Liphyrinae, is a matter of surprise. I consider *Lachnocenema* to be much more closely related to *Thestor*, which Clench exiles to the Liptenidae.

Page 325 :—Subfamily Gerydinae.

Divided into two tribes, Gerydini and Spalgini. Here again I cannot accept this association. The Gerydini (or Miletinae) have a precostal vein on the hind wing and very specialized male genitalia; Spalgis is utterly dissimilar, having no precostal vein and genitalia of a quite common-place Everinae type. Clench draws attention to the fact that in both groups the larvae are carnivorous. I doubt the advisability of using this character, for nothing is known of the larval habits of 80% of the African species. Moreover, such information as is available suggests that this carnivorous habit is not absolutely constant. Some phytophagous species do not disdain honey dew, and the exudates of various Homoptera. Others spend part of their larval life in ant's nests or in shelters constructed by ants. What happens within the ant's nests? Do the larvae subsist on the regurgitated food of the ants, or do they sometimes devour the ant larvae or pupae? I do not know. In the course of rearing the common European Black Hairstreak butterfly in captivity, I noticed that this normally phytophagous larva sometimes becomes a ferocious cannibal during the last instar, even when amply provided with its normal food. I fancy this seldom happens in nature, since these slow-moving larvae live widely dispersed. There is here a clear indication of a carnivorous trend in the Lycaenidae; larvae of other families deprived of their natural food supply die of starvation without recourse to cannibalism.

Page 331 :—Lycaenidae

Page 332 :—Theclinae

Page 357 :—Aphnaeinae

We are in agreement here. No comment is necessary.

Page 364 :—Subfamily Plebeiinae

Divided by Clench into Lampidini, Everini, Plebejini and Zizeerini.

In my view, all these tribes merit to rank as equal subfamilies. The Lampidini occur throughout the Ethiopian and Indo-Malaysian regions and even reach the Neotropical region, but very few penetrate into the holarctic zone. It is probably a very ancient subfamily, for the male genitalia exhibit wide variety. When the Lycaenid fauna of the tropics is fully known, I expect it will prove necessary to divide the group into several subfamilies. The Everinae are mainly holarctic and include only a few genera with relatively little differences in their male genitalia. The Plebejinae are certainly a recent subfamily of Angarian origin. They account for the great bulk of the holarctic "Blues", only two genera, *Freyeria* and *Chilades* having penetrated the tropics of the Old World. The male genitalia throughout the Plebejinae are of a very uniform pattern, with the uncus divided into two more or less subtriangular lobes, and curved subunci. The Zizeeriinae are a primitive subfamily with a very wide distribution in spite of the small number of species.

Their male genitalia often exhibit archaic characters. The subfamily Plebejinae as conceived by Clench appears to me to be decidedly heterogeneous.

I will conclude by repeating that I make no claim to a monopoly of the truth, on the contrary, I have been at pains to show how precarious some of my conclusions seem to be, and I am naturally glad to change my opinions in favour of others that would seem better justified. But I should be lacking in scientific honesty if I failed to indicate, however briefly, my reasons for retaining provisionally the system of classification which I first elaborated some twenty years ago.

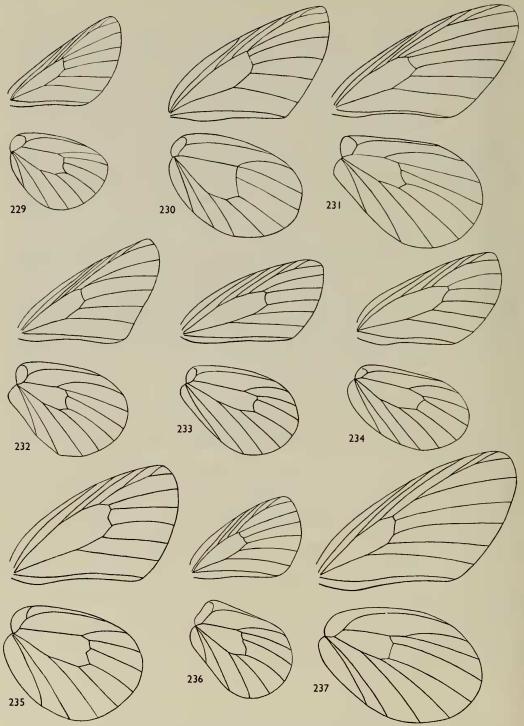
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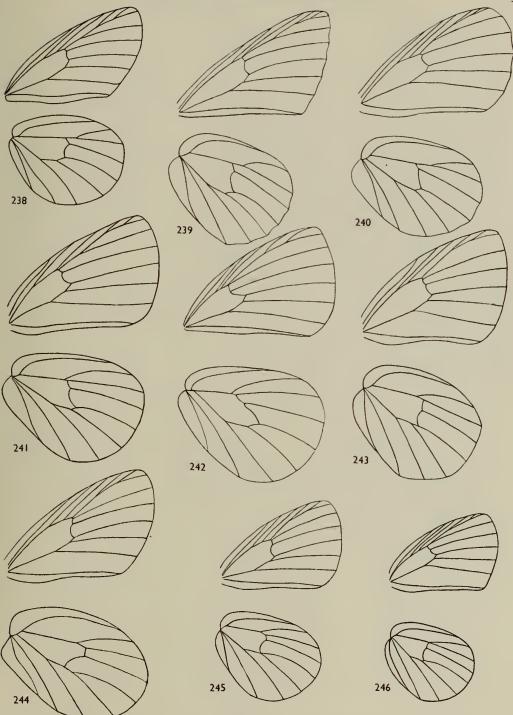
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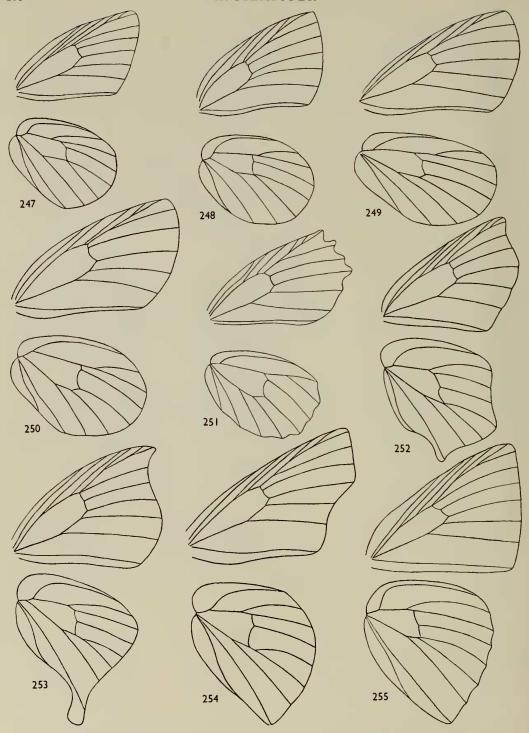
Full references are given in the text only to works not listed above or not covered by references to be found in Aurivillius (1898, 1919–1925).



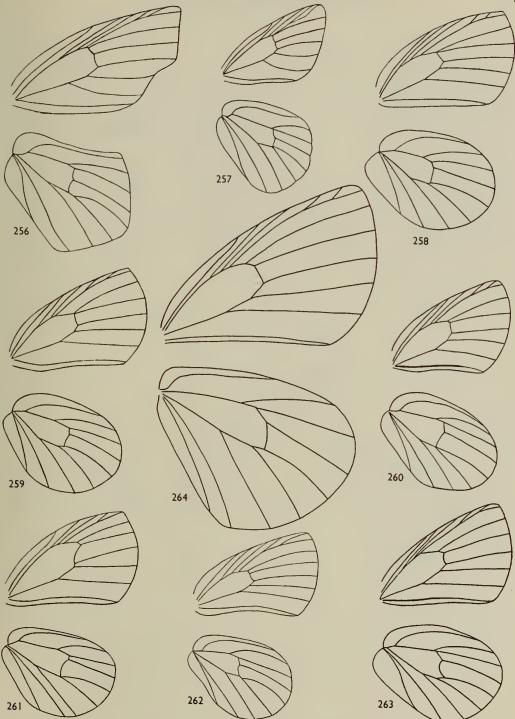
Figs. 229–237. Wing venation of :—229, Alaena amazoula Boisduval; 230, Telipna bimacula semirufa (Smith & Kirby); 231, Cooksonia trimeni Druce; 232, Cooksonia neavei (Druce); 233, Pentila tropicalis tropicalis (Boisduval); 234, Liptenara batesi Bethune Baker; 235, Ornipholidotos kirbyi (Aurivillius); 236, Durbania amakosa Trimen; 237, Mimacraea darwinii Butler.



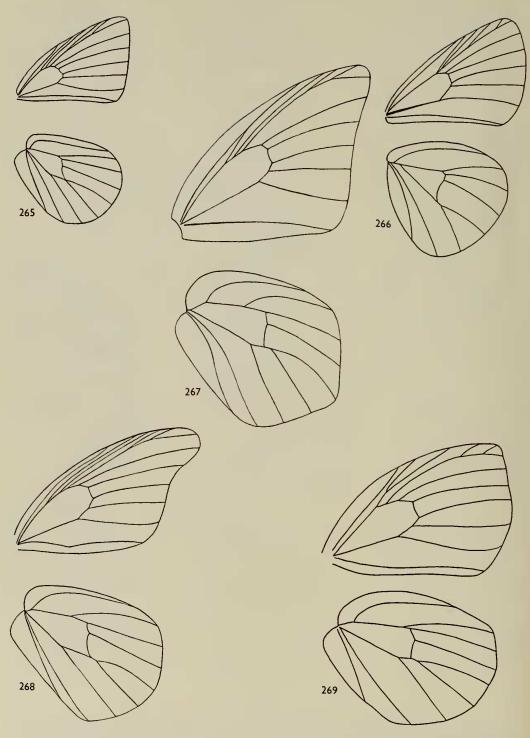
Figs. 238-246. Wing venation of:—238, Mimeresia libentina (Hewitson); 239, Pseuderesia eleaza eleaza (Hewitson); 240, Citrinophila marginalis Kirby; 241, Teromima subpunctata Kirby; 242, Larinopoda lircaea (Hewitson); 243, Falcuna libyssa libyssa (Hewitson); 244, Liptena (Liptena) undularis Hewitson; 245, Liptena (Liptena) decipiens (Kirby); 246, Liptena (Tetrarhanis) ilma daltoni Poulton.



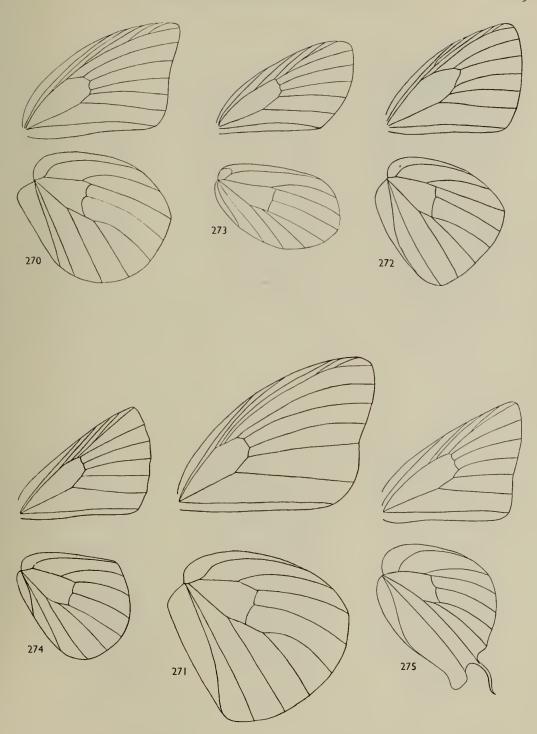
Figs. 247–255. Wing venation of:—247, Micropentila adelgitha (Hewitson); 248, Eresinopsides bichroma Strand; 249, Eresina corynetes (Smith & Kirby); 250, Toxochitona gerda (Kirby); 251, Argyrocheila undifera Staudinger; 252, Aslauga vininga (Hewitson); 253, Paraslauga kallimoides (Schultze); 254, Euliphyra mirifica Holland; 255, Egumbia ernesti (Karsch).



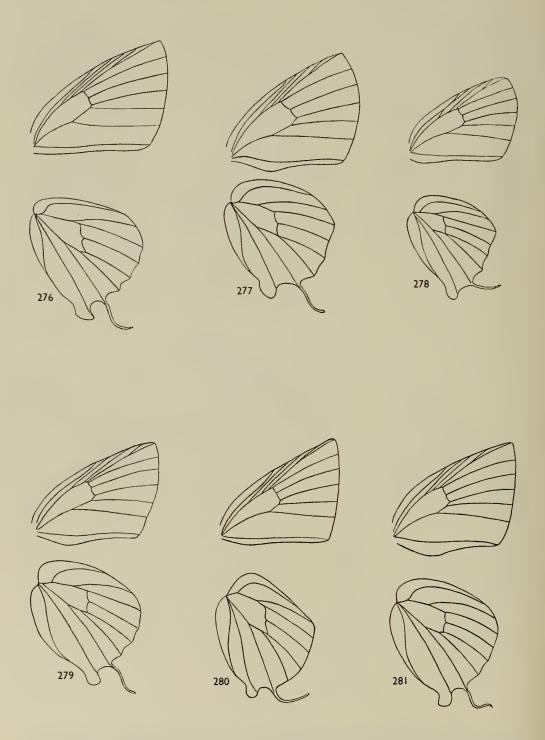
Figs. 256-264. Wing venation of:—256, Teratoneura isabellae Dudgeon; 257, Iridana incredibilis (Staudinger); 258, Deloneura immaculata Trimen; 259, Deloneura millari Trimen; 260, Ebepius ochrascens (Neave); 261, Batelusia zebra Druce; 262, Tumere-pedes flava Bethune Baker; 263, Neaveia lamborni Druce; 264, Pseudoneaveia jacksoni Stempffer.



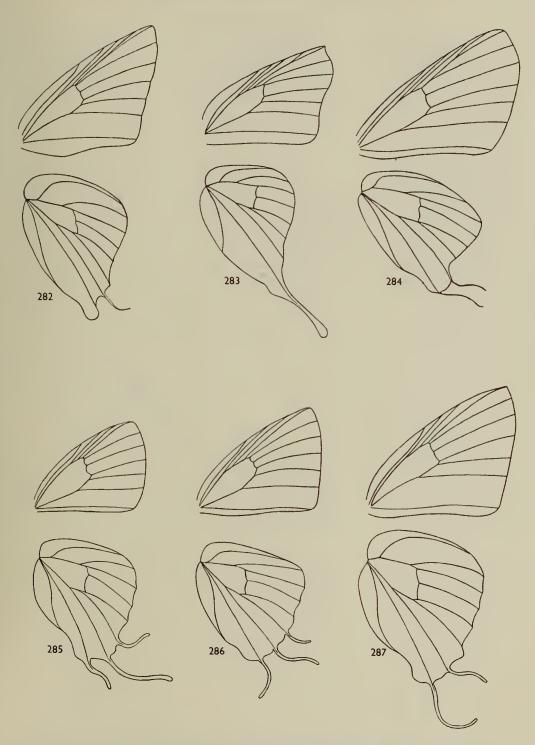
Figs. 265–269. Wing venation of :—265, Epitolina dispar (Kirby): 266, Stempfferia carcassoni Jackson; 267, Phytala elais elais Westwood; 268, Epitola posthumus (Fabricius); 269, Neoepitola barombiensis (Kirby).



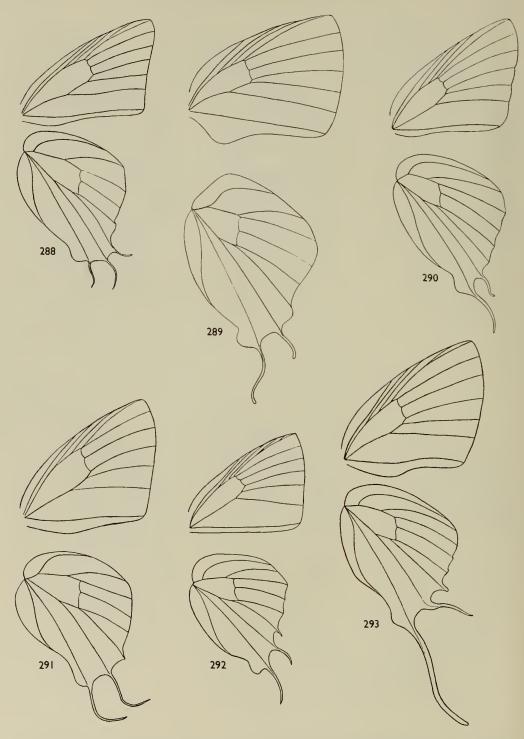
Figs. 270–275. Wing venation of :—270, Aethiopana honorius honorius (Fabricius); 271, Hewitsonia boisduvali boisduvali (Hewitson); 272, Powellana cottoni Bethune Baker; 273, Megalopalpus simplex Röber; 274, Lachnocnema bibulbus (Fabricius); 275, Deudorix epijarbas (Moore).



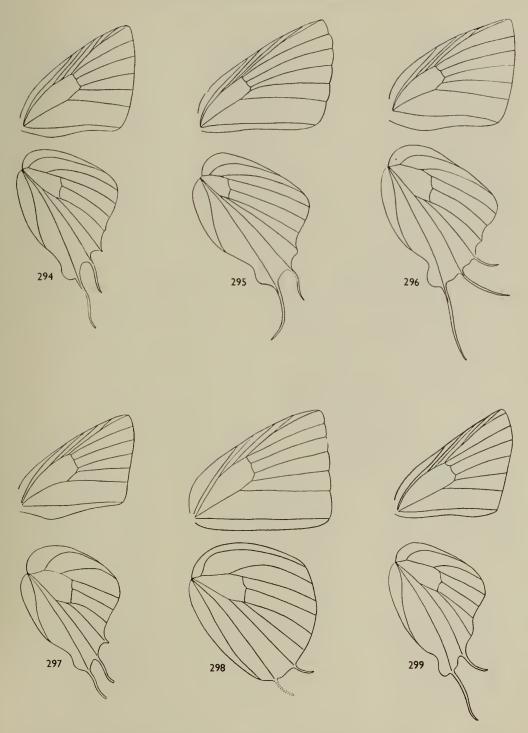
Figs. 276–281. Wing venation of :—276, Hypomyrina nomenia nomion (Staudinger); 277, Actis mimeta mimeta Karsch; 278, Kopelates virgata Druce; 279, Hypokopelates mera (Hewitson); 280, Pilodeudorix camerona (Plötz); 281, Diopetes deritas (Hewitson).



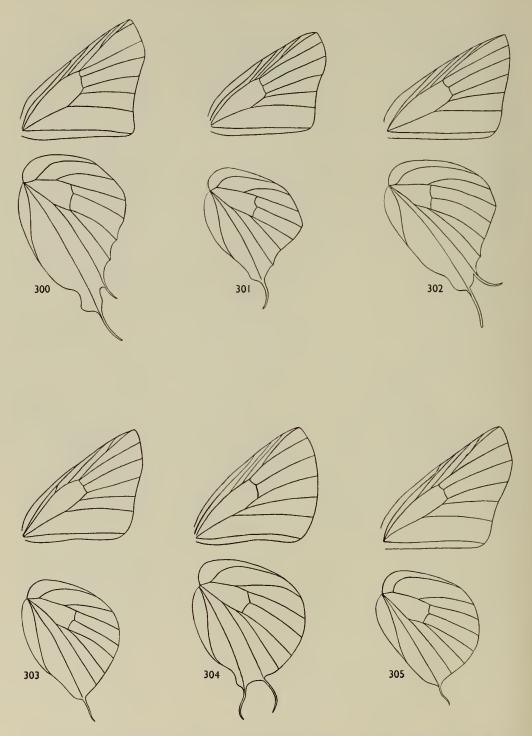
Figs. 282–287. Wing venation of :—282, Virachola perse (Hewitson); 283, Myrina silenus ficedula Trimen; 284, Pseudaletis agrippina Druce; 285, Oxylides faunus (Drury); 286, Syrmoptera melanomitra Karsch; 287, Hypolycaena sipylus (Felder).



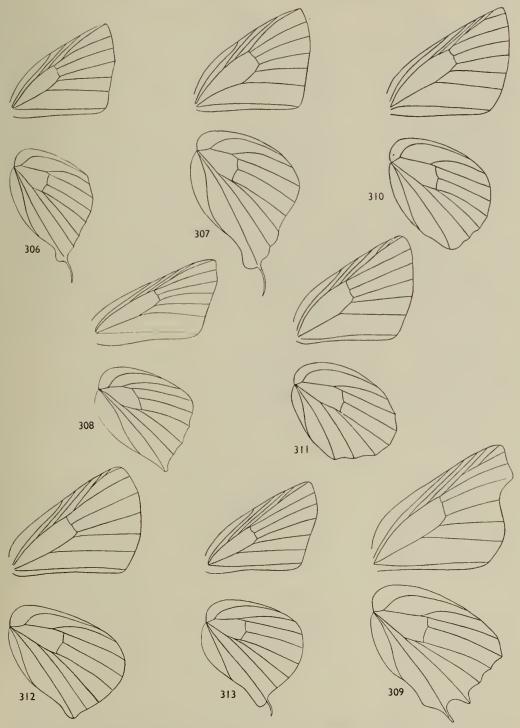
Figs. 288–293. Wing venation of :—288, Dapidodigma hymen (Fabricius); 289, Iolaus (Hemiolaus) coeculus coeculus Hopffer; 290, Iolaus (Stugeta) bowkeri bowkeri Trimen; 291, Iolaus (Pseudiolaus) poultoni Riley; 292, Iolaus (Trichiolaus) mermeros (Mabille); 293, Iolaus (Tanuethira) timon orientius Hulstaert.



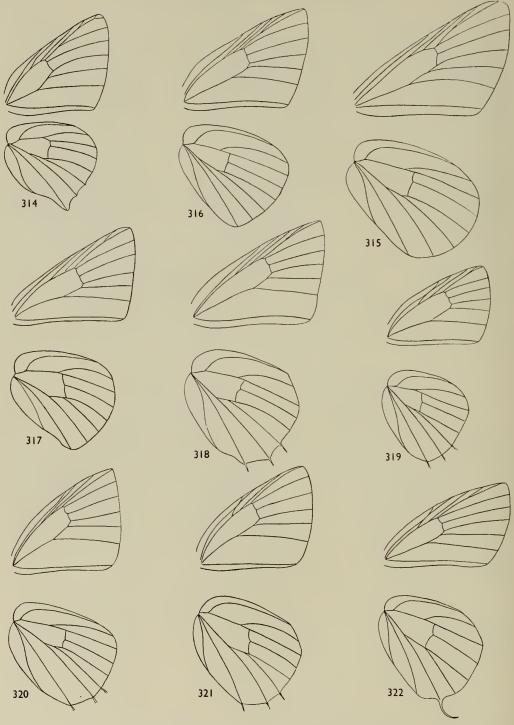
FIGS. 294–299. Wing venation of :—294, Iolaus (Argiolaus) silas silas Westwood; 295, Iolaus (Aphniolaus) pallene (Wallengren); 296, Iolaus (Iolaus) eurisus (Cramer); 297, Iolaus (Epamera) sidus Trimen; 298, Iolaus (Sukidion) inores Hewitson; 299, Aphnaeus oreas oreas (Drury).



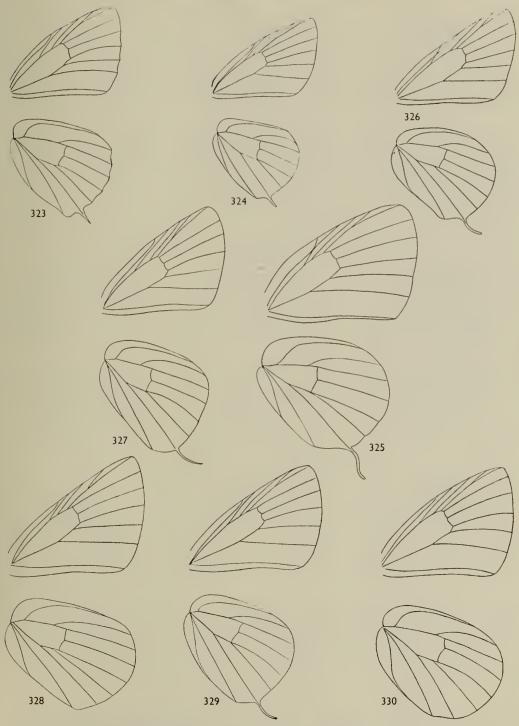
Figs. 300-305. Wing venation of :—300, Paraphnaeus hutchinsoni hutchinsoni (Trimen); 301, Apharitis epargyros (Eversman); 302, Spindasis natalensis (Doubleday & Hewitson); 303, Chloroselas tamaniba esmeralda Butler; 304, Zeritis neriene neriene Boisduval; 305, Desmolycaena mazoensis Trimen.



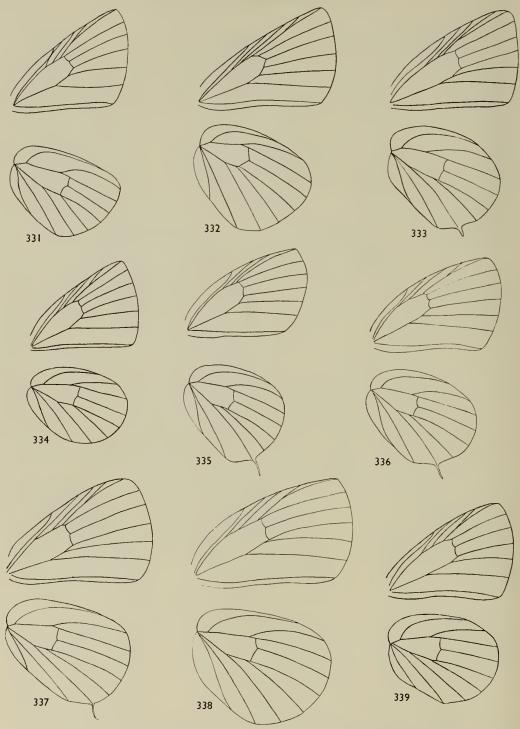
Figs. 306-313. Wing venation of:—306, Axiocerses harpax harpax (Fabricius); 307, Leptomyrina (Leptomyrina) phidias (Fabricius); 308, Capys alphaeus (Cramer); 309, Phasis thero thero (Linnaeus); 310, Aloeides pierus (Cramer); 311, Poecilmitis lycogenes (Trimen); 312, Chrysoritis oreas (Trimen); 313, Crudaria leroma leroma (Wallengren).



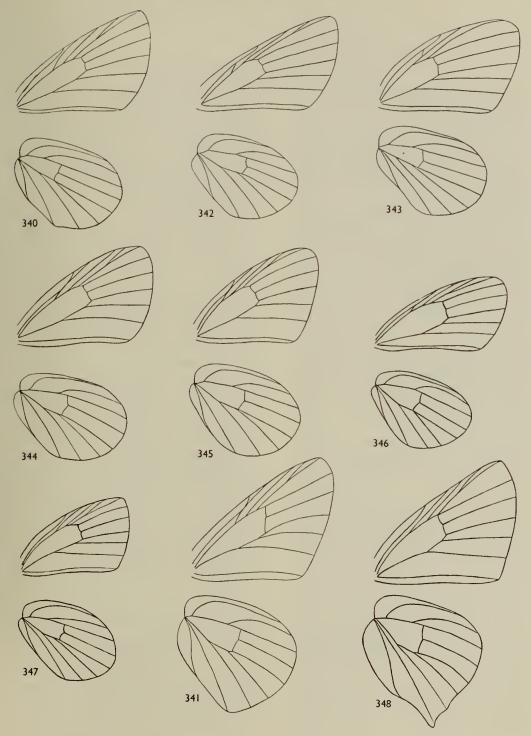
FIGS. 314–322. Wing venation of:—314, Erikssonia acraeina Trimen; 315, Thestor protumnus protumnus (Linnaeus); 316, Spalgis epius (Westwood); 317, Cupidesthes robusta Aurivillius; 318, Anthene larydas larydas (Cramer); 319, Neurypexina lyzanius (Hewitson); 320, Neurellipes lusones (Hewitson); 321, Triclema lucretilis lucretilis (Hewitson); 322, Phlyaria cyara cyara (Hewitson).



Figs. 323–330. Wing venation of :—323, Uranothauma crawshayi Butler; 324, Cacyreus lingeus lingeus (Cramer); 325, Castalius rosimon (Fabricius); 326, Tarucus theophrastus (Fabricius); 327, Lampides boeticus (Linnaeus); 328, Cyclyrius webbianus (Brullé); 329, Syntarucus pirithous pirithous (Linnaeus); 330, Harpendyreus reginaldi Heron.



Figs. 331-339. Wing venation of:—331, Pseudonacaduba aethiops (Mabille); 332, Lepidochrysops parsimon parsimon (Fabricius); 333, Euchrysops cnejus (Fabricius); 334, Eicochrysops eicotrochilus Bethune Baker; 335, Cupidopsis iobates iobates (Hopffer); 336, Thermoniphas plurilimbata plurilimbata Karsch; 337, Oboronia punctatus punctatus (Dewitz); 338, Athysanota ornata ornata (Mabille); 339, Chilades laius (Cramer).



Figs. 340–348. Wing venation of :—340, Freyeria trochylus (Freyer); 341, Azanus ubaldus (Cramer); 324, Brephidium exilis (Boisduval); 343, Oraidium barberae (Trimen); 344, Zizeeria karsandra (Moore); 345, Zizina labradus (Godart); 346, Zizula hylax (Fabricius); 347 Actizera atrigemmata (Butler); 348, Lycaena phlaeas pseudophlaeas (Lucas).

## Synonyms in italics

aberrans, f. of Cupidopsis cissus 240 abottii, Lycaena 264 abraxas, Pentila 17 abruptus, Anthene 190 abyssiniae, subsp. of Euchrysops mauensis abyssiniae, subsp. of Euchrysops nandanus 236 abyssinica, Euthrysops 235 abyssiniensis, subsp. of Lepidochrysops parsimon 232 acares, Hypomyrina 100 acholi, Lepidochrysops 228 acraea, Telipna 11 acraeina, Erikssonia 181 acraeoides, Telipna 11 actinotina, Telipna 111 Actis 101 Actizera 261 adamsi, Aphnaeus 150 adamsi, Iolaus (Epamera) 144 adelgitha, Micropentila 60 adelgunda, Micropentila 60 aderna, Lipaphnaeus 161 adherbal, Anthene 190 adolphifrederici, Epitola 87 adonis, Poecilmitis 177 aelianus, Iolaus (Iolaphilus) 133 aemulus, Iolaus (Epamera) 144 aequatorialis, subsp. of Epitola staudingeri 90 aequatorialis, Harpendyreus 224 aequatorialis, Iolaus (Iolaphilus) 133 aequatorialis, subsp. of Liptena flavicans 54 aequatorialis, Neurellipes 197 aequatorialis, Phytala 85 Aethiopana 91 aethiopia, Lepidochrysops 228 aethiops, Pseudonacaduba 227 aethon, Poecilmitis 177 aethria, (Audeoud) Iolaus (Epamera) 144 aethria, (Karsch) Iolaus (Epamera) 144 affinis, Aphnaeus 150 affixa, Pentila 17 afra, Anthene 190 africana, Triclema 199 agave, Azanus 252 aglaspis, Aloeides 175

agnes, Iolaus (Epamera) 144 agripina, Pseudaletis 114 Alaena 8 alba, Epitola 87 alba, Pentila 17 albata, subsp. of Oxylides faunus 115 alberici, Iolaus 144 alberici, Mimeresia 32 alberici, Thermoniphas 243 alberta, Anthene 190 alberta, Chilades 248 alberta, subsp. of Pentila nyassana 18 alberta, Micropentila 60 albescens, f. of Uranothauma falkensteini albicans, subsp. of Anthene sylvanus 194 albicans, f. of Lepidochrysops parsimon 232 albicans, Liptena (Liptena) 53 albicans, f. of Uranothauma antinorii 202 albicosta, Oborona 245 albida, Cupidesthes 187 albida, subsp. of Pentila cloetensis 17 albifascia, ab. of Hewitsonia boisduvali 94 albifrons, f. of Virachola lorisona 110 albilinea, Lepidochrysops 228 albilunata, f. of Anthene crawshayi 190 albipicta, subsp. of Triclema lucretilis 199 albiradiatus, f. of Cupidopsis cissus 240 albistriatus, Euchrysops 235 albocaerula, Thermoniphas 243 albocaerulea, subsp. of Iolaus (Epamera) pollux 147 albofasciata, subsp. of Telipna bimacula 12 albomacula, Liptena (Liptena) 53 albomaculata, subsp. of Crudaria leroma 180 albomaculata, Epitola 87 albomaculatus, subsp. of Iolaus (Epamera) iasis 146 albula, Liptena (Liptena) 53 alcibiades, Iolaus (Iolaphilus) 133 alcides, Myrina 112 alciopina, subsp. of Mimacraea neurata 31 alenica, ab. of Larinopoda lircaea 45 alenicola, ab. of Larinopoda lircaea 45 aliciae, Cooksonia 14 alienus, Iolaus (Epamera) 145 aliquantum, subsp. of Liptena (Liptena) subvariegata 55

alluaudi, Liptena (Liptena) 53 Apharitis 155 almeida, Aloeides 175 Aloeides 173 Aphnaeus 150 alphaeus, Capys 172 alticola, subsp. of Iolaus (Epamera) silanus apuleia, Spindasis 148 alticola, Virachola 109 amabilis, Durbania 27 amabilis, Liptena (Liptena) 53 amakosa, Durbania 27 amanga, Axiocerses 168 aranda, Aloeides 175. amanica, subsp. of Baliochila minima 43 amanica, Hypolycaena 119 arcas, Lycaena 264 amanica, subsp. of Iolaus (Epamera) nolaensis 147 amarah, Anthene 190 amasa, Syrmoptera 117 amazoula, Alaena 9 amenaida, Pentila 17 amenaidena, subsp. of Pentila nyassana 18 amenaidoides, Pentila 17 amine, subsp. of Zeritis nereine 164 ammon, Epitola 87 anadema, subsp. of Anthene rubricinctus 194 Argyrochiela 65 analogramma, Castalius 211 androgyna, f. of Syrmoptera nivea 117 anerius, subsp. of Lepidochrysops parsimon arnoldi, Anthene 190 anesius, Iolaus (Iolaphilus) 133 anetia, Hypokopelates 103 anetta, Hypokopelates 103 anettae, Myrina 112 Arrugia 182 angelita, Hypokopelates 103 angolensis, subsp. of Falcuna libyssa 47 artemides, Azanus 252 angulosus, Megalopalpus 97 angustata, Mimacraea 30 ashiri, Mimeresia 32 angustifascia, Telipna 11 Aslauga 67 angustimargo, Pseudaletis 114 aslauga, Baliochila 42 ankoleensis, Pilodeudorix 105 anneckei, Telipna 11 anomale, ab. of Hypolycaena lebona 120 ansorgei, Iridana 75 ansorgei, Lepidochrysops 228 anta, Virachola 109 antalus, Virachola 109 Athysanota 245 antanossa, Zizina 259 Anthene 188 antifaunus, Hypolycaena 119 atrinervis, Telipna 12 antimachus, Pseudaletis 114 aucta, Diopetes 107 antinorii, Oranothauma 202 antoto, Eicochrysops 238 auga, Pentila 17 anubis, Euchrysops 235 apatosa, f. of Iolaus (Epamera) aemulus 144 augusta, Liptena (Liptena) 53 apelles, Spindasis 158 aura, Aslauga 69

aphnaeoides, Iolaus (Epamera) 145 Aphniolaus, subg. of Iolaus 137 apicalis, Mimacraea 30 aquilonis, subsp. of Anthene lunulata 192 arabica, Desmolycaena 166 arabicus, subsp. of Anthene butleri 190 arabicus, Lepidochrysops 229 arborifera, Iolaus (Epamera) 145 arctimargo, subsp. of Oboronia punctatus arescopa, Cupidesthes 187 argentarius, Iolaus (Trichiolaus) 128 argentea, Chloroselas 163 argenteola, f. of Aphnaeus asterius 153 argenteomaculata, Axiocerses 168 argenteostriata, Harpendyreus 224 Argiolaus, subg. of Iolaus 129 argyraspis, Phasis 173 argyrocyclus, Aphnaeus 150 ariadne, Lepidochrysops 229 aridus, Poecilmitis 177 aridus, subsp. of Thestor protumnus 184 arnoldi, subsp. of Anthene leptines 192 arrhon, Pseudaletis 114 artemenes, Uranothauma 203 aruma, Hypokopelates 103 asopus, Euchrysops 235 aspasia, subsp. of Pentila cloetensis 17 aspidos, Larinopoda 45 asteris, Euchrysops 235 asteris, Lepidochrysops 229 asterius, Aphnaeus 153 atlantica, Poecilmitis 177 atrigemmata, Actizera 263 audeoudi, Cacyreus 205 auga congoensis, Pentila 17

barnesi, f. of Spindasis ella 159

aurantiaca, Alaena 9 auratus, Uranothauma 203 aurea, Anthene 190 aureobrunnea, subsp. of Anthene butleri 190 aureolineatus, Hypolycaena 119 auricostalis, Hypolycaena 119 aurivilli, Iolaus (Epamera) 145 aurivillii, Telipna 12 aurivillii, Zeritis 164 aurivilliusi, Diopetes 107 aurora, subsp. of Iolaus (Iolaus) bolissus 139 australis, subsp. of Euchrysops osiris 236 australis, Iolaus (Epamera) 145 australis, Lepidochrysops 229 avriko, Spindasis 158 Axiocerses 166 ayresi, subsp. of Durbania amakosa 27 Azanus 250 azurea, Chloroselas 163 azurea, Epitola 87 azurea, Hypokopelates 103 azureus, subsp. of Iolaus (Iolaus) bolissus 139 azureus, Lepidochrysops 229

baboulti, Syntarucus 221 bacchus, Lepidochrysops 229 bacotae, Ornipholidotos 25 badhami, Lepidochrysops 229 badhami, Virachola 109 badia, Epitola 87 baghirmi, Spindasis 158 bakeri, Anthene 190 bakeri, Iolaus (Epamera) 145 bakeriana, Liptena (Liptena) 53 bakotae, Micropentila 60 Baliochila 41 balkanicus, Tarucus 216 bambana, Axiocerses 168 bamendanus, subsp. of Capys disjunctus 172 banco, Iolaus (Epamera) 145 bansana, Iolaus (Epamera) 145 banyoana, Spindasis 158 barbara, subsp. of Iolaus (Epamera) hemicyanus 146 barbatus, Pilodeudorix 105 barberae, Oraidium 256 barca, Deloneura, 77 barkeri, Euchrysops 235 barkleyi, Aloeides 175 barnesi, Anthene 190 barnesi, subsp. of Aphnaeus erikssoni 153 barnesi, Baliochila 42 barnesi, Lepidochrysops 229

barnsi, subsp. of Iolaus (Epamera) hemicyanus 146 barnsi, subsp. of Mimeresia debora 32 barombiensis, Neoepitola 91 baronica, Virachola 109 bassae, Liptena (Liptena) 53 basuta, Thestor 183 Batelusia 79 batesana, Liptena (Liptena) 53 batesi, Epitola 87 batesi, Larinopoda 45 batesi, Liptenara 21 batesi, Pseudaletis 114 batikeli, Virachola 109 batikelides, Virachola 109 baumi, Axiocerses 168 beaufortia, Poecilmitis 177 bella, Aslauga 69 bella, Epitola 87 bella, Oxylides 115 bellatrix, subsp. of Apharitis acama 156 belli, Epitola 87 belli, (Aurivillius), Iolaus (Epamera) 145 belli, (Hewitson), Iolaus (Epamera) 145 bellina, Iolaus (Epamera) 145 bemba, Pilodeudorix 105 benedirensis, subsp. of Anthene otacilia 193 benguellana, subsp. of Pentila nyassana 18 beni, Pseuderesia 34 benigna, Azanus 252 benitensis, Phytala 85 benitonis, ab. of Larinopoda lircaea 45 berbera, subsp. of Iolaus (Epamera) mimosae bergeri, Eresina 63 bergeri, Iolaus (Iolaphilus) 135 bertha, Iolaus (Epamera) 145 bertha, Pentila 17 beryllina, Hewitsonia 94 bibulus, Lachnocnemus 98 bibundana, Thermoniphas 243 bibundica, ab. of Larinopoda lircaea 45 bicaudatus, subsp. of Iolaus (Epamera) alienus 145 bichroma, Eresinopsides 62 bicolor, Lipaphnaeus 161 bicolor, Pseuderesia 34 bicolora, Alaena 9 bicoloria, Megalopalpus 97 bigoti, subsp. of Liptena (Liptena) ferrymani bihe, Anthene 190 bihe, subsp. of Anthene liodes 192

bilinea, Eresina 63 bilineata, Iolaus 150 bimacula, Telipna 12 bimaculata, Virachola 109 bipuncta, Anthene 190 bistrigatus, subsp. of Telipna sanguinea 12 bitje, Anthene 190 bitje, Argyrochiela 67 bitje, subsp. of Lipaphnaeus leonina 161 bitjeana, Hewitsonia 94 bitjeana, Hypolycaena 120 bitjeana, Micropentila 60 bitjeensis, Ornipholidotos 25 bitjeensis, Aslauga 69 bitjensis, Toxochitona 65 boeticus, Lampides 217 boisduvali, Hewitsonia 94 bolivari, Liptena (Liptena) 53 boma, subsp. of Harpendyreus juno 224 bonifacei, Syrmoptera 117 borealis, f. of Axiocerses amanga mendeche 168 borealis, subsp. of Hewitsonia boisduvali 94 borealis, subsp. of Lepidochrysops neonegus 23I boschi, Leptomyrina (Leptomyrina) 169 bowkeri, Iolaus (Stugeta) 125 bowkeri, Tarucus 216 brabo, Lepidochrysops 229 brachycera, Thestor 183 brahami, Aphnaeus 153 braunsi, Thestor 183 brenda, f. of Larinopoda aspidos 45 Brephidium 253 brevidentatus, Syntarucus 221 brimo, Lachnocnemus 98 brooksi, subsp. of Poecilmitis thysbe 178 browni, Euchrysops 235 bruneeli, Aphnaeus 153 brunnea, Micropentila 60 brunnea, Spindasis 158 brunneus, Cupidesthes 187 brunneus, Euchrysops 235 bryki, Iolaus (Epamera) 145 buchanani, Apharitis 156 buchholzi, Anthene 190 budama, Lepidochrysops 229 budduana, subsp. of Epitola conjuncta 88 bueronica, Oboronia 245 bugalla, subsp. of Thermoniphas togara 243 bunyoro, Micropentila 60 burgeoni, subsp. of Mimacraea eltringhami 30 busoga, Lachnocnemus 98

busoga, Pseudaletis 114 butha, Lepidochrysops 229 butleri, Anthene 190 buxtoni, Hypolycaena 120 bwamba, Diopetes 107 bwamba, Epitola 87 bwamba, subsp. of Falcuna orientalis 47 bwamba, Iridana 75

Cacyreus 203 caerulea, Cupidesthes' 187 caerulea, subsp. of Hypolycaena lebona 120 caerulea, subsp. of Iolaus (Epamera) cytaeis 146 caerulea, f. of Iolaus (Stugeta) bowkeri 125 caerulea, Lepidochrysops 229 caerulea, Pilodeudorix 105 caerulea, Thermoniphas 243 caerulea, Triclema 199 caerulescens, ab. of Oxylides faunus 115 caesareus, Iolaus (Iolaphilus) 135 caffer, Spindasis 158 caffraria, Lepidochrysops 229 caissa, Alaena 9 calice (Hopffer), Castalius 211 calice (Wallengren), Castalius 211 caliginosa, Virachola 109 calisto, Iolaus (Iolaphilus) 135 calviniae, subsp. of Thestor dicksoni 183 camerona, Pilodeudorix 106 camerona ugandae, Pilodeudorix 106 cameroona, subsp. of Liptena (Liptena) decipiens 54 cameroonica, subsp. of Falcuna libyssa 47 camerunensis, Ornipholidotos 25 camerunica, Pentila 17 camillus, Azanus 252 campimus, Falcuna 46 cana, subsp. of Leptomyrina (Gonatomyrina) lara 170 canescens, Hypokopelates 103 canissus, Iolaus (Epamera) 145 capensis, Crudaria 180 Capys 171 cerana, Castalius 211 carcassoni, Pentila 17 carcassoni, Stempfferia 84 carcina, Epitola 87 carilla, Epitoma 87 carina, Iolaus (Iolaus) 139 carlota, Mimeresia 33 carnuta, Telipna 12 carpenteri, subsp. of Epitola catuna 87

carpenteri, Iolaus (Stugeta) 126 carsoni, Lepidochrysops 229 casca, Syntarucus 221 Castalius 206 catalina, Liptena (Liptena) 53 catella, Diopetes 107 cataractae, f. of Pentila nyassa 19 catauga, subsp. of Pentila cloetensi 17 catharina, subsp. of Cupidopsis cissus 240 catharina, subsp. of Pseuderesia eleaza 35 catharus, Capys 172 catocata, Micropentila 60 catori, Egumbia 73 catori, Epitolina 83 catori, Hypokopelates 103 catori, Iolaus (Etesiolaus) 149 catori, subsp. of Mimeresia debora 33 catori, subsp. of Pentila picena 19 catori, subsp. of Phytala elais 85 catori, Pseudaletis 114 catuna, Epitola 87 celaeus, (Cramer) Lepidochrysops 229 celaeus, (Trimen) Lepidochrysops 229 cellularis, Mimeresia 33 centralis, subsp. of Micropentila brunnea 60 cephena, Epitola 88 ceraunia, Epitola 88 cercene, Epitola 88 cercenoides, Epitola 88 ceres, Iolaus (Hemiolaus) 124 chaka, Spindasis 158 chalybeata, Virachola 109 chalybeatus, Aphnaeus 153 chapini, Aphnaeus 153 charmian, Mimacrea 30 cherereti, Micropentila 60 chibonotana, subsp. of Phlyaria heritsia 201 Chilades 246 chirinda, Anthene 190 chloauges, Lepidochrysops 229 Chloroselas 162 christina, Pentila 17 chrysantus, Chrysoritis 179 chrysaor, Poecilmitis 177 chryseostictus, Neurellipes 197 Chrysorichia 166 Chrysoritis 179 chyluensis, subsp. of Ornipholidotos peucetia chyulu, subsp. of Pentila tropicalis 20 ciconia, Epitola 88 ciliaris, subsp. of Cacyreus lingeus 205 cinerea, Lepidochrysops 229 cingulum, Micropentila 60

cissus, Cupidopsis 240 Citrinophila 37 citrrifascia, f. of Mimacraea krausei 31 citrimacula, Telipna 12 citronensis, Liptena (Liptena) 53 clarensis, subsp. of Pentila nyassa 18 clarki, Durbaniella 28 clavum, subsp. of Phasis thero 172 clenchi, Pseuderesia 34 cleodora, Zizula 261 cloetensi, Pentila 17 clymenus, Pseudaletis 114 Cnodontes 43 coarctata, ab. of Aethiopana honorius 92 cobalitina, Hypokopelates 103 cobaltina, Iolaus (Hemiolaus) 124 coeculus (Hopffer), Iolaus (Hemiolaus) 124 coeculus (Mabille), Iolaus (Hemiolaus) 124 coelestis, Iolaus (Epamera) 145 coerulea, Epitola 88 coeruleoalbus, Azanus 252 coeruleoarcuatus, Eicochrysops 238 coffea, subsp. of Virachola lorisona 110 colarata, subsp. of Thermoniphas micylus collitorum, Liptena (Tetrarhanis) 57 concepcion, Epitola 88 condamini, Hypolycaena 120 condamini, subsp. of Pentila cloentensi 18 confusa, Liptena (Liptena) 53 congoana, subsp. of Alaena amazoula 9 congoana, Epitola 88 congoana, Liptena (Liptena) 53 congoana, Pilodeudorix 106 congoana, f. of Pentila occidentalium 19 congoensis, subsp. of Hewitsonia boisduvali congoensis, subsp. of Iolaus (Tanuetheira) timon 129 congoensis, subsp. of Liptena (Liptena) praestans 55 congoensis, Ornipholidotos 25 congoensis, subsp. of Teratoneura isabellae congolanus, subsp. of Spindasis trimeni 159 conguensis, Euchrysops 235 conjuncta, Epitola 88 conjungens, f. of Cupidopsis iobates 240 connectens, subsp. of Alaena picata 9 connectens, subsp. of Pentila umangiana 20 connexa, ab. of Anthene zenkeri 195 connexivus, subsp. of Capys disjunctus 172 conradsi, Aloeides 175 conradti, Eresina 63

consanguinea, Telipna 12	daltoni, subsp. of Liptena (Tetrarhanis) ilm
contracta, Chilades 248	57
contrastata, Anthene 190	dama, subsp. of Pentila nyassana 18
convexa, Epitola 88	damarensis, Aloeides 176
cooksoni, Erikssonia 181	Dapidodigma 121
cooksoni, Euthecta 41	dardanella, Pseudaletis 114
Cooksonia 13	dariaves, Virachola 109
coomasiensis, subsp. of Liptena (Liptena)	darius, Cacyreus 205
xanthostola 56	darwinii, Mimacraea 30
corax, ab. of Myrina silenus 112	daveyi, Epitola 88
cordatus, Uranothauma 203	debilis, Pilodeudorix 106
cordelia, f. of Epitolina dispar 83	debora, Mimeresia 33
cornucopiae, Pseuderesia 36	deborula, subsp. of Mimeresia debora 33
coronae, Aphnaeus 153	
corruscans, Diopetes 107	decaryi, Euchrysops 235
corynetes, Eresina 63	decellei, Epitola 88
costleyi, Mimacraea 30	decempunctata, Liptena (Liptena) 54
	decipiens, Liptena (Liptena) 54
cottoni, subsp. of Iolaus (Etesiolaus) catori	decolorata, f. of Mimeresia moreelsi tas
149 cottoni Powallana or	manni 33
cottoni, Powellana 95	deficiens, f. of Azanus mirza 252
cottrelli, Iolaus (Iolaphilus) 135	deficiens, ab. of Larinopoda lagyra f. emilia
cottrelli, subsp. of Lepidochrysops intermedia	45
230	definita, Anthene 191
coxii, Lepidochrysops 230	delagoensis, Crudaria 180
crawshayi, Anthene 190	delatorum, Uranothauma 203
crawshayi, Euchrysops 235	delegorguei, Lachnocnemus 98
crawshayi, Iolaus (Argiolaus) 131	delicata, Lepidochrysops 230
crawshayi, Uranothauma 203	delicatula, Eicochrysops 238
crawshayinus, Euchrysops 235	delicatula, Teriomima 40
creta, Iolaus (Epamera) 145	Deloneura 76
cretosus, Castalius 211	demaculata, ab. of Pseuderesis isca 35
crippsi, Hewitsonia 94	demeter, Dapidodigma 122
croesus, Axiocerses 168	dentatis, subsp. of Aloeides thyra 176
crola, Eresina 63	depuncta, subsp. of Telipna sanguinea 12
crowleyi, Epitola 88	derema, Pentilia
Crudaria 180	deritas, Diopetes 107
cruenta, Axiocerses 168	dermaptera, Myrina 112
crustaria, Spindasis 158	derona, Virachola 109
cuneatum, Uranothauma 203	Desmolycaena 164
Cupidesthes 185	desmondi, Lepidochrysops 230
Cupidopsis 239	
cuprescens, Euchrysops 235	despecta, Liptena (Liptena) 54
cupreus, Lepidochrysops 230	Deudorix 98
cuprifascia, Cupidesthes 187	dewitzi, Anthene 191
cyanea, Epitola 88	dewitzi, Epitola 88
cyara, Phlyaria 201	dexamene, Pseudonacaduba 227
cyclopteris, Euchrysops 235	diametra, subsp. of Iolaus (Epamera
Cyclyrius 217	aphnaeoides 145
cydaria, subsp. of Pentila picena 19	dicksoni, Cacyreus 205
cynica, Spindasis 159	dicksoni, subsp. of Lepidochrysops methymn
cytaeis, Iolaus (Epamera) 146	231
	dicksoni, Poecilmitis 178
	dicksoni, Thestor 183
daemon, Liptena (Liptena) 53	dilatata, subsp. of Falcuna campimus 46

dinochares, Virachola 109 dinomenes, Virachola 109 dinora, Mimeresia 33 diocles, Virachola 110 diomedes, Virachola 110 Diopetes 106 diopolis, Virachola 110 discimacula, Anthene 191 discirubra, subsp. of Mimeresia dinora 33 disjunctus, Capys 172 dispar, Epitolina 83 disrupta, Lachnocnemus 98 distincta, Thermoniphas 243 distinctesignatus, f. of Uranothauma nubifer 203 distractus, Eicochrysops 238 divaricatus, subsp. of Iolaus (Philiolaus) parasilanus 137 divergens, Lachnocnemus 98 diversa, Liptena (Tetrarhanis) 57 divisa, subsp. of Aethiopana honorius 92 divisa, f. of Hypolycaena buxtoni 120 diyllus, Pilodeudorix 106 docilis, Pseudonacaduba 227 docus, Spalgis 185 dohertyi, subsp. of Mimacraea marshalli 31 dohertyi, Virachola 110 doleta, Epitola 88 dollmani, Lepidochrysops 230 dolores, f. of Iolaus (Hemiolaus) coeculus 124 dolorosa, Epitola 88 dolorosa, Euchrysops 235 dondoensis, subsp. of Deloneura millari 77 dorothea, Epitola 88 dorothea, Falcuna 47 dorothea, Micropentila 60 drucei, subsp. of Actizera lucida 263 drucei, Mimeresia 33 drucei, subsp. of Paraphnaeus hutchinsoni dubia, Epitola 88 dubia, f. of Epitolina dispar 83 dubia, Hypolycaena 120 dubiosa, Baliochila 42 dubiosa, Iolaus (Epamera) 146 dudgeoni, Eiocochrysops 238 dukei, Thestor 183 dulcis, Anthene 191 dunia, Epitola 88 duponti, ab. of Iolaus (Hemiolaus) coeculus 124 durbani, Lachnocnemus 98 Durbania 27 durbania, Liptena (Liptena) 54

Durbaniella 28 Durbaniopsis 28

Ebepius 78 echo, subsp. of Telipna bimacula 12 ecaudata, f. of Cacyreus palemon 206 ecaudata, Virachola 110 edwardsi, Virachola 110 efulena, subsp. of Axiocerses harpax 168 egerides, f. of Aloeides thyra 176 Egumbia 71 Eicochrysops 237 eicotrochilus, Eicochrysops 238 eketi, Liptena (Liptena) 54 elaidina, Phytala 85 elais, Phytala 85 eleala, Hypokopelates 104 elealodes, Hypokopelates 104 eleaza, Pseuderesia 35 eleusis, Chilades 248 elfrieda, Pentila 18 elfriedana, f. of Pentila cloetensi 17 elgonae, subsp. of Iolaus (Argiolaus) crawshayi 131 elgonae, Lepidochrysops 230 elgonae, subsp. of Mimacraea krausei 31 elgonensis, subsp. of Anthene hobleyi III elicola, Chilades 248 elion, Epitola 88 elisa, Iolaus (Iolaphilus) 135 elizabetha, subsp. of Pentila nyassana 18 elissa, Epitola 88 ella, Spindasis 159 elorea, Oboronia 245 elpinice, Pentila 18 eltringhami, Mimacraea 30 emarginata, Ornipholidotos 25 emilia, f. of Larinopoda lagyra 45 emma, Iolaus (Epamera) 146 emolus, Anthene 146 emperamus, Lachnocnemus 98 endymion, Poecilmatis 178 entebbae, subsp. of Iolaus (Iolaphilus) aelianus 133 entebbeana, subsp. of Epitola cephena 88 Epamera, subg. of Ilaus 139 Epitola 85 Epitolina 82 erasmus, ab. of Citrinophila erastus 39 erastus, Citrinophila 39 Eresina 63 Eresinopsis 61 erica, Telipna 12

ericus, Cacyreus 205 erikssoni, Aphnaeus 153 Erikssonia 181 erna, Spindasis 159 ernesti, Egumbia 73 erosine, Phasis ertli, Castalius 211 ertli, subsp. of Iolaus (Epamera) tajoraca 148 ertli, subsp. of Mimacraea charmian 30 erycinoides, Liptena (Liptena) 54 erylus, Hypolycaena 120 erythropoecilus, Anthene 191 esmeralda, subsp. of Chloroselas tamaniba Etesiolaus, subg. of Iolaus 148 ethiopica, subsp. of Iolaus (Stugeta) bowkeri ethiopicus, Cacyreus 205 ethode, Azanus 252 etoumbe, Liptena (Tetrarhanis) 57 euadrus, Aloeides 176 Euchrysops 233 eukrinaria, Liptena (Liptena) 54 eukrines, Liptena (Liptena) 54 eukrinoides, Liptena (Liptena) 54 Euliphyra 70 Euliphyrodes 71 euprepes, Iridana 75 eurema, Larinopoda 45 eurisus, Iolaus (Iolaus) 139 eustorgia, Lipaphnaeus 161 Euthecta 51 evanescens, Liptena (Liptena) 54 exclusa, Lepidochrysops 230 exiguus, Lachnocnemus 98 exquisita, subsp. of Iolaus (Epamera) bellina exquisita, Iridana 75 exsuperia, Telipna 12 extincta, ab. of Telipna consanguinea 12

Falcuna 46
falkensteini, Epitola 88
falkensteini, Uranothauma 203
fallax, Lipaphnaeus 161
fallax, Pentila 18
farquharsoni, Iolaus (Epamera) 146
fasciatus, Triclema 199
fatima, Liptena (Liptena) 54
faunus, Oxylides 115
favillacea, Mimeresia 33
fayei, subsp. of Pentila preussi 19
felthami, Poecilmitis 178

felthami, f. of Uranothauma antinorii 203 femina, Oxylides 115 feminina, Hypokopelates 104 ferruginea, Liptena (Liptena) 54 ferrulineata, Alaena 9 ferrymani, Liptena (Liptena) 54 frater, Iolaus (Epamera) 146 fervida, Telipna 12 fescennia, Euchrysops 235 fidecula, subsp. of Myrina silenus 112 fidonioides, Pentila 18 flava, subsp. of Athysanota ornata 246 flava, Tumerepedes 80 flavefasciata, Mimacraea 30 flaveola, ab. of Citrinophila erastus 39 flavescens, Aphnaeus 153 flavescens, ab. of Mimacraea landbecki 31 flavicans, Liptena (Liptena) 54 flavelinea, Iolaus (Epamera) 146 flavisquamata, Lepidochrysops 230 flavoantennata, Epitola 88 flavomaculatus, Anthene 191 flavopunctata, Micropentila 60 fontainei, f. of Aphnaeus orcas 153 fontainei, Eicochrysops 239 fontainei, Eresina 63 fontainei, Iolaus (Epamera) 146 fontainei, Micropentila 60 fontainei, subsp. of Myrina sharpei 113 fontainei, subsp. of Pentila umangiana 20 fontainei, subsp. of Pseudaletis nigra 114 fontainei, Thermoniphas 243 fontainei, Zeritis 164 fournierae, subsp. of Hypomyrina nomenia fracta, f. of Cacyreus palemon 206 fracta, f. of Spindasis homeyeri 159 fragilis, Baliochila 42 Freyeria 248 freyi, Baliochila 43 frommi, subsp. of Hypolycaena auricostalis 119 fulvaria, Mimacraea 30 fulvescens, Lepidochrysops 230 fulvicans, Liptena (Liptena) 54 fulvimacula, Neuryllipes 197 fulvula, Micropentila 60 fulvus, Neurellipes 197 fumata, Diopetes 107 fumata, f. of Ornipholidotos kirbyi 26 fumosa, Lepidochrysops 230 fumosa, Thermoniphas 243 fusca, Eresina 63 fusca, subsp. of Falcuna synesia 47

fusca, Hypokopelates 104 fusca, Pseuderesia 36 fuscomarginata, Iolaus (Epamera) 146 fuscula, Micropentila 60

gabonensis, subsp. of Falcuna synesia 47 gabonensis, subsp. of Iolaus (Iolaus) carina gabonensis, Ornipholidotos 25 gabunica, subsp. of Cupidesthes voltae 188 gabunica, Iolaus (Iolaphilus) 135 gabunica, Iridans 75 gabunica, Micropentila 60 gabunica, subsp. of Pentila occidentalium 19 gaika, Zizula 261 galathea, Virachola 110 galla, subsp. of Anthene butleri 190 gambius, Virachola 110 gamra, Azanus 252 gazei, Iolaus (Epamera) 146 gelinea, Mimacraea 30 gemmarius, Iolaus (Epamera) 146 gemmifera, Neurellipes 197 genuba, Hypokopelates 104 gerda, Toxochitona 65 gerina, Epitola 88 ghanana, Iridana 75 ghesquierei, Epitola 88 ghimirra, subsp. of Cacyreus palemon 206 gigantea, Lepidochrysops 230 gigas, Megalopalpus 97 gilletti, Apharitis 156 gilloni, Aphnaeus 153 girthii, Liptena (Liptena) 54 glagoessa, Pentila 18 glauca, Lepidochrysops 230 glaucus, Iolaus (Epamera) 146 gloveri, Oxylides 115 gomensis, subsp. of Hewitsonia intermedia Gonatomyrina, subg. of Leptomyrina 170 goodi, Epitola 88 gordoni, subsp. of Epitola staudingeri 90 gordoni, Liptena (Liptena) 54 gorgias, Leptomyrina (Gonatomyrina) 170 gracilis, Kopelates 102 graeseri, Mimacraea 31 grahami, Lepidochrysops 230 grammicus, Tarucus 216 grandis, Lepidochrysops 230 gregorii, subsp. of Castalius calice 211 griqua, subsp. of Castalius melaena 212 grisea, subsp. of Anthene levis 192

griseata, subsp. of Mimeresia favillacea 33 griseus, Aloeides 176 grosei, Anthene 191 guichardi, Lepidochrysops 230 gussfeldti, Oboronia 245 gyrala, f. of Larinopoda lagyra 45 haemus, subsp. of Iolaus (Epamera) mimosae handmani, Iolaus (Epamera) 146 handmani, Leptomyrina (Leptomyrina) 169 hapale, Liptena (Liptena) 54 harpax, Axiocerses 168 harpax piscatoris, Axiocerses 168 Harpendyreus 221 hatita, Hypolycaena 120 hauttecoeuri, subsp. of Alaena interposita 9 hawkeri, Lepidochrysops 230 hedwiga, Pentila 18 helena, Liptena (Liptena) 54 heliodorus, Aphnaeus 153 helius, Iolaus (Iolaus) 139 hemicyanus, Iolaus (Epamera) 146 Hemiolaus, subg. of Iolaus 123 henleyi, Phytala 85 henryi, Iolaus (Iolaphilus) 135 Heodes 263 heritsia, Phylaria 201 hermansi, f. of Larinopoda lircaea 45 hesiodus, Oxylides 115 hewitsoni, Anthene 191 hewitsoni (Mabille), Epitola 88 hewitsoni (Staudinger), Epitola 89 hewitsoni, Euliphyra 71 hewitsoni, Pentila 18 Hewitsonia 93 hewitsonioides, Epitola 89 hiendlmayri, Liptenara 21 hilarion, Cupidesthes 181 hildegarda, Baliochila 43 hintza, Castalius 211 hippocrates, Eicochrysops 239 hirundo, Leptomyrina (Leptomyrina) 169 hobleyi, Anthene 191 hodsoni, Anthene 191 hollandi, subsp. of Aphnaeus orcas 153 hollandi, Falcuna 47 hollandi, Pilodeudorix 106 hollandi, Telipna 12 holmsei, Thestor 183 homeyeri, Liptena (Liptena) 54 homeyeri, Spindasis 159 homeyeri, Syrmoptera 117 honorius, Aethiopana 92 horus, Euchrysops 235

hulstaerti, Liptena (Liptena) 54
hutchinsoni, Paraphnaeus 155
hyetta, Phytala 85
hyettina, Phytala 85
hylax, Zizula 261
hymen, Dapidodigma 122
hypocala, Iridana 75
Hypokopelates 102
hypoleucus, Lepidochrysops 230
Hypolycaena 117
Hypomyrina 100
hypopolia, Lepidochrysops 230
Hyreus, 203

iasis, Iolaus (Epamera) 146 iaspis, Iolaus (Epamera) 146 ideoides, Liptena (Liptena) 54 ignota, Lepidochrysops 230 ikoya, Epitola 89 ilala, subsp. of Liptena (Tetrarhanis) diversa ilma, Liptena (Tetrarhanis) 57 ilogo, Aphnaeus 153 immaculata, Deloneura 77 immaculata, subsp. of Liptena (Liptena) opaca 55 immaculata, f. of Pentila occidentalium 19 immaculata, f. of Virachola lorisona 110 immaculatus, f. of Cupidopsis cissus 240 imperialis, subsp. of Lepidochrysops cinerea inconspicua, Triclema 199 inconspicua, Pentila 18 incredibilis, Iridana 76 incurvata, f. of Mimacraea neurata 31 indefinita, Anthene 191 inferna, Triclema 199 infima, Liptena (Liptena) 54 inframacula, Liptena (Liptena) 54 infusca, Hypokopelates 104 innesi, Deloneura, 77 innocentia, f. of Larinopoda lircaea 45 inores, Iolaus (Sukidion) 150 insulana, Epitola 89 insulanus, subsp. of Syntarucus pirithous 221 intermedia, Epitola 89 intermedia, Hewitsonia 94 intermedia, Lepicochrysops 230 intermedia, Liptena (Liptena) 54 intermedia, subsp. of Phlyaria heritsia 201 intermixta, Phytala 85 interposita, Alaena 9 interrupta, subsp. of Alaena picata 9

interruptus, subsp. of Castalius melaena 212 inundifera, Argyrochiela 67 inyangae, Lepidochrysops 230 iobates, Cupidopsis 240 Iolaphilus, subg. of Iolaus 131 lolaus 122 Iolaus, subg. of Iolaus 138 Iridana 74 Iridopsis 74 Iris 74 iruma, Cupidesthes 188 irvingi, Lepidochrysops 230 isabellae, f. of Mimeresia libentina 33 isabellae, Teratoneura 74 isarchus, Azanus 252 isca, Pseuderesia 35 isis, Azanus 252 ismenius, Iolaus (lolaphilus) 135 itea, Azanus 252 iturensis, Iolaus (Iolaphilus) 135 ituri, Hypokopelates 104 ituria, Anthene 191 ituriensis, Triclema 199 iturina, Epitola 89 iturina, Falcuna 47 ivoirensis, subsp. of Lipaphnaeus leonina iza, Spindasis 159

ja, Anthene 191 ja, Telipna 12 jacksoni, Aphnaeus 153 jacksoni, Axiocerses 168 jacksoni, Epitola 89 jacksoni, Eresina 63 jacksoni, Euchrysops 235 jacksoni, Hypolycaena 120 jacksoni, Iolaus (Epamera) 146 jacksoni, Iridana 76 jacksoni, Lepidochrysops 230 jacksoni, Liptena (Liptena) 54 jacksoni, subsp. of Liptena (Liptena) rubromacula 55 jacksoni, Micropentila 60 jacksoni, subsp. of Oborona punctatus 245 jacksoni, Ornipholidotos 25 jacksoni, Pseudoneaveia 82 jacksoni, Virachola 110 jamesoni, Iolaus (Iolaphilus) 135 janna, subsp. of Anthene opalina 193 jansei, Lepidochrysops 230 japhusa, subsp. of Hypolycaena hatita 120 jeanneli, subsp. of Anthene rubricinctus 194 jeanneli, Syntarucus 221
jefferyi, Aphnaeus 153
jefferyi, subsp. of Eresinopsides bichroma 62
jefferyi, Lepidochrysops 230
jesous, Azanus 252
johanna, Alaena 9
jordanus, subsp. of Iolaus (Epamera) glaucus 146
juba, Anthene 191
julianus, Iolaus (Iolaphilus) 135
julius, Iolaus (Iolaphilus) 135
julus, Iolaus (Iolaphilus) 135
julus, Iolaus (Iolaphilus) 135
juno, Harpendyreus 224

kabrosae, Euchrysops 235 kedassa, Diopetes 107 kadiskos, Hypolycaena 120 kadugli, f. of Axiocerses harpax 168 kaduglii, f. of Apharitis nilus 156 kaffana, Castalius 212 kafuensis, Hypokopelates 104 kagera, subsp. of Alaena caissa 9 kalinzu, Neurypexina 196 kallimoides, Paraslauga 70 kallimon, subsp. of Spindasis homeyeri 159 kallipygos, Hypokopelates 104 kama, Euchrysops 235 kamerunica, subsp. of Iolaus (Epamera) frater 146 kamilila, Triclema 199 kamitugensis, subsp. of Liptena (Liptena) praestans 55 kamitugensis, Telipna 12 kamitugensis, Thermoniphas 243 kampala, Anthene 191 kamyengensis, Epitola 89 karsandra, Zizeeria 258 karschioides, subsp. of Mimacraea krausei 31 kasai, Falcuna 47 katanga, subsp. of Pilodeudorix camerona 106 katangae, Euchrysops 235 katangae, subsp. of Mimeresia russulus 33 katangae, Ornipholidotes 25 katangae, Telipna 12 katangana, Egumba 73 katangana, Eresina 63 katanganae, Micropentila 60 katanganus, Iolaus (Epamera) 147 katera, Anthene 191 katera, Eresina 63 katera, subsp. of Iolaus (Epamera) neavi 147 katera, Iridana 76

katera, subsp. of Liptena (Liptena) flavicans katera, subsp. of Pseuderesia eleaza 35 katerae, Epitola 89 katerae, Micropentila 60 katherinae, Epitole 89 kayonza, Iolaus (Iolaphilus) 135 kayonza, Virachola 110 kedonga, Chilades 248 kedonga, subsp. of Iolaus (Stugeta) bowkeri kelle, Liptena (Liptena) 54 kelleana, Micropentila 60 kersteini, subsp. of Anthene larydus 192 kholifa, Epitola 89 kiellandi, Alaena 9 kiellandi, subsp. of Aloeides molomo 176 kigezi, subsp. of Anthene hobleyi 191 kigezi, subsp. of Iolaus (Argiolaus) lalos 131 kigezi, subsp. of Liptena (Tetrarhanis) stempfferi 57 kigezi, Thermoniphas 243 kikuyu, subsp. of Anthene otacilia 193 kilimandjarensis, Lepidochrysops 231 kinumbensis, f. of Hypokopelates mera 104 kirbyi, Hewitsonia 94 kirbyi, Ornipholidotos 25 kisaba, subsp. of Harpendyreus meruanus kitale, subsp. of Lepidochrysops cinerea 229 kitobolensis, subsp. of Virachola antalus 109 kivuensis, subsp. of Lepidochrysops parsimon knobeli, f. of Phasis sardonyx 173 knysna, Zizeeria 258 koena, Lepidochrysops 231 kohli, Pilodeudorix 106 knotu, subsp. of Castalius carana 211 Kopelates 102 krausei, Mimacraea 31 kulala, Tarucus 216 kumboae, subsp. of Iolaus (Epamera) 146

labuschagnei, f. of Phasis argyraspis 173 labwor, Lepidochrysops 231 lachares, Anthene 191 Lachocnema 97 lacides, Triclema 199 lacrimosa, Lepidochrysops 231 lacta, Mimacraea 31 lacteata, Falcuna 47 lactinatus, subsp. of Castalius cretosus 211 lagyra, Larinopoda 45

lalos, Iolaus (Argiolaus) 131 leonina, Lipaphnaeus 161 lamborni, Alaena 9 leonis, subsp. of Iolaus (Epamera) cytaeis lamborni, Aslauga 69 146 lamborni, Epitola 89 leonis, Pseudaletis 114 lamborni, Neaveia 81 leopardina, subsp. of Pentila nyassana 19 lamias, Triclema 199 Lepidochrysops 227 laminiter, Liptena (Tetrarhanis) 57 leptala, Anthene 192 Lampides 217 leptines, Anthene 192 lamprocles, Neurypexina 196 Leptomyrina (Leptomyrina) 168 landana, f. of Falcuna synesia 47 leroma, Crudaria 180 landbecki, Mimacraea 31 lerothoda, Lepidochrystops 231 letsea, Lepidochrysops 231 landbecki, Pentila 18 laon, Iolaus (Epamera) 147 leucocerus, Iolaus (Trichiolaus) 128 lanonides, Iolaus (Iolaphilus) 135 leucocyana, Thermoniphas 243 Leucolepis 47 laplumei, subsp. of Telipna acraeoides 11 lara, Larinopoda 45 leucon, Lepidochrysops 231 lara, Leptomyrina (Gonatomyrina) 170 leucostola, Liptena (Liptena) 55 Larinopoda 44 leucyana, Euliphyra 71 larydas, Anthene 192 leura, f. of Pentila hewitsoni 18 lasti, Anthene 192 levis, Anthene 192 lianna, Epitola 89 lasti, f. of Pentila tropicalis 20 latifascia, f. of Pentila cloentensi 17 liara, Hypolycaena 120 lathyi, subsp. of Liptena (Tetrarhanis) ilma libentina, Mimeresia 33 liberiana, Oboronia 245 laticlavia, Diopetes 107 libussa, Larinopoda 45 latifascia, ab. of Mimacraea landbecki 31 libyssa, Falcuna 47 latifasciata, f. of Anthene rubricinctus libyssa confluens, Falcuna 47 jeanneli 194 libyssa latimarginata, Falcuna 47 latifasciata, ab. of Hypolycaena antifaunus licinia (Mabille), Virachola 110 licinia (Trimen), Virachola 110 ligures, Anthene 192 latifimbriata, Lipaphnaeus 161 latimaculata, subsp. of Hypolycaena antiliguroides, subsp. of Anthene definita 191 likouala, Eresina 63 likouala, Neurellipes 197 latimarginata, subsp. of Baliochila minima lilacina, subsp. of Lepidochrysops pterou 232 latimarginata, f. of Larinopoda aspidos 45 limbata, Citrinophila 39 latimarginata, Spalgis 185 limbata, Durbania 27 latimargo, Ornipholidotes 26 limbata, subsp. of Pentila hewitsoni 18 latruncula, Euchrysops 235 linearis, subsp. of Tarucus sybaris 216 latruncularia, Liptena (Liptena) 55 lineata, subsp. of Mimacraea neurata 31 lavinia, Pentila 18 lineosa, f. of Hypokopelates ituri 104 lebona, Hypolycaena 120 lingeus, Cacyreus 205 Lactiles, 56 liodes, Anthene 192 legrasi, Tarucus 216 Lipaphnaeus 159 lekanion, Iolaus (Iolaphilus) 135 lipara, Baliochila 43 lemnos, Anthene 192 liparis, Anthene 192 lemolea, Spalgis 185 Liptena 47 leonae, Aslauga 69 Liptena, subg. of Liptena 53 leonensis, Epitola 47 Liptenaria 20 leonensis, Falcuna 47 lircaea, Larinopoda 45 leonina, Cupidesthes 188 lithas, Anthene 192 leonina (Bethune-Baker), Epitola 89 lithas, Cupidesthes 188 leonina (Staudinger), Epitola 89 littoralis, subsp. of Aphnaeus coronae 153 leonina, Hypokopelates 104 littoralis, subsp. of Ebepius ochrascens 79

littoralis, subsp. of Iolaus (Argiolaus) crawshayi 131 littoralis, ab. of Iolaus (Hemiolaus) coeculus livia, Virachola 110 livida, subsp. of Anthene butleri 190 loa, subsp. of Anthene lemnos 192 lochias, Anthene 192 locra, Anthene 192 locuples, Anthene 192 loewensteini, Lepidochrysops 231 lois, Euchrysops 235 longicauda, Iolaus (Epamera) 147 longifalces, Lepidochrysops 231 lorisona, Virachola 110 lotana, Lepidochrysops 231 louisae, Tarucus 216 loveni, Lepidochrysops 231 loxura, Lipaphnaeus 161 lucayensis, subsp. of Pentila cloentensi 18 lucida, Actizera 263 lucretia, Triclema 199 lucretilis, Triclema 199 lukabas, Iolaus (Iolaphilus) 135 lukenia, Lepidochrysops 231 lukokesha, Anthene 192 lulua, Iolaus (Pseudiolaus) 127 luna, Lachnocnemus 98 lunaris, Pentila 18 lunifer, subsp. of Uranthauma poggei 203 lunulata, Anthene 192 lunulifer, subsp. of Lepidochrysops cinerea lusambo, Pseudaletis 114 lusones, Neurellipes 197 luteomaculata, Mimacraea 31 lutosa, Spindasis 159 lutzi, Triclema 199 lybia, Falcuna 47 Lycaena 263 lycaenoides, Larinopoda 45 lycegenes, Poecilmitis 178 lychnaptes, Anthene 192 lychnides, Anthene 192 lychnoptera, Anthene 192 lycia, Poecilmitis 178 lycotas, Anthene 192 lydia, Triclema 199 lyncurium, Poecilmitis 178 lysander, Poecilmitis 178 lysias, Anthene 192 lysimon, Zizeeria 258 lysimon, Zizula 261 lyzanius, Neurypenxinus 196

mabangi, Micropentila 60 mabillei, f. of Anthene princeps smithi 193 mabillei, subsp. of Iolaus (Philiolaus) parasilanus 137 macalanga, Azanus 252 maculata, Alaena 9 maculata, Epitola 89 maculata, subsp. of Pentila abraxas 17 madiberensis, Alaena 9 madiberensis, Anthene 193 maesa, Iolaus (Epamera) 147 maesseni, Eresina 63 maesseni, subsp. of Iolaus (Philiolaus) prasilanus 137 maeseni, Triclema 200 mafugae, subsp. of Iolaus (Epamera) aphnaeoides 145 magda, Virachola 110 magdalenae, Hewitsonia 94 magna, f. of Anthene lunulata 192 magna, Lachnocnema 98 magnifica, Epitola 89 magnifica, Iridana 76 mangificans, f. of Uranothauma antinorii magnimaculata, subsp. of Pseuderesia isca mahallakoena, Eicochrysops 239 mahota, Anthene 193 major, Alaena 9 major, subsp. of Euchrysops subpallida 236 major, Harpendyreus 224 major, subsp. of Lepidochrysops lacrimosa 23I makala, Anthene 193 makala, subsp. of Hypokopelates angelita 103 makala, Leptomyrina (Leptomyrina) 169 makoniensis, ab. of Larinopoda lircaea 45 melaena, Castalius 212 malagrida, Aloeides 176 malathana, Euchrysops 235 mandersi, Cyclyrius 219 mapongua, Pseuderesis 36 mara, Epitola 89 margaritacea, Alaena 9 margaritaceus, Castalius 212 margarita, Falcuna 47 marginalis, Aslauga 69 marginalis, Citrinophila 39 marginalis, Syntarucus 221 marginaria, subsp. of Aslauga purpurascens marginata, subsp. of Anthene crawshayi 190

marginata, Aslauga 69	mendeche, subsp. of Axiocerses amanga 168
marginata, Epitola 89	mendeche bistrigata & Axiocerses 168
marginata, Hypokopelates 104	mendeche bistrigata 2 Axiocerses 168
margites, Iolaus (Hemiolaus) 124	menelas, Spindasis 159
maria, subsp. of Iolaus (Stugeta) bowkeri 126	mengoensis, Epitola 89
mariae, Mimacraea 31	menna, Lepidochrysops 231
mariae, Telipna 12	mera, Hypok pelates 104
mariana, Hypokopelates 104	mercedes, Epitola 89
mariana, subsp. of Pentila rothi 20	mermeros, Iolaus (Trichiolaus) 128
marina, Iridana 76	mermis, Iolaus (Epamera) 147
maris, subsp. of Iolaus (Epamera) bellina 145	meruanus, Harpendyreus 224
maritimus, Iolaus (Iolaphilus) 135	mesia, Pentila 18
marlieri, Harpendyreus 224	messapus, Eicochrysops 239
marmorata, subsp. of Alaena nyassae 9	metaleucus, Megalopalpus 97
marmorea, Iolaus (Stugeta) 126	methymna, Lepidochrysops 231
mars, Aloeides 176	metophis, Brephidium 254
marshalli, Aloeides 176	micra, Teriomima 40
marshalli, Aphnaeus 153	Micropentila 58
marshalli, subsp. of Aslauga purpurascens 69	micylus, Thermoniphas 243
marshalli, Cacyreus 205	inidas, Poecilmitis 178
marshalli, Mimacraea 31	migiurtinensis, Euchrysops 236
marshalli, Triclema 200	inildbraedi, subsp. of Iolaus (Epamera
maryra, Iolaus (Hemiolaus) 124	hemicyanus 146
maryra mabillei, Iolaus (Hemiolaus) 123	mildbraedi, Pseuderesia 35
marungensis, Harpendyreus 224	millari, Anthene 193
masai, Eicochrysops 239	millari, Deloneura 77
masaka, Eresina 63	Mimacraea 29
	Mimeresia 32
mashuna, Lepidochrysops 231 mashunae, subsp. of Aphnaeus erikssoni 153	mimeta, Actis 101
masilikazi, Spindasis 159	mimetica, Cupidesthes 188
masindae, subsp. of Alogides there 126	mimetica, Iolaus (Stugeta) 126
masuruna, subsp. of Aloeides thyra 176	mimosae, Iolaus (Epamera) 147 minima, Anthene 193
matilda, Iolaus (Iolaphilus) 135	The state of the s
mauensis, Euchrysops 235 maureli, Axiocerses 168	minima, Baliochila 43
maureli, subsp. of Iolaus (Argiolaus) craw-	minima, Chloroselas 163
shayi 131	minium, Pseuderesia 35
	minor, Cupidesthes 188
mauritanica, Cupidopsis 240 mayottensis, Syntarucus 221	minuscula, Freyeria 249 minuta, subsp. of Anthene crawshayi 190
mazanguli, Pseudaletis 114	mirabilis, Iolaus (Epamera) 147
mazoensis, Desmolycaena 166	
meander, Neurellipes 197	miranda, Epitola 89 mirifica, Epitola 89
media, subsp. of Mimacraea marshalli 31	mirifica, Euliphyra 71
mediterraneae, Tarucus 216	
medjensis, Telipna 12	mirza, Azanus 252 mittoni, Hewitsonia 94
Megalopalpus 95	mixtura, Syrmoptera 117
melama, Castalius 212	
melambrotus, Anthene 193	mnestra, Epitolina 83 modesta, Aslauga 69
melandeta, Falcuna 47	modesta, Asiauga 09 modesta, f. of Hypokopelates otraeda 104
melanis, Pseudonacaduba 227	modesta, Liptena (Liptena) 55
melanomitra, Syrmoptera 117	
melas, Castalius 212	modestus, Spindasis 159
melissa, f. of Epitolina dispar 83	modestissima, Liptena (Liptena) 55 molomo, Aloeides 176
menas, Iolaus (Iolaphilus) 135	
menas, rotaus (rotapinius) 135	mombasae, subsp. of Pentila tropicalis 20

mombase, subsp. of Iolaus (Stugeta) bowkeri moncus, Anthene 193 mondo, Mimeresia 33 moneta, Micropentila 60 Monile, 196 montana, Spindasis 159 montanus, Thestor 183 monteironis, Anthene 193 moreelsi, Mimeresia 33 moriqua, Azanus 252 moyambina, Epitola 89 moyambina, Hypokopelates 104 moyambina, Iolaus (Epamera) 147 moyambina, Mimeresia 33 moyo, subsp. of Lepidochrysops elgonae 230 mozambica, Spindasis 159 mpanda, Lepidochrysops 231 mpangensis, Epitola 89 mpigi, Micropentila 60 muhata, Ornipholidotos 26 mulsa, Alaena 9 multiplagiata, subsp. of Pentila nyassa 19 multipunctata, f. of Pentila nyassa 19 mumbuensis, subsp. of Alaeides molomo 176 murrayi, Thestor 183 mus, Epitola 89 musagetes, Anthene 193 musagetes elgonensis, Anthene 193 muzizii, f. of Zeritis neriene 164 mwagensis, Liptena (Liptena) 55 *mylica*, Zizula 261 mylothrina, Pentila 18 Myrina 111 mysteriosa, subsp. of Spindasis crustaria 159

naara (Hewitson), Hypolycaena 120 naara (Karsch), Hypolycaena 120 nacrescens, Lepidochrysops 231 naidina, Lepidochrysops 231 nairobiensis, subsp. of Spindasis apelles 158 nais, Poecilmitis 178 namaqua, Spindasis 159 nandanis, Euchrysops 236 nandianus, Eicochrysops 239 of Iolaus (Epamera) nasissii, subsp. aphnaeoides 145 natalensis, Azanus 252 natalensis, subsp. of Durbania amakosa 27 natalensis, Spindasis 159 ndolae, Iolaus (Iolaphilus) 135 neavei, Aphnaeus 153 neavei, Baliochila 43

neavei, Cooksonia 14 neavei, Iolaus (Epamera) 147 neavei, Lepidochrysops 231 neavei, Mimacraea 31 neavei, Mimeresia 33 neavei, subsp. of Telipna angustifascia 11 Neaveia 80 neglecta, Anthene 193 negus (Felder), Lepidochrysops 231 negus Karsch), Lepidochrysops 231 Neochrysops 227 Neoepitola 91 neokoton, Mimacraea 31 neongegus, Lepidochrysops 231 neriene, Zeritis 164 nero, Pentila 18 neurata, Mimacraea 31 Neurellipes 196 Neurypexina 195 nevillei, Lepidochrysops 231 ngoko, Anthene 193 ngonga, Alaena 9 nicephora, Virachola 110 nigeriae, Chilades 248 nigeriae, Epitola 89 nigeriae, Lepidochrysops 231 nigeriae, Ornipholidotos 26 nigeriae, Triclema 200 nigeriana, Iridana 76 nigeriana, Micropentila 60 nigeriana, Pentila 18 nigeriana, Pseuderesia 36 nigeriensis, subsp. of Hewitsonia boisduvali 94 nigra, Epitola 89 nigra, Hpyolycaena 120 nigra, Pseudaletis 114 nigra, subsp. of Pseuderesia eleaza 35 nigra, subsp. of Telipna acraea 11 nigrescens, Phytala 85 nigribasis, f. of Pentila nyassa 19 nigricans, subsp. of Falcuna hollandi 47 nigricans, Poecilmitis 178 nigrita, Lepidochrysops 231 nigrita, f. of Telipna bimacula 12 nigrocaudata, Anthene 193 nigromarginata, Liptena (Liptena) 55 nigropunctata, Anthene 193 nigrostriata, ab. of Actis ula 101 nigrovenata, Epitola 89 nilotica, subsp. of Euchrysops malathana 235 niloticus, subsp. of Iolaus (Argiolaus) crawshayi 131 nilus, Apharitis 156

niobe, Lepidochrysops 231	obscura, Triclema 200
nurmo, Diopetes 107	obscurata, subsp. of Pilodeudorix caerulea
nitide, Epitola 89	105
nivea, Syrmoptera 117	obscurus, Iolaus (Epamera) 147
niveus, subsp. of Anthene sylvanus 194	obscurus, f. of Iolaus (Hemiolaus) coeculus
niveus, Lachnocnemus 98	124
niveocincta, subsp. of Euchrysops reducta 236	obscurus, Thestor 183
nobilis, Pilodeudorix 106	obsolescens, subsp. of Anthene lachares 191
nodieri, subsp. of Castalius cretosus 211	obsolescens, ab. of Mimacraea krausei 31
nolaensis, Iolus (Epamera) 147	obsoleta, subsp. of Liptena (Liptena) eukrines
nonienia, Hypomyrina 100	54
nomion, subsp. of Hypomyrina nomenia 100	obsoleta, subsp. of Neurellipes staudinger
noquasa, subsp. of Harpendyreus tsomo 224	197
notobia, Harpendyreus 224	obsoleta, subsp. of Pentila nyassana 19
ntebi, Ornipholidotos 26	occidentalis, subsp. of Aphnaeus chapini 153
nuba, subsp. of Myrina subornata 113	occidentalis, Azanus 252
nubifer, Uranothauma 203	occidentalis, subsp. of Iolaus (Stugeta
nubifera, Liptena (Tetrarhanis) 57	bowkeri 126
nunu, Pentila 18	occidentalis, Liptena (Liptena) 55
nuptus, subsp. of Dapidodigma demeter 122	occidentalis, Pseudaletis 114
nursei, Iolaus (Epamera) 147	occidentalium, Pentila 19
nyansa, Telipna 12	occidentalium congoana, Pentila 19
nyanzae, Aphnaeus 153	ochracea, ab. of Alaena nyassae 9
nyanzana, Hypokopelates 104	ochraceus, f. of Aloeides conradsi 176
nyanzae, subsp. of Iolaus (Argiolaus) craw-	ochrascens, Ebepius 79
shayi 131	ochrea, subsp. of Alaena maculata 9
nyanzana, subsp. of Iolaus (Stugeta) bowkeri	ochrea, Liptena (Liptena) 55
126	ochreofascia, Anthene 193
nyasae, Baliochila 43	ochreopuncta, subsp. of Cupidopsis iobates
nyasae, Lepidochrysops 231 nyasae, subsp. of Myrina dermaptera 112	oculata, f. of Anthene indefinita 191
	oculatus, Triclema 200
nyasana, subsp. of Alaena amazoula 9 nyasana, subsp. of Iolaus (Stugeta) bowkeri	oculus, subsp. of Lepidochrysops parsimon
126	232
nyassae, Alaena 9	odana, Virachola 110
nyassae, Spindasis 159	ogađenensis, Chloroselas 163
nyassana, Pentila 18	ogađenensis, Hypolycaena 120
nycetus, Aloeides 176	ogojae, Liptena (Tetrarhanis) 57
nyika, Lepidochrysops 231	ogojae, Micropentila 60
nzoia, subsp. of Mimacraea marshalli 31	olalae, subsp. of Iolaus (Stugeta) marmorea
nzoiae, subsp. of Myrina silenus 112	126
, ,	olombo, Liptena (Liptena) 55
oberthuri, Alaena 9	olympus, Anthene 193
oberthueri, subsp. of Iolaus (Epamera) pollux	onesso, Pseuderesia 36
147	onias, Anthene 193
obliquisigna, Lachnocnemus 98	oniens, subsp. of Liptena (Liptena) flavicans
obliterata, f. of Virachola lorisona 110	54
Oboronia 243	oniensis, Epitola 90
obscura, Epitola 89	onitshae, Liptena (Tetrarhanis) 57
obscura, Hypokopelates 104	onitshae, Ornipholidotos 26
obscura, subsp. of Hypolycaena liara 120	opaca, Liptena (Liptena) 55
obscura, Iridana 76	opalina, Anthene 193
obscura, Phytala 85	Oraidium 254
obscura, f. of Spindasis natalensis 159	orcas, Aphnaeus 153

oreas, Chrysoritis 179 oreas, Lepidochrysops 231 orejus, Hypolycaena 120 orientalis, subsp. of Cupidesthes arescopa 187 orientalis, Epitola 90 orientalis, subsp. of Euchrysops osiris 236 orientalis, Falcuna 47 orientalis, subsp. of Neaveia lamborni 81 orientalis, subsp. of Ornipholidotos peucetia orientalis, subsp. of Pilodeudorix diyllus 106 orientalis, subsp. of Pseuderesia paradoxa 36 orientus, subsp. of Iolaus (Tanuetheira) timon 129 ornata, Athysanota 246 ornata, f. of Azanus mirza 252 Ornipholidotos 21 orontius, Lepidochrysops 231 orphna, subsp. of Anthene amarah 190 orthrus, Aloeides 176 ortygia, Lepidochrysops 232 o-rubrum, Liptena (Liptena) 55 orus, Lycaena 264 osbecki, subsp. of Poecilmitis thysbe 178 osheba, Pseuderesia 36 osiris (Hopffer), Euchrysops 236 osiris (Trimen), Euchrysops 236 otacilia (Hewitson), Anthene 193 otacilia (Trimen), Anthene 193 otacilia mashuna, Anthene 193 otlauga, Liptena (Liptena) 55 otraede, Hypokopelates 104 ouesso, Epitola 90 overlaeti, f. of Aphnaeus orcas 153 overlaeti, Chloroselas 163 overlaeti, Falcuna 47 overlaeti, Ornipholidotos 26 owerri, subsp. of Mimeresia drucei 33

pachalica, Hypolycaena 120
palemon, Cacyreus 206
pallene, Iolaus (Aphniolaus) 138
palemon, Cacyreus 206
pallene, Iolaus (Aphniolaus) 138
pallida, f. of Aloeides thyra 176
pallida, subsp. of Citrinophila erastus 39
pallida, Cnodontes 44
pallida, f. of Megalopalpus zymna 97
palmus, Poecilmitis 178
paludicola, Cupidesthes 188
pampolis, Lepidochrysops 232
pan, subsp. of Lipaphnaeus aderna 161
pan, Poecilmitis 178

Oxylides 114

pandora, Aslauga 69 paneperata, Iolaus (Iolaphilus) 135 paradoxa, subsp. of Lipaphnaeus leonina 161 paradoxa, Ornipholidotos 26 paradoxa, Pseuderesia 36 paragora, Mimacraea 31 paralithas, Cupidesthes 188 parallela, subsp. of Anthene crawshayi 191 parapetreia, subsp. of Pentila rogersi 19 Paraphnaeus 154 Parapontia 47 parasilanus, Iolaus (Philiolaus) 137 Paraslauga 69 pardalina, subsp. of Pentila abraxas 17 parrhasius, Chilades 248 parsimon (Fabricius), Lepidochrysops 232 parsimon (Wallengren), Leipodochrysops 232 parva, Freyeria 249 parva, Iolaus (Epamera) 147 parva, subsp. of Telipna carnuta 12 parva, Teriomima 40 pasteon, Dioptes 107 patricia, Lepidochrysops 232 paucipuncta, Pentila pauli, subsp. of Pentila nyassana 19 pauliani, Eicochrysops 239 pauperi, f. of Aphnaeus orcas 153 peculiaris, Lepidochrysops 232 pelion, Poecilmitis 178 pelotus, Uranothauma 203 penningtoni, Capys 172 penningtoni, Cnodontes 44 penningtoni, subsp. of Durbania amakosa 27 penningtoni, Iolaus (Epamera) 147 penningtoni, subsp. of Ornipholidotos peucetia 26 penningtoni, Poecilmitis 178 penningtoni, Thestor 183 penningtoni, Virachola 110 Pentila 15 pephedro, Lepidochrysops 232 perdita, Iridana 76 perfragilis, Ornipholidotos 26 pericles, Anthene 193 perigrapha, Actis 101 peringueyi, f. of Phasis sardonix 173 perion, f. of Axiocerses harpax 168 perobscura, Liptena (Liptena) 55 perparva, Zizula 261 perpulchra, Lepidochrysops 232 petalus, Thestor 183 petersi, Baliochila 43 petersi, Hypokopelates 104 petra, Thestor 183

petreia, Pentila 19	praeusta, subsp. of Liptena (Liptena)
petreoides, Pentila 19	flavicans 54
peuceda, Ornipholidotos 26	praestans, Liptena (Liptena) 55
peucetia, Ornipholidotos 26	praeterite, Lepidochrysops 232
phaeochiton, Pseuderesia 36	preussi, Hewitsonia 94
phanes, Spindasis 159	preussi, Pentila 19
pharaonis, Chilades 248	princeps, Anthene 193
Phasis 172	princeps, subsp. of Lepidochrysops cinere
phasma, f. of Castalius margaritaceus 212	230
phasma, Lepidochrysops 232	procera, Lepidochrysops 232
phidia, subsp. of Pentila abraxas 17	proclus, subsp. of Lepidochrysops plebej
phidias, Leptomyrina (Leptomyrina) 170	232
philbyi, Euchrysops 236	prodita, subsp. of Pentila umangiana 20
Philiolaus, subg. of Iolaus 136	promethus, subsp. of Iolaus (Tanuetheira
philippus, Hypolycaena 120	timon 129
phlaeas ethiopica, Lycaena 264	propinguus, Aphnaeus 153
phlaeas menelicki, Lycaena 264	protumnus, Thestor 183
phoa, Euchrysops 236	Pseudaletis 113
	pseudepaea, Mimacraea 31
phoenicis, Triclema 200	Pseuderesia 33
phosphor, Poecilmitis 178	33
Phlyaria 200	Pseudiolaus, subg. of Iolaus 126
Phyltaa 84	Pseudocapys 172
piaggiae, subsp. of Iolaus (Iolaphilus)	pseudoconjuncta, Epitola 90
ismenius 135	pseudoderitas, Diopetes 107
picata, Alaena 9	pseudofrater, Iolaus (Epamera) 147
picena, Pentila 19	pseudofusca, Eresina 63
picta, Pseuderesia 36	Pseudonacaduba 225
pictus, subsp. of Thestor montanus 183	Pseudoneaveia 81
pierus, Aloeides 176	pseudophlaeas, subsp. of Lycaena phlaea
pierus var. A, Aloeides 176	264
pierus var. B, Aloeides 176	pseudopollux, Iolaus (Epamera) 147
Pilodeudorix 104	pseudopunctatus, Oboronia 245
pilos, Spalgis 185	pseudorotha, Pentila 19
pinodes, Epitole 90	pseudosoyauxi, Athysanota 246
pinodoides, Epitola 90	pseudozeritis, Chloroselas 163
pirithous, Syntarucus 221	pterou, Lepidochrysops 232
pitho, Tarucus 216	publia, Zizina 259
pitmani, Anthene 193	puella, Teriomima 40
plagiata, Telipna 12	puellaris, Teriomima 40
plana, subsp. of Hypolycaena liara 120	pulcher, Anthene 194
plebeja, Lepidochrysops 232	pulcher, Syntarucus 221
pluricauda, Neurellipes 197	pulcherrima, Zeritis 164
plurilimbata, Thermoniphas 243	pulchra, Epitola 90
podorina, Chilades 248	pulchristriata, Chilades 248
poecilaon, Iolaus (Iolaphilus) 135	pulsius, Phasis 173
Poecilmitis 176	pulverulenta, Epitola 90
poggei, Uranothauma 203	pulverulenta, Mimacraea 31
pollux, Iolaus (Epamera) 147	punctata, f. of Aloeides damarensis 176
polydialecta, Lepidochrysops 232	punctata, f. of Larinopoda lagyra 45
posthumus, Epitola 90	punctatus, Oboronia 245
poultoni, Iolaus (Pseudiolaus) 127	puncticilia, Lepidochrysops 232
poultoni, subsp. of Mimacraea krausei 31	punicea, Axiocerses 168
Poultonia 78	purpurascens, Aslauga 69
Powellana oa	nurnurea subsp. of Mimeresia moreelsi 22

pusillus, Eicochrysops 239 pusio, Citrinophila 39 pygmoea, Zizula 261 pyramus, Poecilmitis 178 pyroeis, Poecilmitis 178 pyroptera, Anthene 194 pyrrhops, Euchrysops 236

quadratus, Tarucus 216 quadricaudata, Neurypexina 196 quadriocularis, Lycaena 239 quassi, Lepidochrysops 232 questiauxi, Aphnaeus 153

rabe, Leptomyrina (Leptomyrina) 170 rabefaner, Syntarucus 221 radiata, Anthene 194 radiata, subsp. of Epitola viridana 90 radiata, f. of Pentila nyassana 19 ramonza, subsp. of Hypolycaena philippus 120 ras, subsp. of Pentila nyassana 19 rattrayi, Aphnaeus 153 rava, Hypokopelates 104 rectifascia, Lachnocnemus 98 rectifascia, Liptena (Liptena) 55 reducta, Euchrysops 236 reducta, Falcuna 47 reducta, Phytala 85 regillus, Neurypexina 196 regularis, Citrinophila 39 reichenowi, Lepidochrysops 232 renidens, Virachola 110 resplendens, subsp. of Castalius hintza 211 reticulata, Alaena 9 reticulum, subsp. of Pseudonacaduba sichela reutlingeri, Lachnocnemus 98 rex, subsp. of Aphnaeus erikssoni 153 rezia, Phytala 85 rhodesendae, Lepidochrysops 232 rhodesiana, Anthene 194 rhodesiensis, subsp. of Virachola dinochares rhodosense, subsp. of Iolaus (Epamera) mimosae 147 richardi, Pseudaletis 114 rileyi, subsp. of Capys catharus 172 rileyi, Epitola 90 rileyi, Thestor 184 ringa, Lepidochrysops 233 robusta, Cupidesthes 188

rochei, Liptena (Liptena) 55 rogersi, Desmolycaena 166 rogersi, Eicochrysops 239 rogersi, subsp. of Hypolycaena buxtoni 120 rogersi, Pentila 19 roidesta, f. of Pentila tachyroides 20 rollei, Alaena 9 rosacea, Tarucus 216 rosimon, Castalius 212 rotha, Pentila 20 rothi, Telipna 12 rothioides, Telipna 12 rothschildi, Anthene 194 rougemonti, f. of Aloeides aranda 175 rougeoti, Eresina 63 rougeoti, Iridana 76 rougeoti, Liptena (Tetrarhanis) 57 rougeoti, Pseuderesia 36 ruandensis, subsp. of Harpendyreus meruanus rubrica, Mimeresia 33 rubricinctus, Anthene 194 rubrimaculata, Anthene 194 rubromacula, Liptena (Liptena) 55 rubropuncta, Azanus 252 rufilla, Telipna 12 rufomarginata, Anthene 194 rufoplagiata, Triclema 200 rumina, Phasis 173 ruspinoides, Telipna 12 russulus, Mimeresia 33 ruthica, Lepidochrysops 233 rutila, Virachola 110 rutilans, Virachola 110 rutilo, Pseuderesia 36 rutshurensis, subsp. of Thermoniphas plurilimbatus 243 ruwenzoricus, Anthene 194

sabulosa, f. of Apharitis nilus 156
sadducus, Anthene 194
sadeska, Diopetes 107
saga, Durbaniopsis 29
salmoneus, Phasis 173
sanctithomae, Chilades 248
sanguigutta, Eicochrysops 239
sanguinea, Anthene 194
sanguinea, Telipna 12
sankuru, Micropentila 60
sankuru, Toxochitona 65
sapphirinus, Eicochrysops 239
sapphirinus, Iolaus (Epamera) 148
sappirus, Iolaus (Epamera) 148

sardonyx, Phasis 173	silenus, Myrina 112
sauberi, Liptena (Liptena) 55	silvius, Thestor 184
saundersi, Erisina 64	simekoa, ab. of Larinopoda lircaea 45
schmidtii, Mimacraea 31	similis, Citrinophila 39
schoutedeni, Anthene 194	similis, Hewitsonia 94
schoutedeni, Liptena (Tetrarhanis) 57	similis, Hypolycaena 120
schoutedeni, Liptenara 21	similis, Liptena (Liptena) 55
schubotzi, Hypolycaena 120	similis, Megalopalpus 97
schubotzi, Mimacraea 31	simplex, Aloeides 176
schultzei, subsp. of Hypokopelates angelita	simplex, subsp. of Hypokopelates aruma 10
103	simplex, Megalopalpus 97
schultzei, Iolaus (Iolaphilus) 135	simplex, Liptena (Tetrarhanis) 57
schultzei, Phytala 85	simplex, Pilodeudorix 106
scintilla, Lycaena 239	simplicia, Liptena (Liptena) 55
scintillans, subsp. of Hypolycaena lebona	singularis, Baliochila 43
120	sitalces, Azanus 252
scintillans, Iolaus (Epamera) 148	siwani, Cupidopsis 240
scintillula, Anthene 194	sjoestedti, subsp. of Euliphyra mirifica 71
sciophilus, Iolaus (Epamera) 148	skoptolos, Mimacraea 31
Scoptes 171	skotios, Lepidochrysops 233
scotti, Spindasis 159	smithi, subsp. of Anthene princeps 193
seamani, Hypolycaena 120	soalalicus, subsp. of Azanus jesous 252
sebagadis, f. of Eicochrysops messapus 239	sobrina, subsp. of Anthene crawshayi 191
sebasta, Hypolycaena 120	sobrina, subsp. of Leptomyrina (Gonatomy
semibrunnea, Epitola 80	rina) lara 170
semilimbata, Liptena (Liptena) 55	socotranus, Syntarucus 221
semirufa, Mimeresia 33	solwezi, Lepidochrysops 233
semirufa, subsp. of Telipna bimacula 12	somalina, subsp. of Anthene pitmani 193
semliki, Falcuna 47	somalina, Jolaus (Stugeta) 126
septentrionalis, Iolaus, (Epamera) 148	somalina, Foliaus (Stugeta) 120
septistrigata, Liptena (Liptena) 55	somereni, f. of Mimacraea marshalli doherty
serena, Citrinophila 39	
sesse, subsp. of Virachola lorisona 110	31 sorhageni, Zeritis 164
severini, Euchrysops 236	souanke, Liptena (Tetrarhanis) 57
seydeli, subsp. of Aphnaeus affinis 150	souanke, Micropentila 60
sharpei, Myrina 112	soyauxii, Larinapoda 45
sharpiae, subsp. of Harpendyreus aequa-	Spalgis 184
torialis 224	Spindasis 157
sheffieldi, Telipna 12	spindasoides, subsp. of Lipaphnaeus aderna
Sheffieldia 13	161
sheppardi, Anthene 194	splendens, ab. of Hypolycaena lebona 120
sheppardi, subsp. of Deloneura millari 77	splendens, Poecilmitis 178
shima, subsp. of Lycaena phlaeas 264	splendens, f. of Uranothauma antinorii 203
sibella, Iolaus (Epamera) 148	spolia, Pseudaletis 114
sichela, Anthene 194	spuma, f. of Larinopoda lircaea 45
sichela, Pseudonacaduba, 227	spurcus, subsp. of Hypolycaena buxtoni 120
sidus, Iolaus (Epamera) 148	s-signata, Spalgis 185
sigiensis, f. of Pentila tropicalis 20	stactalla, subsp. of Phylaria cyara 201
sigillata, Azanus 252	staphyla, Eresinopsides 62
silanus, Iolaus (Epamera) 148	staudingeri, Epitola 90
silarus, subsp. of Iolaus (Argiolaus) silas 131	staudingeri, Pertola 90 staudingeri, Neurellipes 197
silas, Iolaus (Argiolaus) 131	staudingeri, Neurempes 197 staudingeri, Oboronia 245
silas ab. lasius, Iolaus (Argiolaus) 131	stellata, Actizera 263
silenus subsp. of Iolaus (Epamera) silanus 148	stempfferi subsp. of Anthene butleri 100

stempfferi, Epitola 90 stempfferi, Liptena (Tetrarhanis) 37 stempfferi, Thermoniphas 243 Stempfferia 83 stenogrammica, Iolaus (Epamera) 148 stormsi, Lepidochrysops 233 stratola, Pseudonacaduba 227 strigatus, subsp. of Chilades eleusis 248 strutti, Thestor 184 Stugeta, subg. of Iolaus 124 stygia, Baliochila 43 styx, subsp. of Axiocerses harpax 168 subalba, Epitola 90 subangulata, subsp. of Pseudaletis clymenus subargentea, Epitola 90 subaureus, Apharitis 156 subcoerulea, Epitola 90 subdita, Euchrysops 236 subfulvida, Aslauga 69 subfusca, Deloneura 77 subfuscata, Pentila 20 subgriseata, Epitola 90 subhyalina, Telipna 12 subinfuscata, subsp. of Iolaus (Stugeta) bowkeri 126 sublustris, Epitola 90 submacula, Liptena (Liptena) 55 subnotens, Triclema 200 subochracea, subsp. of Pentila abraxas 17 subornata, Myrina 113 subpallida, Euchrysops 236 subplagiata, Micropentila 60 subpunctata, Liptena (Liptena) 55 subpunctata, Teriomima 40 subrubra, Alaena 9 subsuffusa, Liptena (Liptena) 55 subundularis, Liptena (Liptena) 55 subvariegata, Lepidochrysops 233 subvariegata, Liptena (Liptena) 55 sudanica, subsp. of Iolaus (Epamera) umbrosa 148 sudanica, Lachnocnemus 98 sudanica, Leptomyrina (Leptomyrina) suetonus, Aloeides 176 suffusa, subsp. of Falcuna hollandi 47 suk, subsp. of Lepidochrysops pterou 232 suk, Virachola 110 sukidion, subg. of Iolaus 149 sulpitia, Telipna 12 suquala, Anthene 194 swanepoeli, Lepidochrysops 233 swanepoeli, Poecilmitis 178

swinburnei, subsp. of Lepidochrysops glauca 230
swynnertoni, subsp. of Pentila tropicalis 20
sybaris, Tarucus 216
syllidus, Anthene 194
sylpha, Ornipholidotos 26
sylphida, Ornipholidotos 26
sylvanus, Anthene 194
sylvius, Lepidochrysops 233
symmacha, Hypolycaena 120
symplocus, Liptena (Tetrarhanis) 57
synchrematiza, Lepidochrysops 233
synesia, Falcuna 47
Syntarucus 219
Syrmoptera 116

tachyroides, Pentila 20 taikosama, Aloeides 176 tajoraca, Iolaus (Epamera) 148 talboti, Anthene 194 tamaniba, Chloroselas 163 tanganyikensis, subsp. of Epitola urania 90 tantalus, Lepidochrysops 233 Tanuethiera, subg. of Iolaus 128 taposana, Chloroselas 163 Tarucus 212 tavetensis, Spindasis 159 teita, subsp. of Anthene hobleyi 191 telesippe, Pentila 20 telicanus, Syntarucus 221 Telipna 9 telloides, Mimacraea 31 tempe, Thestor 184 tenera, Cirtinophila 39 tenuimarginata, f. of Phlyaria cyara 201 tenuivittata, Hypokopelates 104 tera, Larinopoda 45 Teratoneura 73 terias, Citrinophila 39 Teriomima 39 teroana, subsp. of Liptena (Liptena) o-rubrum 55 teroensis, Ornipholidotos 26 terpsichore, subsp. of Cooksonia trimeni 14 terranus, Syntarucus 221 tespis, Cacyeus 206 tessmanni, subsp. of Mimeresia moreelsi 33 Tetrarhanis, subg. of Liptena 57 thebana, Azanus 252 theodori, Eresina 64 theodota, Lepidochrysops 233 theophrastus, Tarucus 216 Thermoniphas 241 thero, Phasis 173

thespis, Tarucus 216 Thestor 182 thuraui, Iolaus (Iolaus) 139 thyra, Aloeides 176 thysbe, Poecilmitis 178 thyrsis, Cupidesthes 188 timon, Iolaus (Tanuetheira) 129 tintinga, Spalgis 185 tiressa, Euchrysops 236 tirza, Ornipholidotos 26 tisamenus, Triclema 200 tjoane, Axiocerses 168 togara, Thermoniphas 243 tongidensis, Anthene 194 toroensis, subsp. of Anthene lachares 192 toroensis, Eresina 64 toroensis, Iolaus (Epamera) 148 tororo, Iridana 76 torrida, Pentila 20 Toxochitona 64 transverstigma, Telipna 12 triangularis, Micropentila 60 Trichiolaus, subg. of Iolaus 127 Triclema 198 tricolor, Pseudaletis 114 tricolora, Liptena (Liptena) 55 trifasciata, Pseudaletis 114 trimeni, Cooksonia 14 trimeni, Iolaus (Iolaphilus) 135 trimena, Lepidochrysops 233 trimeni, subsp. of Poecilmitis thysbe 178 trimeni, Spindasis 159 tripunctata, subsp. of Liptena (Liptena) o-rubrum 55 tripunctata, (Aurivillius) Pentila 20 tripunctata (Druce), Pentila 20 trisignatus, subsp. of Eicochrysops mahallakoena 239 trochylus, Freyeria 249 tropicalis, Pentila 20 tsavao, subsp. of Alaena johanna 9 tsiphana, ab. of Virachola batikeli 109 tsomo, Harpendyreus 224 tullia, Liptena (Liptena) 55 tulliana, Liptena (Liptena) 55 tumentia, Epitola 90 Tumerepedes 80 turbata, Liptena (Liptena) 56 turbatus, Neurypexina 196 turkana, subsp. of Anthene contrastata 190 turneri, Poecilmitis 178 tytleri, subsp. of Chloroselas pseudozeritis 163

uelensis, subsp. of Pentila cloetensi 18 ugandae, subsp. of Anthene princeps 194 ugandae, subsp. of Aphnaeus chapini 153 ugandae, subsp. of Argyrocheila undifera 67 ugandae, subsp. of Epitolina catori 83 ugandae, subsp. of Hewitsonia similis 94 ugandae, Hypokopelates 104 ugandae, subsp. of Hypolycaena hatita 120 ugandae, subsp. of Iolaus (Epamera) alienus ugandae, subsp. of Liptena (Tetrarhanis) ilma 57 ugandae, Micropentila 61 ugandae, subsp. of Mimeresia drucei 33 ugandae, Ornipholidotos 26 ugandae, subsp. of Phytala elais 85 ugandae, Pseudaletis 114 ugandae, subsp. of Telipna arica 12 ugandana, subsp. of Axiocerses harpax 168 ukerewensis, Anthene 194 ula, Actis 101 ultramarina, Hypokopelates 104 umangiana, Pentila 20 umbra, Pentila 20 umbra, f. of Uranothauma falkensteini 203 umbratilis, subsp. of Epitola marginata 89 umbrosa, f. of Chloroselas pseudozeritis tytleri 163 umbrosa, Iolaus (Epamera) 148 umbrya, Lachnochemus 98 unda, subsp. of Actis mimeta 101 undifera, Argyrocheila 67 undina, Liptena (Liptena) 56 undularis, Liptena (Liptena) 56 ungemachi, Tarucus 216 unicolor, f. of Cupidesthes thyrsis 188 unicolor, subsp. of Toxochitona gerda 65 uniformis, Epitola 90 unigemmata, Euchrysops 236 unimaculata, Alaena 9 unipunctata, Citrinophila 39 unyoro, Iridana 76 unyoro, subsp. of Mimeresia russulus 33 urania, Epitola 90 uranochroa, subsp. of Cupidopsis iobates 240 Uranothauma 201 uranus, Poecilmitis 178 usamba, subsp. of Anthene hodsoni 191 usambara, subsp. of Iolaus (Iolaphilus) maritima 135 usemia, subsp. of Castalius cretosus 211

vansomereni, Chloroselas 163 vansomereni, Cnodontes 44 vansomereni, Iolaus (Iolaphilus) 135 vansomereni, subsp. of Lepidochrysops victoriae 233 vansomereni, Phytala 85 vansomereni, Toxochitona 65 vansomereni, Uranothauma 203 vansomereni, Virachola 110 vansoni, Cnodontes 44 vansoni, Lepidochrysops 233 vansoni, Thestor 184 vansoni, Virachola 110 variegata, Lepidochrysops 233 variegata, Pseuderesia 36 varipes, Larinopoda 45 varnieri, Iolaus (Hemiolaus) 124 venanigra, Telipna 12 vera, Lepidochrysops 233 versatilis, Anthene 194 versicolor, Epitola 90 vestalis, subsp. of Athysanota ornata 246 vexillarius, Iolaus (Iolaus) 139 victoriae, Lepidochrysops 233 victoriae, Micropentila 61 victoriae, Spindasis 159 vidua, Cupidesthes 188 vidua, subsp. of Epitola miranda 89 vidua, subsp. of Pseuderesia eleaza 35 villiersi, Telipna 12 vinalli, Epitola 90 vininga, Aslauga 69 violacea, Iolaus (Epamera) 148 violetta, Diopetes 107 violetta, Lepidochrysops 233 Virachola 107 virgata, Kopelates 102 virginea, Epitola 90 virginea, Powelli 95 virgo, Phylaria 201 viridana, Epitola 90 viridis, Hypokopelates 104 virilis, Cacyreus 206 virilis, ab. of Hewitsonia boisduvali 94 vittigera, ab. of Hypolycaena philippus ramonza 120 viviana, f. of Mimacraea krausei 31 voltae, Cupidesthes 188 vosseleri, subsp. of Virachola diocles 110 vulcanica, ab. of Citrinophila erastus 39 vulcanica, subsp. of Harpendyreus aequatorialis 224

waggae, Spindasis 159
wallengreni, Aloeides 176
wardii, Virachola 110
wau, f. of Lepidochrysops negus 231
webbianus, Cyclyrius 219
weberi, Powellana 95
williamsi, subsp. of Aphnaeus flavescens 153
williamsi, Uranothauma 203
wilsoni, Cupidesthes 188
wollastoni, subsp. of Harpendyreus
marungensis 224
woodi, Baliochila 43
wykehami, Lepidochrysops 233

xantha, Liptena (Liptena) 56 xanthis, Liptena (Liptena) 56 xanthopoecilus, Anthene 194 xanthostola, Liptena (Liptena) 56

yalae, subsp. of Iolaus (Epamera) bansana 145 yuanda, Pentila 20 yokoana, Iolaus (Epamera) 148 ysobelae, Cupidesthes 188 yukadumae, Liptena (Liptena) 56

zanzibarensis, Paraphnaeus 155 zaraces, Thestor 184 zebra, Batelusia 80 zebra, Pseudaletis 114 zela, Pilodeudorix 106 zelica, Epitola 90 zeloides, subsp. of Pilodeudorix zela 106 zelomina, Pilodeudorix 106 zelza, Epitola 90 Zeltus 117 zena, Azanus 252 zenkeri, Anthene 195 zerita, f. of Mimeresia libentina 33 Zeritis 163 zeuxo, Chrysoritis 179 zilka, subsp. of Aloeides aranda 175 Zizeeria 256 Zizina 258 Zizula 260 zonarius, subsp. of Chrysoritis zeuxo 179 zoraida, Mimeresia 33 zorites, Crudaria 180 zuluana, Teriomima 40 zymna, Megalopalpus 97

